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(54) ENGINEERED NUCLEIC ACIDS AND METHODS OF USE THEREOF

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(56) References Cited

U.S. PATENT DOCUMENTS

2,008,526 A	7/1935	Wrappler et al.
3,552,394 A	1/1971	Horn et al.
3,737,524 A	6/1973	Ebel et al.
3,766,907 A	10/1973	Muenzer
3,906,092 A	9/1975	Hilleman et al.
4,373,071 A	2/1983	Itakura
4,399,216 A	8/1983	Axel et al.
4,401,796 A	8/1983	Itakura
4,411,657 A	10/1983	Galindo
4,415,732 A	11/1983	Caruthers et al.
4,458,066 A	7/1984	Caruthers et al.
4,474,569 A	10/1984	Newkirk

4,500,707 A	2/1985	Caruthers et al.
4,579,849 A	4/1986	MacCoss et al.
4,588,585 A	5/1986	Mark et al.
4,668,777 A	5/1987	Caruthers et al.
4,737,462 A	4/1988	Mark et al.
4,816,567 A	3/1989	Cabilly et al.
4,879,111 A	11/1989	Chong
4,957,735 A	9/1990	Huang
4,959,314 A	9/1990	Mark et al.
4,973,679 A	11/1990	Caruthers et al.
5,012,818 A	5/1991	Joishy
5,017,691 A	5/1991	Lee et al.
5,021,335 A	6/1991	Tecott et al.
5,036,006 A	7/1991	Sanford et al.
5,047,524 A	9/1991	Andrus et al.
5,116,943 A	5/1992	Koths et al.
5,130,238 A	7/1992	Malek et al.
5,132,418 A	7/1992	Caruthers et al.
5,153,319 A	10/1992	Caruthers et al.
5,168,038 A	12/1992	Tecott et al.
5,169,766 A	12/1992	Schuster et al.
5,194,370 A	3/1993	Berninger et al.
5,199,441 A	4/1993	Hogle
5,240,855 A	8/1993	Tomes
5,240,885 A	8/1993	Aitken et al.
5,262,530 A	11/1993	Andrus et al.
5,273,525 A	12/1993	Hofman
5,298,422 A	3/1994	Schwartz et al.
5,332,671 A	7/1994	Ferrara et al.
5,399,491 A	3/1995	Kacian et al.
5,409,818 A	4/1995	Davey et al.
5,105,510 71		•
	(Con	tinued)

(Continued)

FOREIGN PATENT DOCUMENTS

CA	2376634	12/2000
CA	2473135	6/2003
	(Co	ntinued)

OTHER PUBLICATIONS

Kariko, K., et al. Molecular Therapy, vol. 16, No. 11, pp. 1833-1840, Nov. 2008.*

Egeter, O. et al., Eradication of disseminated lymphomas with CpG-DNA activated T helper type 1 cells from nontransgenic mice. Cancer Res. Mar. 15, 2000;60(6):1515-20.

El Ouahabi, A., et al., Double long-chain amidine liposome-mediated self replicating RNA transfection. FEBS Letters. Feb. 1996; 380(1-2): 108-112.

Elango, N., et al., Optimized transfection of mRNA transcribed from a d(A/T)100 tail-containing vector. Biochem Biophys Res Commun. 2005; 330: 958-966.

El Bashir, S.M. et al. Duplexes of 21 nucleotide RNAs mediate RNA interference in cultured mammalian cells. Nature. May 24, 2001;411(6836):494-8.

(Continued)

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(57) ABSTRACT

Provided are compositions and methods for delivering biological moieties such as modified nucleic acids into cells to kill or reduce the growth of microorganisms. Such compositions and methods include the use of modified messenger RNAs, and are useful to treat or prevent microbial infection, or to improve a subject's heath or wellbeing.

17 Claims, No Drawings

(56)		Referen	ces Cited	6,234,990 6,235,883			Rowe et al. Jakobovits et al.
	211	PATENT	DOCUMENTS	6,239,116			Krieg et al.
	0.5.	IMILIVI	DOCOMENTS	6,251,665			Cezayirli et al.
	5,426,180 A	6/1995	Kool	6,255,076			Widner et al.
	5,437,990 A		Burg et al.	6,258,558			Szostak et al.
	5,457,041 A		Ginaven et al.	6,261,584 6,265,387			Peery et al. Wolff et al.
	5,466,586 A 5,484,401 A		Davey et al. Rodriguez et al.	6,265,389		7/2001	
	5,514,545 A		Eberwine	6,267,987		7/2001	Park et al.
	5,527,288 A		Gross et al.	6,291,170			Van Gelder et al.
	5,545,522 A		Van Gelder et al.	6,300,484		10/2001	
	5,554,517 A		Davey et al.	6,303,378 6,303,573			Bridenbaugh et al. Ruoslahti et al.
	5,580,859 A 5,588,960 A		Felgner et al. Edwards et al.	6,322,967		11/2001	
	5,589,466 A		Felgner et al.	6,326,174			Joyce et al.
	5,663,153 A		Hutherson et al.	6,334,856			Allen et al.
	5,665,545 A		Malek et al.	6,355,245 6,368,801		3/2002 4/2002	Evans et al.
	5,672,491 A 5,674,267 A		Khosla et al. Mir et al.	6,376,248			Hawley-Nelson et al.
	5,677,124 A		DuBois et al.	6,395,253		5/2002	Levy et al.
	5,679,512 A		Laney et al.	6,399,061			Anderson et al.
	5,693,622 A		Wolff et al.	6,406,705			Davis et al.
	5,693,761 A		Queen et al.	6,410,276 6,413,942			Burg et al. Felgner et al.
	5,697,901 A 5,700,642 A		Ericksson Monforte et al.	6,433,155		8/2002	Umansky et al.
	5,702,384 A		Umeyama et al.	6,440,096	B1	8/2002	Lastovich et al.
	5,703,055 A		Felgner et al.	6,455,043			Grillo-Lopez
	5,712,127 A		Malek et al.	6,491,657			Rowe et al. Hone et al.
	5,716,785 A		Van Gelder et al.	6,500,419 6,500,919			Adema et al.
	5,736,137 A 5,756,264 A		Anderson et al. Schwartz et al.	6,514,498			Antonsson et al.
	5,759,179 A		Balbierz	6,514,948	B1	2/2003	Raz et al.
:	5,766,903 A	6/1998	Sarnow et al.	6,517,869			Park et al.
	5,773,244 A		Ares, Jr. et al.	6,520,949 6,525,183			St. Germain Vinayak et al.
	5,776,456 A		Anderson et al.	6,527,216			Eagelman et al.
	5,789,554 A 5,807,707 A		Leung et al. Andrews et al.	6,528,262			Gilad et al.
	5,824,307 A	10/1998		6,534,312			Shiver et al.
:	5,824,497 A		Andrews et al.	6,552,006			Raz et al.
	5,840,299 A		Bendig et al.	6,555,525 6,565,572		4/2003	Chappuis
	5,843,439 A 5,848,996 A	12/1998	Anderson et al.	6,572,857			Casimiro et al.
	5,849,546 A		Sousa et al.	6,586,524		7/2003	
	5,851,829 A		Marasco et al.	6,589,940			Raz et al.
	5,861,501 A		Benseler et al.	6,610,044 6,610,661			Mathiesen Carson et al.
	5,869,230 A		Sukhatme Scaringe et al.	6,613,026			Palasis et al.
	5,889,136 A 5,891,636 A		Van Gelder et al.	6,617,106		9/2003	
	5,914,269 A		Bennett et al.	6,623,457			Rosenberg
	5,955,310 A		Widner et al.	6,652,886			Ahn et al.
	5,958,688 A		Eberwine et al.	6,653,468 6,664,066		12/2003	Guzaev et al.
	5,962,271 A 5,962,272 A		Chenchik et al. Chenchik et al.	6,670,178			Selden et al.
	5,965,720 A	10/1999	Gryaznov et al.	6,676,938	B1	1/2004	Teti et al.
	5,965,726 A	10/1999	Pavlakis et al.	6,696,038			Mahato et al.
	5,980,887 A		Isner et al.	6,743,211 6,743,823			Prausnitz et al. Summar et al.
	5,989,911 A 5,994,511 A		Fournier et al. Lowman et al.	6,777,187			Makarov et al.
	6,004,573 A		Rathi et al.	6,808,888			Zhang et al.
	6,019,747 A		McPhee	6,818,421			Kossmann et al.
	6,022,715 A		Merenkova et al.	6,835,393			Hoffman et al.
	6,057,494 A		Koops et al.	6,835,827 6,890,319			Vinayak et al. Crocker
	6,063,603 A 6,074,642 A		Davey et al. Wang et al.	6,896,885		5/2005	
	6,090,382 A		Salfeld et al.	6,900,302	B2		Teti et al.
(6,090,591 A		Burg et al.	6,902,734			Giles-Komar et al.
	6,096,503 A		Sutcliffe et al.	6,924,365 6,949,245			Miller et al. Sliwkowski
	6,100,024 A		Hudson et al.	6,960,193			Rosenberg
	6,124,091 A 6,132,419 A		Petryshyn Hofmann	6,962,694			Soegaard et al.
	6,147,055 A		Hobart et al.	7,001,890			Wagner et al.
(6,177,274 B1	1/2001	Park et al.	7,052,891			Leung et al.
	6,187,287 B1		Leung et al.	7,074,596			Darzynkiewicz et al.
	6,190,315 B1		Kost et al.	7,125,554			Forsberg et al.
	6,210,931 B1		Feldstein et al.	7,135,010 7,195,761			Buckman et al. Holtzman et al.
	6,214,804 B1 6,217,912 B1		Felgner et al. Park et al.	7,193,761			Schleyer et al.
	6,228,640 B1		Cezayirli et al.	7,202,226			Murray et al.
	,,		,	, -,			,

(56)		Referen	ces Cited	8,242,258 8,246,958			Dellinger et al. Bendig et al.
	U.S.	PATENT	DOCUMENTS	8,278,036	B2	10/2012	Kariko et al.
7 200 4	70 D2	4/2007	C	8,304,183 8,304,532			Sooknanan Adamo et al.
7,208,4° 7,226,4°			Carson et al. Prausnitz et al.	8,309,706			Dellinger et al.
7,226,59			Antonsson et al.	8,329,172			Grillo-Lopez et al.
7,268,12			Horton et al.	8,329,182 8,329,887			Peters et al. Dahl et al.
7,276,43 7,316,93			Agrawal et al. Draghia-Akli et al.	8,333,799			Bales, Jr. et al.
7,320,90			Kempf et al.	8,344,153	B2		Cottrell et al.
7,329,74		2/2008		8,349,321 8,367,328			Burke et al. Asada et al.
7,335,4° 7,348,00			Guillerez et al. Peters et al.	8,367,631		2/2013	
7,354,7			Kamme et al.	8,383,340			Ketterer et al.
7,371,40			Panzner et al.	8,394,763 8,399,007			Forte et al. Taft et al.
7,374,7° 7,374,9°			Hoffman et al. Oh et al.	8,404,222		3/2013	
7,378,20		5/2008	Sobek et al.	8,404,799			Podobinski et al.
7,384,73			Kitabayashi et al.	8,414,927 8,415,325			Richard Kiick et al.
7,404,93 7,422,73			Peters et al. Anderson et al.	8,420,123			Troiano et al.
7,476,50			Schleyer et al.	8,420,605			Ulijn et al.
7,476,70			Moody et al.	8,431,160 8,435,504			O'Hagan et al. Kozlowski et al.
7,479,54 7,498,4		3/2009	Tsuchiya et al.	8,440,231			Smyth et al.
7,501,4			Zhang et al.	8,440,614		5/2013	
7,521,0			Pastan et al.	8,444,992 8,449,884			Borkowski et al. Rivera et al.
7,547,6′ 7,550,20			Kempf et al. Getts et al.	8,449,916			Bellaire et al.
7,575,5			Sweeney	8,450,298			Mahon et al.
7,579,3			Divita et al.	8,454,946 8,454,948			Shen et al. Pearlman et al.
7,615,22 7,629,3		11/2009	Forsberg et al. Tobinick	8,460,696			Slobodkin et al.
7,641,9			Goldenberg et al.	8,460,709			Ausborn et al.
7,667,03			Alvarado	8,461,132 8,466,122			Cohen et al. Heyes et al.
7,682,6 7,699,8			White et al. Frankel et al.	8,470,560			Bergmann-Leitner et al.
7,708,99			Benyunes	8,470,771			Gao et al.
7,709,4		5/2010		8,476,234 8,496,945			Fima et al. Schlesinger et al.
7,718,42 7,737,10			Reinke et al. Hoffman et al.	8,506,928			Ferrara et al.
7,745,39			Mintz et al.	8,506,966			Podda et al.
7,763,2			Hedlund et al.	8,512,964 8,518,871			Tontonoz et al. Hsu et al.
7,776,52 7,794,7			Garcia et al. Bardroff et al.	8,519,110	B2		Kowalska et al.
7,799,90	00 B2		Adams et al.	8,529,538			Pang et al.
7,820,10			Curd et al.	8,529,939 8,530,429			Masters et al. Robbins et al.
7,820,62 7,829,09			Hart et al. Lobb et al.	8,530,625	B2		Kaplan et al.
7,846,89	95 B2	12/2010	Eckert et al.	8,535,655			O'Shea et al.
7,862,82 7,884,13			Peters et al. DeGroot et al.	8,535,701 8,535,702			Peery et al. Richard et al.
7,906,49		3/2011		8,545,843	B2		Curd et al.
7,943,10	58 B2	5/2011	Schlesinger et al.	8,557,231			Langer et al.
7,943,53 7,964,5			Divita et al. Fewell et al.	8,557,244 8,562,992			White et al. Adams et al.
7,999,0			Dellinger et al.	8,563,041	B2	10/2013	Grayson et al.
8,003,1		8/2011	Hoffman et al.	8,568,784 8,569,256			Lillard et al. Heyes et al.
8,008,44 8,039,2	49 B2 14 B2		Korman et al. Dahl et al.	8,580,297			Essler et al.
8,048,99			Yamanaka et al.	8,603,499	B2	12/2013	Zale et al.
8,057,82			Slobodkin et al.	8,603,500 8,603,501			Zale et al. Zale et al.
8,058,06 8,101,33			Yaworski et al. Cload et al.	8,603,534			Zale et al.
8,105,59			Goldenberg et al.	8,603,535	B2	12/2013	Troiano et al.
8,108,3		1/2012	Kraft et al.	8,609,142 8,609,822			Troiano et al. Elson et al.
8,137,9 8,153,70			Dahl et al. Kunz et al.	8,613,951			Zale et al.
8,158,36			Heise et al.	8,613,954	B2	12/2013	Zale et al.
8,158,60	01 B2	4/2012	Chen et al.	8,617,608			Zale et al.
8,178,60 8,183,3			Weiner et al. Fay et al.	8,618,240 8,623,367			Podobinski et al. Momm et al.
8,183,3			Ayyavoo et al.	8,628,801			Garreta et al.
8,202,9	83 B2	6/2012	Dellinger et al.	8,636,696			Ross et al.
8,217,0			Hoerr et al.	8,636,994 8,637,028			Bossard et al. Alexis et al.
8,226,93 8,242,03			Lobb et al. Divita et al.	8,637,028			Troiano et al.
8,242,0			Adelfinskaya et al.	8,642,076			Manoharan et al.

(56)	Referen	ices Cited	2004/0236268 A1		Mitragotri et al.
211	PATENT	DOCUMENTS	2004/0259081 A1 2005/0032730 A1		Watzele et al. Von Der Mulbe et al.
0.5	. I AILLINI	DOCUMENTS	2005/0037494 A1		Hecker et al.
8,652,487 B2	2/2014	Maldonado	2005/0054026 A1		Atsushi et al.
8,652,528 B2		Troiano et al.	2005/0059624 A1 2005/0064596 A1		Hoerr et al. Riemen et al.
8,658,211 B2		Rozema et al.	2005/0004390 A1 2005/0089913 A1		Williams
8,658,733 B2 8,663,599 B1		Jorgedal et al. Sung et al.	2005/0112141 A1		Terman
8,663,692 B1		Muller et al.	2005/0130201 A1		Deras et al.
8,663,700 B2		Troiano et al.	2005/0137155 A1 2005/0147618 A1	6/2005	McSwiggen et al. Rivera et al.
8,668,926 B1 8,685,368 B2		Mousa et al. Reineke	2005/0147018 A1 2005/0153333 A1		Sooknanan
8,685,458 B2		Miller et al.	2005/0181016 A1	8/2005	Freyman et al.
8,691,223 B2		Van Den Brink et al.	2005/0232919 A1		Grasso et al.
8,691,750 B2		Constein et al.	2005/0250723 A1 2006/0008910 A1		Hoerr et al. MacLachlan et al.
8,691,785 B2 8,691,963 B2		Teng et al. Brahmbhatt et al.	2006/0008910 A1 2006/0018971 A1		Scott et al.
8,696,637 B2	4/2014		2006/0032372 A1		Dauber et al.
8,697,098 B2		Perumal et al.	2006/0035226 A1		Scheinert et al.
8,703,204 B2		Bloom et al.	2006/0057111 A1 2006/0083780 A1		Hedlund et al. Heyes et al.
8,709,483 B2 8,715,677 B2		Farokhzad et al. Bartlett et al.	2006/0063780 A1 2006/0160743 A1		Zhang et al.
8,715,689 B2		Kinney et al.	2006/0172003 A1		Meers et al.
8,715,694 B2		Apt et al.	2006/0172966 A1		Lipford et al.
8,715,736 B2		Sachdeva et al.	2006/0188490 A1 2006/0204566 A1	8/2006 9/2006	Hoerr et al. Smyth-Templeton et al.
8,715,741 B2 8,722,341 B2		Maitra et al. Fouchier et al.	2006/0241076 A1		Uhlmann et al.
8,728,491 B2		Sesardic et al.	2006/0247195 A1	11/2006	Ray
8,728,527 B2	5/2014	Singh et al.	2006/0265771 A1		Lewis et al.
8,728,772 B2		Suzuki et al.	2006/0275747 A1 2007/0037147 A1		Hardy et al. Garcia et al.
8,734,832 B2 8,734,846 B2		O'hagan et al. Ali et al.	2007/0037147 A1 2007/0037148 A1		Fong et al.
8,734,853 B2		Sood et al.	2007/0048741 A1	3/2007	
8,735,566 B2		Brahmbhatt et al.	2007/0054278 A1		Cargill
8,735,570 B2		Miller et al.	2007/0072175 A1 2007/0087437 A1	3/2007 4/2007	Cooper et al.
2001/0001066 A1		Cezavirli et al.	2007/0087437 A1 2007/0105124 A1	5/2007	
2001/0005506 A1 2001/0014753 A1		Cezayirli et al. Soloveichik et al.	2007/0117112 A1		Diener et al.
2002/0001842 A1		Chapman et al.	2007/0122882 A1	5/2007	2
2002/0064517 A1		Cederholm-Williams	2007/0141030 A1	6/2007 6/2007	Yu et al.
2002/0111471 A1		Lo et al.	2007/0143878 A1 2007/0178103 A1		Bhat et al. Fey et al.
2002/0123099 A1 2002/0123723 A1		Weiner et al. Sorenson et al.	2007/0213287 A1		Fewell et al.
2002/0127592 A1		Ichihara et al.	2007/0224635 A1	9/2007	
2002/0130430 A1		Castor et al.	2007/0252295 A1 2007/0265220 A1	11/2007 11/2007	Panzner et al. Rossi et al.
2002/0132788 A1 2002/0143204 A1		Lewis et al. Evain et al.	2007/0280929 A1	12/2007	
2003/0026841 A1		Trubetskoy et al.	2008/0008711 A1	1/2008	Schleyer et al.
2003/0032615 A1		Felgner et al.	2008/0020431 A1	1/2008	
2003/0050468 A1		Shiver et al.	2008/0025944 A1 2008/0075698 A1	1/2008 3/2008	Hoerr et al. Sawada et al.
2003/0073619 A1 2003/0077604 A1		Mahato et al. Sun et al.	2008/0075098 A1 2008/0076174 A1		Selden
2003/0077004 A1 2003/0082768 A1		Baskerville et al.	2008/0119645 A1		Griffey et al.
2003/0083272 A1		Wiederholt et al.	2008/0166414 A1		Hanes et al.
2003/0092653 A1		Kisich et al 514/44	2008/0166793 A1 2008/0171711 A1		Beer et al. Hoerr et al.
2003/0138419 A1 2003/0143743 A1		Radic et al. Schuler et al.	2008/02/20471 A1		Davis et al.
2003/0153735 A1		Breece et al.	2008/0260706 A1		Rabinovich et al.
2003/0158133 A1		Movsesian	2008/0261905 A1		Herdewijn et al.
2003/0170273 A1		O'Hagan et al.	2008/0267873 A1 2008/0274463 A1		Hoerr et al. Chen et al.
2003/0171253 A1 2003/0186237 A1		Ma et al. Ginsberg et al.	2008/0275468 A1		Chuang et al.
2003/0191303 A1		Vinayak et al.	2008/0286813 A1	11/2008	
2003/0192068 A1		Deboer et al.	2008/0293143 A1 2009/0042829 A1	11/2008 2/2009	Lin et al. Matar et al.
2003/0225016 A1		Fearon et al. Ratti et al.	2009/0042829 A1 2009/0048167 A1	2/2009	Hillman
2004/0005667 A1 2004/0018525 A1		Wirtz et al.	2009/0053775 A1	2/2009	Dahl et al.
2004/0106567 A1		Hagstrom et al.	2009/0093433 A1	4/2009	Woolf et al.
2004/0110191 A1		Winkler et al.	2009/0144839 A1	6/2009	Inana et al.
2004/0122216 A1		Nielsen et al. Mahato et al.	2009/0169550 A1 2009/0170090 A1	7/2009 7/2009	Dummer Ignatov et al.
2004/0142474 A1 2004/0147027 A1		Troy et al.	2009/01/0090 A1 2009/0208418 A1	8/2009	Kohler et al.
2004/0167090 A1		Monahan et al.	2009/0208500 A1	8/2009	Joly et al.
2004/0171041 A1	9/2004	Dahl et al.	2009/0226470 A1	9/2009	Mauro et al.
2004/0171980 A1		Mitragotri et al.	2009/0227660 A1	9/2009	Oh et al.
2004/0197802 A1 2004/0202658 A1		Dahl et al. Benyunes	2009/0264511 A1 2009/0281298 A1	10/2009 11/2009	de Fougerolles et al. Manoharan et al.
2004/0202038 A1 2004/0209274 A2	10/2004	•	2009/0281298 A1 2009/0286852 A1	11/2009	
200 0205217 112	10,2004			11.2007	

(56)	Referer	nces Cited	2012/0128699			Kandimalla et al.
211	PATENT	DOCUMENTS	2012/0129759 2012/0156679			Liu et al. Dahl et al.
0.5	. IAILAT	DOCOMENTS	2012/0171290	A1		Coursaget et al.
2009/0324584 A1		Hoerr et al.	2012/0177724			Irvine et al.
2010/0003337 A1		Hanes et al.	2012/0178702 2012/0189700		7/2012 7/2012	Huang Aguilar et al.
2010/0004313 A1 2010/0004315 A1		Slobodkin et al. Slobodkin et al.	2012/0195917		8/2012	
2010/0009424 A1		Forde et al.	2012/0195936			Rudolph et al.
2010/0009865 A1		Herdewijn et al.	2012/0207840 2012/0213818			de los Pinos Hoerr et al.
2010/0015232 A1 2010/0021429 A1		Besenbacher et al. Brentzel, Jr. et al.	2012/0213818			Baumhof et al.
2010/0021429 A1 2010/0028943 A1		Thomas et al.	2012/0225070		9/2012	Smith et al.
2010/0047261 A1		Hoerr et al.	2012/0232133			Balazs et al.
2010/0086922 A1 2010/0120024 A1		Bryant et al.	2012/0237975 2012/0251618			Schrum et al. Schrum et al.
2010/0120024 A1 2010/0129877 A1		Cload et al. Sahin et al.	2012/0252117			Selden et al.
2010/0137407 A1		Abe et al.	2012/0258046			Mutske
2010/0178271 A1		Bridger et al.	2012/0276048 2012/0282247			Panzara et al. Schneewind et al.
2010/0189729 A1 2010/0196318 A1		Hoerr et al. Lieberburg	2012/0282249			Fox et al.
2010/0203076 A1		Fotin-Mleczek et al.	2012/0295832			Constien et al.
2010/0215580 A1		Hanes et al.	2012/0301955 2012/0321719			Thomas et al. McDonnell et al.
2010/0233141 A1 2010/0239608 A1		Polach et al. Von Der Mulbe et al.	2012/0321719			Rossi et al.
2010/0259008 AT 2010/0260817 AT		Slobodkin et al.	2012/0322865			Rossi et al.
2010/0261231 A1	10/2010	Kore et al.	2013/0012426			de los Pinos
2010/0266587 A1		McLachlan	2013/0012450 2013/0012566			de los Pinos De Los Pinos
2010/0273220 A1 2010/0285135 A1		Yanik et al. Wendorf et al.	2013/0017223			Hope et al.
2010/0291156 A1		Barner et al.	2013/0017265			Farokhzad et al.
2010/0293625 A1	11/2010		2013/0022538 2013/0029418		1/2013	
2010/0297750 A1 2010/0305196 A1		Natsume et al. Probst et al.	2013/0059360			Angel et al. Bossard et al.
2011/0002934 A1		Luqman et al.	2013/0064894		3/2013	Martin et al.
2011/0020352 A1		Garcia et al.	2013/0065942			Matar et al.
2011/0045022 A1	2/2011		2013/0071450 2013/0072670		3/2013	Copp-Howland Srivastava et al.
2011/0053829 A1 2011/0065103 A1		Baumhof et al. Sahin et al.	2013/0072709			McManus et al.
2011/0003103 A1 2011/0077287 A1		Von Der Mulbe et al.	2013/0084289			Curd et al.
2011/0086904 A1		Russell	2013/0090287 2013/0090372			Alessi et al. Budzik et al.
2011/0091473 A1 2011/0091879 A1		Golab et al. Hillebrand et al.	2013/0090372		4/2013	
2011/0091879 A1 2011/0097716 A1		Natt et al.	2013/0102545		4/2013	Gao et al.
2011/0112040 A1	5/2011	Liu et al.	2013/0108629			Dumont et al.
2011/0143397 A1		Kariko et al. Dahl et al.	2013/0111615 2013/0115192			Kariko et al. Ali et al.
2011/0143436 A1 2011/0165123 A1		Hartmann et al.	2013/0115196	A1		Hantash et al.
2011/0165159 A1		Grillo-Lopez et al.	2013/0115247		5/2013	de Los Pinos
2011/0172126 A1	7/2011		2013/0115272 2013/0115273			De Fougerolles et al. Yang et al.
2011/0182919 A1 2011/0200582 A1		Peters et al. Baryza et al.	2013/0115274			Knopov et al.
2011/0218231 A1		Fewell et al.	2013/0115293		5/2013	Sabnis et al.
2011/0244026 A1		Guild et al.	2013/0116408 2013/0121954			de Los Pinos Wakefield et al.
2011/0245756 A1 2011/0247090 A1	10/2011	Arora et al.	2013/0121934			Hoerr et al.
2011/0250225 A1		Fotin-Mleczek et al.	2013/0122104	A1	5/2013	Yaworski et al.
2011/0269950 A1		Von Der Mulbe et al.	2013/0123338			Heyes et al.
2011/0274697 A1		Thomas et al. Debart et al.	2013/0123351 2013/0129627			Dewitt Delehanty et al.
2011/0275793 A1 2011/0287006 A1		Friess et al.	2013/0129726			Lee et al.
2011/0294717 A1	12/2011	Ali et al.	2013/0129785			Manoharan et al.
2011/0300205 A1		Geall et al.	2013/0129794 2013/0129830			Kleiner et al. Chen et al.
2011/0311472 A1 2012/0009145 A1		Hoerr et al. Slobodkin et al.	2013/0130348			Gu et al.
2012/0009221 A1		Hoerr et al.	2013/0133483			Yang et al.
2012/0009649 A1		Dahl et al.	2013/0136746 2013/0137644			Schneewind Alluis et al.
2012/0015899 A1 2012/0021043 A1		Lomonossoff et al. Kramps et al.	2013/013/044			Kim et al.
2012/0021043 A1 2012/0027813 A1		Podda et al.	2013/0142818	A1	6/2013	Baumhof et al.
2012/0046346 A1	2/2012	Rossi et al.	2013/0142868			Hoekman et al.
2012/0053333 A1 2012/0060293 A1		Mauro et al. Stelter et al.	2013/0142876 2013/0149318			Howard et al. Reynolds et al.
2012/0060293 A1 2012/0065252 A1		Schrum et al.	2013/0149318		6/2013	
2012/0076836 A1		Hori et al.	2013/0149783			Yockman et al.
2012/0094906 A1		Guyon et al.	2013/0150295			Jaworowicz
2012/0095077 A1		Burrows et al. Schneewind et al.	2013/0150625		6/2013 6/2013	Budzik et al.
2012/0114686 A1 2012/0121718 A1		Lai et al.	2013/0150822 2013/0156721			Cheng et al.
						0*

(56)	References Cited	2013/0273039		Grillo-Lopez
211	PATENT DOCUMENTS	2013/0273047 2013/0273081		Rivera et al. Monaci et al.
0.5.	TATENT DOCUMENTS	2013/0273117		Podobinski et al.
2013/0156776 A1	6/2013 Chang et al.	2013/0274194		Dumont et al.
2013/0156845 A1	6/2013 Manoharan et al.	2013/0274504 2013/0274523		Colletti et al. Bawiec, III et al.
2013/0164219 A1 2013/0164343 A1	6/2013 Brinkmann et al. 6/2013 Hanes et al.	2013/02/4323		Karp et al.
2013/0164348 A1	6/2013 Palasis et al.	2013/0280339		
2013/0164400 A1	6/2013 Knopov et al.	2013/0281658		Rozema et al.
2013/0165499 A1	6/2013 Vaishnaw et al.	2013/0281671 2013/0287832		
2013/0165772 A1 2013/0171138 A1	6/2013 Traverso et al. 7/2013 Peters et al.	2013/0287832		Bhat et al.
2013/0171175 A1	7/2013 Pierce et al.	2013/0295183		Troiano et al.
2013/0171183 A1	7/2013 Schneewind	2013/0295191		Troiano et al.
2013/0171241 A1	7/2013 Geall	2013/0302432 2 2013/0302433 2		Zale et al. Troiano et al.
2013/0171646 A1 2013/0172406 A1	7/2013 Park et al. 7/2013 Zale et al.	2013/0315831		Shi et al.
2013/0172600 A1	7/2013 Chang et al.	2013/0317079		
2013/0177499 A1	7/2013 Brahmbhatt et al.	2013/0323179		
2013/0177523 A1	7/2013 Ghandehari et al.	2013/0323310 2 2013/0330401 2		Smyth et al. Payne et al.
2013/0177587 A1 2013/0177611 A1	7/2013 Robinson et al. 7/2013 Kaplan et al.	2013/0338210		Manoharan et al.
2013/0177633 A1	7/2013 Schutt et al.	2013/0344091		Berger et al.
2013/0177634 A1	7/2013 Schutt et al.	2013/0344158 2 2014/0005379 2		Zale et al.
2013/0177635 A1 2013/0177636 A1	7/2013 Schutt et al. 7/2013 Schutt et al.	2014/0003379		Cheng et al.
2013/0177637 A1	7/2013 Schutt et al.	2014/0017329		
2013/0177638 A1	7/2013 Schutt et al.	2014/0030351		
2013/0177639 A1	7/2013 Geall et al.	2014/0037573 2014/0037660		Eliasof et al. Fotin-Mleczek et al.
2013/0177640 A1 2013/0178541 A1	7/2013 Geall et al. 7/2013 Stanton et al.	2014/0037714		
2013/0183244 A1	7/2013 Hanes et al.	2014/0039032		
2013/0183355 A1	7/2013 Jain et al.	2014/0044772		
2013/0183372 A1	7/2013 Schutt et al. 7/2013 Schutt et al.	2014/0044791 2014/0045913		Basilion et al. Kuboyama et al.
2013/0183373 A1 2013/0183375 A1	7/2013 Schutt et al.	2014/0045950		Lacko et al.
2013/0183718 A1	7/2013 Rohayem et al.	2014/0050775		Slobodkin et al.
2013/0184207 A1	7/2013 Fares et al.	2014/0056867 2014/0056970		Lebowitz et al. Panzer et al.
2013/0184443 A1 2013/0184453 A1	7/2013 Bentley et al. 7/2013 Davis et al.	2014/0057109		Mechen et al.
2013/0189295 A1	7/2013 Arico et al.	2014/0065172		Echeverri et al.
2013/0189351 A1	7/2013 Geall	2014/0065204		Hayes et al.
2013/0189741 A1	7/2013 Meis et al.	2014/0065228 2014/0066363		Yarowoski et al. Bhunia et al.
2013/0195759 A1 2013/0195765 A1	8/2013 Mirkin et al. 8/2013 Gho et al.	2014/0073715		Fonnum et al.
2013/0195846 A1	8/2013 Friess et al.	2014/0073738		
2013/0195898 A1	8/2013 O'Hagan et al.	2014/0079774 2014/0079776		Brinker et al. Lippard et al.
2013/0195967 A1 2013/0195968 A1	8/2013 Guild et al. 8/2013 Geall et al.	2014/0080766		
2013/0195969 A1	8/2013 Geall et al.	2014/0081012		DeSimone et al.
2013/0197068 A1	8/2013 Kariko et al.	2014/0093575		
2013/0202595 A1	8/2013 Peirce et al.	2014/0093579 2014/0100178		Zale et al. Ansari et al.
2013/0202645 A1 2013/0202684 A1	8/2013 Barner et al. 8/2013 Geall et al.	2014/0106260		Cargnello et al.
2013/0203115 A1	8/2013 Schrum et al.	2014/0107227		Masters et al.
2013/0209454 A1	8/2013 Diskin et al.	2014/0107229 2014/0107349		Kormann et al. Bentley et al.
2013/0209456 A1 2013/0236419 A1	8/2013 Kano et al. 9/2013 Schneewind et al	2014/0107504		Guo et al.
2013/0236500 A1	9/2013 Zale et al.	2014/0113137		Podobinski et al.
2013/0236533 A1	9/2013 Von Andrian et al	. 2014/0121263		Fitzgerald et al. Manoharan et al.
2013/0236550 A1	9/2013 Ausborn et al.	2014/0121393 2014/0121587		Sallberg et al.
2013/0236556 A1 2013/0236968 A1	9/2013 Lai et al. 9/2013 Manoharan et al.	2014/0127227		Chang
2013/0243747 A1	9/2013 Fima et al.	2014/0127301		Alexis et al.
2013/0243827 A1	9/2013 Troiano et al.	2014/0128269		Hinz et al. Gore et al.
2013/0243848 A1 2013/0243867 A1	9/2013 Lobovkina et al. 9/2013 Mohapatra et al.	2014/0128329 2 2014/0134129 2		Thalhamer et al.
2013/0244972 A1	9/2013 Monapada et al. 9/2013 Ben-Shalom et al	2014/0134201	A 1 5/2014	Tureci et al.
2013/0245091 A1	9/2013 Rozema et al.	2014/0134230		Frank et al.
2013/0251679 A1	9/2013 Pearlman et al.	2014/0135380 2 2014/0135381 2		Hadwiger et al. Hadwiger et al.
2013/0251766 A1 2013/0251816 A1	9/2013 Zale et al. 9/2013 Zale et al.	2014/0133381 2		Kudirka et al.
2013/0251810 A1 2013/0251817 A1	9/2013 Zale et al.	2014/0141070		Geall et al.
2013/0259923 A1	10/2013 Bancel et al.	2014/0141089	A 1 5/2014	Liang
2013/0266553 A1	10/2013 Ballance et al.	2014/0141483		Bossard et al.
2013/0266611 A1 2013/0266617 A1	10/2013 Rabinovich et al. 10/2013 Mirosevich et al.	2014/0142165 2 2014/0142254 2		Grayson et al. Fonnum et al.
2013/0200017 A1 2013/0272994 A1	10/2013 Minosevich et al. 10/2013 Fu et al.	2014/0142234 1		Bancel et al.
		201.011.132		

(56)	Referen	ces Cited	WO	9819710 A	
	HC DATENIT	DOCUMENTS	WO WO	9834640 9847913 <i>A</i>	8/1998 A2 10/1998
	U.S. PATENT	DOCUMENTS	wo	9855495	12/1998
2014/0147454	A1 5/2014	Chakraborty et al.	WO	99/06073	2/1999
2014/0148502		Bancel et al.	WO WO	9914346 9920766	3/1999 4/1999
2014/0148503	A1 5/2014	Giangrande et al.	WO	9920766	4/1999
EC	DEICN DATE	NT DOCUMENTS	WO	9933982	7/1999
FC	KEION FAIE	NI DOCUMENTS	WO	9942618	8/1999
CA	2795695	10/2011	WO WO	9943835 9952503	9/1999 10/1999
EP	0194809	3/1986	WO	9954457	10/1999
EP EP	0204401 0366400	12/1986 5/1990	WO	0026226	5/2000
EP	0427073	5/1990	WO WO	0027340 0029561	5/2000 5/2000
EP	0427074	5/1991	WO	0029301	7/2000
EP	0735144 B1	3/1996	WO	0050586	8/2000
EP EP	0726319 0737750	8/1996 10/1996	WO	0075304	12/2000
EP	0771873 A3	7/1997	WO WO	0075356 0100650	12/2000 1/2001
EP	0839912	5/1998	wo	0104313	1/2001
EP EP	0969862 1026253	1/2000 8/2000	WO	01/14424 A	
EP	1020233 1083732 B1	3/2001	WO WO	0121810 0155306	3/2001 8/2001
EP	1404860	5/2002	WO	01/78779 A	
EP EP	1224943	7/2002 11/2003	WO	0192523	12/2001
EP EP	1361277 1393745	3/2004	WO	0193902	12/2001
EP	1083232	2/2005	WO WO	0208435 0224873	1/2002 3/2002
EP	1301614	11/2006	wo	0246477	6/2002
EP EP	1873180 A1 1905844 A2	1/2008 2/2008	WO	02064799	8/2002
EP	1964922 A1	3/2008	WO WO	02065093 02102839	8/2002 12/2002
EP	2072618	6/2009	WO	03002604	1/2003
EP EP	1056873 2191840	3/2010 6/2010	WO	03018798	3/2003
EP EP	2092064	9/2010	WO	03028656	4/2003
EP	2246422	11/2010	WO WO	03029401 03046578	4/2003 6/2003
EP	1619254	12/2010	WO	03050258	6/2003
EP EP	2292771 2377938	3/2011 10/2011	WO	03051923	6/2003
EP	2468290 A1	6/2012	WO WO	03059194 03059381	7/2003 7/2003
EP	2476430 B1	7/2012	wo	03066649	8/2003
EP EP	2484770 1907590	8/2012 9/2012	WO	03086280	10/2003
EP	2535419	12/2012	WO WO	03087815 03101401	10/2003 12/2003
EP	2188379	1/2013	wo	2004005544	1/2004
EP EP	2548960 2620161	1/2013 7/2013	WO	2004010106	1/2004
EP	2623121	7/2013	WO WO	2004035607 <i>A</i> 2004037972	A2 4/2004 5/2004
EP	2073848	8/2013	WO	2004057572	7/2004
EP EP	2623121 2695608 A2	8/2013 2/2014	WO	2004065561	8/2004
EP	2160464 B1	5/2014	WO WO	2004067728 2004085474	8/2004 10/2004
EP	2607379 B1	5/2014	WO	2004083474	10/2004
EP WO	2732825 A1 89/07947 A1	5/2014 3/1989	WO	2004092329	10/2004
WO	8906700	7/1989	WO	2005005622	1/2005
WO	8909622 A1	10/1989	WO WO	2005009346 2005017107 A	2/2005 \(\)2\(\)2\(\)2005
WO	9011092	10/1990	WO	2005/044859 A	
WO WO	9201813 A1 92/16553 A1	2/1992 10/1992	WO	2005040416	5/2005
WO	9309236	5/1993	WO WO	2005047536 2005/062967 <i>A</i>	5/2005 7/2005
WO	9314778	8/1993	WO	2005098433	10/2005
WO WO	9512665 9524485	5/1995 9/1995	WO	2005103081 A	
wo	9526204	10/1995	WO WO	2005117557 2005118857	12/2005 12/2005
WO	9529697 A1	11/1995	WO	2005118857 2006008154 A	
WO WO	95/35375 A1 9533835	12/1995 12/1995	WO	2006/013107 A	1 2/2006
WO	9617086	6/1996	WO	2006022712	3/2006
WO	9711085	3/1997	WO WO	2006044456 2006044503	4/2006 4/2006
WO	9712519	4/1997 8/1007	WO	2006044505	4/2006
WO WO	9730064 A1 9741210	8/1997 11/1997	WO	2006044682	4/2006
WO	9746680	12/1997	WO	2006046978 A	
WO	9748370	12/1997	WO	2006058088	6/2006
WO WO	9800547 9812207	1/1998 3/1998	WO WO	2006063249 2006065479	6/2006 6/2006
***	7012201	3/17/0	***	2000003777	0,2000

(56)	References Cited	WO	2010084371 A1	7/2010
	EODEICNI DATENIT DOCUMENTO	WO WO	2010088537 2010088927	8/2010 8/2010
	FOREIGN PATENT DOCUMENTS	wo	2010088927	9/2010
WO	2006065480 6/2006	WO	2010111290	9/2010
WO	2006071903 7/2006	WO WO	2010120266 2010129709	10/2010 11/2010
WO WO	2006095259 9/2006 2006110581 10/2006	wo	2010141135	12/2010
WO	2006110585 10/2006	WO	2010141135 A2	12/2010
WO WO	2006110599 10/2006	WO WO	2010144740 2011/005799 A2	12/2010 1/2011
WO	2007005645 1/2007 2007024323 3/2007	WO	2011005341 A3	1/2011
WO	2007024708 3/2007	WO	2011005799	1/2011
WO WO	2007064952 3/2007 2007059782 A1 5/2007	WO WO	2011/032633 A1 2011026641	3/2011 3/2011
WO	2007039782 A1 3/2007 2007062495 6/2007	WO	2011026641 A9	3/2011
WO	2007067968 6/2007	WO WO	2011062965 2011/069164 A2	5/2011 6/2011
WO WO	2007069068 A2 6/2007 2007095976 A2 8/2007	WO	2011/069164 A2 2011068810	6/2011
wo	2007100699 9/2007	WO	2011069528	6/2011
WO	2007100789 9/2007	WO WO	2011069529	6/2011 6/2011
WO WO	2007104537 9/2007 2008/003319 A1 1/2008	WO	2011069586 2011069587	6/2011
WO	2008/003319 A1 1/2008	WO	2011071931	6/2011
WO	2008/019371 A 2/2008	WO WO	2011071936 2011076807	6/2011 6/2011
WO WO	2008014979 2/2008 2008014979 A2 2/2008	WO	2011070807	7/2011
wo	2008014979 A2 2/2008 2008022046 A2 2/2008	WO	2011088309	7/2011
WO	2008042973 4/2008	WO WO	2011120053 2011127032 A1	9/2011 10/2011
WO WO	2008051245 5/2008 2008052770 5/2008	WO	2011127032 AT 2011127255	10/2011
wo	2008032770 3/2008	WO	2011127933 A1	10/2011
WO	2008078180 7/2008	WO WO	2011128444 2011130624	10/2011 10/2011
WO WO	2008078180 A2 7/2008 2008083949 7/2008	WO	2011130024 2011133868 A2	10/2011
wo	2008083949 A2 7/2008	WO	2011137206	11/2011
WO	2008091799 7/2008	WO WO	2011144358 2011161653	11/2011 12/2011
WO WO	2008/096370 A2 8/2008 2008107388 A1 9/2008	WO	2011101033 2012003474 A2	1/2011
wo	2008107388 A1 9/2008 2008115504 A2 9/2008	WO	2012006359	1/2012
WO	2008/134724 A2 11/2008	WO WO	2012006369 2012006372	1/2012 1/2012
WO WO	2008/143878 A2 11/2008 2008140615 11/2008	WO	2012006372	1/2012
wo	2008144365 11/2008	WO	2012006377	1/2012
WO	2008151049 A2 12/2008	WO WO	2012006378 2012006380	1/2012 1/2012
WO WO	2008151058 12/2008 2008153705 12/2008	wo	2012010855	1/2012
WO	2008157688 12/2008	WO	2012013326	2/2012
WO	2009006438 1/2009	WO WO	2012019168 2012019630	2/2012 2/2012
WO WO	2009015071 1/2009 2009024599 2/2009	wo	2012019780	2/2012
WO	2009030254 3/2009	WO	2012023044	2/2012
WO	2009030254 A1 3/2009	WO WO	2012024526 2012030683	2/2012 3/2012
WO WO	2009030481 3/2009 2009042971 4/2009	WO	2012030901	3/2012
WO	2009046738 4/2009	WO WO	2012030904 A2 2012031043	3/2012 3/2012
WO WO	2009046739 4/2009 2009046974 4/2009	WO	2012031043	3/2012
WO	2009046975 4/2009	WO	2012034067 A1	3/2012
WO	2009/068649 A2 6/2009	WO WO	2012034077 A2 2012045075	3/2012 4/2012
WO WO	2009077134 6/2009 2009095226 8/2009	WO	2012045082	4/2012
wo	2009101407 8/2009	WO	2012050975 A2	4/2012
WO	2009/113083 A1 9/2009	WO WO	2012064429 2012065164	5/2012 5/2012
WO WO	2009/120927 A2 10/2009 2009127060 10/2009	wo	2012068295	5/2012
wo	2009127230 10/2009	WO	2012068360	5/2012
WO	2009149253 12/2009	WO WO	2012068470 2012072269	5/2012 6/2012
WO WO	2010009065 1/2010 2010009277 1/2010	wo	2012072209	6/2012
WO	2010027903 3/2010	WO	2012088381	6/2012
WO	2010033906 3/2010	WO	2012089225	7/2012
WO WO	2010037408 4/2010 2010037408 A1 4/2010	WO WO	2012089338 2012094304	7/2012 7/2012
wo	2010037408 A1 4/2010 2010037539 4/2010	wo	2012094574	7/2012
WO	2010042490 4/2010	WO	2012099755	7/2012
WO	2010042877 4/2010 2010054406 5/2010	WO WO	2012099805 2012103985	7/2012 8/2012
WO WO	2010054406 5/2010 2010/068918 A2 6/2010	WO	2012103985 2012110636 A2	8/2012
0				

(56)	References	Cited	WO	2013045505	4/2013
	FOREIGN PATENT	DOCLIMENTS	WO WO	2013049234 2013049247	4/2013 4/2013
	FOREIGN PAIENT	DOCUMENTS	WO	2013049247	4/2013
WO	2012112582	8/2012	WO	2013052167	4/2013
WO	2012113413	8/2012	WO	2013052523	4/2013
WO		8/2012	WO WO	2013054307 2013055331	4/2013 4/2013
WO WO		9/2012 9/2012	wo	2013055905	4/2013
wo		9/2012	WO	2013055971	4/2013
WO		9/2012	WO WO	2013056132 2013057687	4/2013 4/2013
WO WO		9/2012 9/2012	WO	2013057087	4/2013
WO		9/2012 9/2012	WO	2013059496	4/2013
WO	2012125812	9/2012	WO	2013059509	4/2013
WO		9/2012	WO WO	2013/066866 A1 2013059922	5/2013 5/2013
WO WO		9/2012 0/2012	WO	2013061208	5/2013
wo		0/2012	WO	2013062140	5/2013
WO		0/2012	WO WO	2013063468 2013063530	5/2013 5/2013
WO WO		0/2012 0/2012	WO	2013063930	5/2013
WO		0/2012	WO	2013066274	5/2013
WO	2012142240 1	0/2012	WO	2013066427	5/2013
WO		0/2012	WO WO	2013066903 2013067355	5/2013 5/2013
WO WO		1/2012 1/2012	wo	2013067530	5/2013
wo		1/2012	WO	2013067537	5/2013
WO		1/2012	WO WO	2013068413 2013068431	5/2013 5/2013
WO WO		1/2012 1/2012	WO	2013068431	5/2013
WO		1/2012	WO	2013068847	5/2013
WO	2012/149376 A2 1	1/2012	WO	2013070653	5/2013
WO		1/2012	WO WO	2013070872 2013071047	5/2013 5/2013
WO WO		1/2012 1/2012	wo	2013071047	5/2013
wo		1/2012	WO	2013072929	5/2013
WO		1/2012	WO WO	2013074696	5/2013
WO		1/2012	WO	2013075068 2013077907	5/2013 5/2013
WO WO		1/2012 1/2012	WO	2013078199	5/2013
WO		1/2012	WO	2013/087911 A1	6/2013
WO		1/2012	WO WO	2013079604 2013082111	6/2013 6/2013
WO WO		2/2012 2/2012	wo	2013082418	6/2013
WO		2/2012	WO	2013082427	6/2013
WO		2/2012	WO WO	2013082470 2013082529	6/2013 6/2013
WO WO		2/2012 2/2012	WO	2013082529	6/2013
wo		2/2012	WO	2013084000	6/2013
WO	2012170930 1	2/2012	WO	2013085951	6/2013
WO WO		2/2012 2/2012	WO WO	2013086008 2013086322	6/2013 6/2013
WO		2/2012	WO	2013086354	6/2013
WO		1/2013	WO	2013086373	6/2013
WO		1/2013	WO WO	2013086486 2013086502	6/2013 6/2013
WO WO		1/2013 1/2013	WO	2013086505	6/2013
wo		1/2013	WO	2013086526	6/2013
WO	2013006834	1/2013	WO WO	2013087083 2013087791	6/2013 6/2013
WO WO		1/2013 1/2013	WO	2013087791 2013087912 A1	6/2013
WO		1/2013	WO	2013088250	6/2013
WO		1/2013	WO	2013090294	6/2013
WO		1/2013	WO WO	2013090601 2013090648	6/2013 6/2013
WO WO		1/2013 1/2013	wo	2013090841	6/2013
wo		1/2013	WO	2013090897	6/2013
WO	2013019669	2/2013	WO	2013090961	6/2013
WO		2/2013	WO WO	2013091001 2013093648	6/2013 6/2013
WO WO		3/2013 3/2013	WO	2013093648	6/2013
wo		3/2013	WO	2013096812 A1	6/2013
WO		3/2013	WO	2013098589	7/2013
WO		3/2013	WO	2013103842	7/2013
WO WO		3/2013 3/2013	WO WO	WO 2013/103659 A1 WO 2013/109713 A1	7/2013 7/2013
wo		3/2013	wo	2013112778	8/2013
WO		3/2013	WO	2013112780	8/2013

(56)	References Cited	WO 2014/043618 A1 3/2014
	EODEIGNI DATENTE DOGLINA	WO 2014/047649 A1 3/2014 RNTS WO 2014/052634 A1 4/2014
	FOREIGN PATENT DOCUME	WO 2014/053654 A1 4/2014 WO 2014/053654 A1 4/2014
WO	2012112226 8/2012	WO 2014/054026 A1 4/2014
WO	2013113326 8/2013 2013113501 8/2013	WO 2014/059022 A1 4/2014
wo	2013113502 8/2013	WO 2014053622 A1 4/2014
WO	2013113736 8/2013	WO 2014053624 A1 4/2014
WO	WO 2013/120497 A1 8/2013	WO 2014053628 A1 4/2014
WO	WO 2013/120498 A1 8/2013	WO 2014053629 A1 4/2014 WO 2014053634 A1 4/2014
WO	WO 2013/120499 A1 8/2013	WO 2014053634 A1 4/2014 WO 2014053879 A1 4/2014
WO WO	WO 2013/120500 A1 8/2013	WO 2014053880 A1 4/2014
WO	WO 2013/120626 A1 8/2013 WO 2013/120627 A1 8/2013	WO 2014053881 A1 4/2014
wo	WO 2013/120627 AT 6/2013 WO 2013/120628 A1 8/2013	WO 2014053882 A1 4/2014
WO	WO 2013/120629 A1 8/2013	WO 2014062697 A2 4/2014
WO	2013128027 9/2013	WO 2014063059 A1 4/2014
WO	2013130161 9/2013	WO 2014/064534 A2 5/2014 WO 2014/064543 A1 5/2014
WO	2013130535 9/2013	WO 2014/066811 A1 5/2014 WO 2014/066811 A1 5/2014
WO WO	2013135359 9/2013 2013136234 9/2013	WO 2014/066898 A9 5/2014
wo	2013138254 9/2013	WO 2014/066912 A1 5/2014
WO	2013138346 9/2013	WO 2014/071072 A2 5/2014
WO	2013142349 A1 9/2013	WO 2014/072468 A1 5/2014
WO	2013143555 10/2013	WO 2014/072747 A1 5/2014 WO 2014/072997 A1 5/2014
WO	2013143683 10/2013	WO 2014/072997 A1 5/2014 WO 2014/072999 A1 5/2014
WO WO	2013143698 10/2013 2013143699 10/2013	WO 2014/074218 A1 5/2014
WO	2013143699 10/2013 2013143700 10/2013	WO 2014/074299 A1 5/2014
WO	2013148186 10/2013	WO 2014/074597 A1 5/2014
WO	2013148541 10/2013	WO 2014064258 A1 5/2014
WO	2013149141 10/2013	WO 2014064687 A1 5/2014
WO	2013151650 10/2013	WO 2014067551 A1 5/2014 WO 2014068542 A1 5/2014
WO	2013151669 10/2013	WO 2014071219 A1 5/2014
WO WO	2013151672 10/2013 2013151771 10/2013	WO 2014071963 A1 5/2014
wo	2013152351 10/2013	WO 2014072061 A1 5/2014
WO	2013153550 10/2013	WO 2014072481 A1 5/2014
WO	2013154766 10/2013	WO 2014074289 A1 5/2014
WO	2013154774 10/2013	WO 2014074823 A1 5/2014 WO 2014074905 A1 5/2014
WO	2013155487 10/2013	WO 20140749303 AT 5/2014 WO 2014074912 A1 5/2014
WO WO	2013155493 10/2013 2013155513 10/2013	WO 2014075047 A2 5/2014
wo	2013158127 10/2013	WO 2014076709 A1 5/2014
WO	2013158141 10/2013	WO 2014078399 A1 5/2014
WO	2013158579 10/2013	WO 2014078636 A1 5/2014
WO	2013/177421 A2 11/2013	WO 2014081299 A1 5/2014 WO 2014081300 A1 5/2014
WO WO	2013166385 11/2013 2013166498 11/2013	WO 2014081303 A1 5/2014
WO	2013173582 A1 11/2013	WO 2014081507 A1 5/2014
wo	2013173657 11/2013	WO 2014081849 A1 5/2014
WO	2013173693 11/2013	
WO	2013174409 11/2013	OTHER PUBLICATIONS
WO	2013182683 12/2013	Ellem, K.A.O., and Colter, J.S. The isolation of three variants of
WO WO	2013184945 12/2013 2013185069 12/2013	merigo virus differing in plaque morphology and hemagglutinating
WO	2013188979 12/2013	characteristics. Virology. Nov. 1961; 15(3): 340-347.
WO	2014004436 A2 1/2014	Ellem, K.A.O., and Colter, J.S. The interaction of infectious ribo-
WO	2014012479 A1 1/2014	nucleic acid with a mammalian cell line: I. Relationship between the
WO	2014012994 1/2014	osmotic pressure of the medium and the production of infectious
WO WO	2014012996 1/2014 2014014613 1/2014	centers. Virology. Jun. 1960; 11(2): 434-443.
WO	2014014890 A1 1/2014	Ellem, K.A.O. and Colter, J.S. The interaction of infectious ribo-
wo	2014015334 1/2014	nucleic acid with a mammalian cell line: II. Kinetics of the
WO	2014015422 1/2014	formation of infectious centers. Virology, Dec. 1960: 12(4): 511-
WO	2014016439 1/2014	520.
WO	2014018675 1/2014	Ellen K.A.O and Colter, J.S. The interaction of infectious ribo-
WO	2014024193 2/2014	nucleic acids with mammalian cells: III. Comparison of infection
WO WO	2014025312 2/2014 2014025795 2/2014	and RNA uptake in the HeLa cell-polio RNA and L cell-rnengo
WO	2014023793 2/2014 2014025890 2/2014	RNA systems. Virology. Oct. 1961; 15(2): 113-126.
wo	2014026044 2/2014	Epicentre Forum. Tools and Techniques for Genomics and RNA
WO	2014026284 2/2014	Research. 2006; 13(2): 1-3.
WO	2014027006 2/2014	Epicentre Forum. Tools and Techniques for Genomics and RNA
WO	2014028209 2/2014	Research. 2007; 14(1): 1-24.
WO WO	2014028487 2/2014	Esposito, S., Effect on Leukaemic Cells of Ribonucleic Acid Extracted from Calf's Spleen. Nature. Sep. 1964; 203: 1078-1079.
WO	2014028763 2/2014 2014/039185 A1 3/2014	Esvelt, K., et al., A system for the continuous directed evolution of
WO	2014/039183 A1 3/2014 2014/042920 A1 3/2014	biomolecules. Nature. Apr. 2011; 472(7344): 499-503.
		T. 2011, 130, 130

OTHER PUBLICATIONS

Fahy, E. et al., Self-sustained sequence replication (3SR): an isothermal transcription-based amplification system alternative to PCR. PCR Methods Appl. Aug. 1, 1991(1):25-33.

Faissner, A. et al., Analysis of polypeptides of the tree shrew (Tupaia) herpesvirus by gel electrophoresis. J Gen Virol. Jan. 1982;58 Pt 1:139-48.

Fan, X.C., et al., Overexpression of HuR, a nuclear-cytoplasmic shuttling protein, increases the in vivo stability of ARE-containing mRNAs. Embo J 1998; 17(12): 3448-3460.

Fandrich, F. et al., Preimplantation-stage stem cells induce long term allogeneic graft acceptance without supplementary host conditioning. Nat Med. Feb. 2002;8(2):171-8.

Fang, S.H. et al., Functional measurement of hepatitis C virus core-specific CD8(+) T-cell responses in the livers or peripheral blood of patients by using autologous peripheral blood mononuclear cells as targets or stimulators. J Clin. Microbiol. Nov. 2001;39(11):3895-901.

Fearnley, D.B. et al., Monitoring human blood dendritic cell numbers in normal individuals and in stem cell transplantation. Blood. Jan. 15, 1999;93(2):728-36.

Felgner, P.L., et al., Lipofection: a highly efficient, lipid-mediated DNA-transfection procedure Proc Natl Acad Sci U S A. Nov. 1987;84(21):7413-7.

Felgner, P.L. Particulate systems and polymers for in vitro and in vivo delivery of polynucleotides. Adv. Drug Delivery Rev. 1990; 5(3): 163-187.

Felgner, P.L. Cationic lipid/polynucleotide condensates for in vitro and in vivo polynucleotide delivery—the cylofectins. J. of Liposome Research. 1993; 3(1): 3-16.

Fisch, P. et al., Generation of antigen-presenting cells for soluble protein antigens ex vivo from peripheral blood CD34+hematopoietic progenitor cells in cancer patients, Eur J Immunol. Mar. 1996;26(3):595-600.

Fisher, K.J. and Wilson, J.M. The transmembrane domain of diphtheria toxin improves molecular conjugate gene transfer. Biochem. J. Jan. 1997; 321(1): 49-58.

Fishman, M., et al., In vitro transfer of macrophage RNA to lymph node cells. Nature. May 11, 1963;198:549-51.

Fisk, B. et al., Identification of an immunodominant peptide, of HER-2/neu protooncoclene recognized by ovarian tumor-specific cytotoxic T lymphocyte lines. J Exp Med. Jun. 1, 1995;181(6):2109-17

Frank, B. et al., Interanimal "memory" transfer: results from brain and liver homogenates. Science. Jul. 24, 1970;169(3943):399-402. Franklin, R.M., Purification and properties of the replicative intermediate of the RNA bacteriophage R17. Proc Natl Acad Sci U S A. Jun. 1966;55(6):1504-11.

Frey, M.R. et al., RNA-mediated interaction of Cajal bodies and U2 snRNA genes. J Cell Biol. Aug. 6, 2011;154(3):499-509.

Fukuda, N., et al., In vitro evolution of single-chain antibodies using mRNA display. Nucleic Acids Res. 2006;34(19): e127. Epub Sep. 29, 2006.

Fusaki, N., et al., Efficient induction of transgene-free human pluyripotent stem cells using a vector based on Sendai virus, an RNA virus that does not integrate into the host genome. Proc Jpn Acad Ser B Phys Biol Sci. 2009; 85(8): 348-362.

Fynan E.F. et al., DNA vaccines: protective immunizations by parenteral, mucosal, and gene-gun inoculations. Proc Natl Acad Sci U S A. Dec. 15, 1993;90(24):11478-82.

Gall, J.G. et al., A role for Cajal bodies in assembly of the nuclear transcription machinery. FEBS Lett. Jun. 8, 2001;8;498(2-3):164-7. Gall, J.G. The centennial of the Cajal body. Nat Rev mol Cell Biol. Dec. 2003;4(12):975-80.

Gallie, D.R. A tale of two termini: a functional interaction between the termini of an mRNA is a prerequisite for efficient translation initiation. Gene. Aug. 17, 1998;216(1):1-11.

Gallie, D.R. The cap and poly(A) tail function synergistically to regulate mRNA translational efficiency. Genes Dev. Nov. 1991;5(11):2108-16.

Ganot, P. et al., Site-specific pseudouridine formation in preribosomal RNA is guided by small nucleolar RNAs. Cell. May 30, 1997;89(5):799-809.

Gao, M. et al., A novel mRNA-decapping activity in HeLa cytoplasmic extracts is regulated by AU-rich elements. EMBO J. Mar. 1, 2001;20(5):1134-43.

Gao, X. et al., Nonviral gene delivery, what we know and what is next. AAPS J. Mar. 23, 2007;9(1):E92-104.

Garbe, C. et al., [Epidemiology of malignant melanoma in West Germany in an international comparison]. Onkologie. Dec. 1989;12(6):253-62.

Gardiner-Garden, M. et al., CpG islands in vertebrate genomes. J Mol Biol. Jul. 20, 1987;196(2):261-82.

Gasche, C. et al., Sequential treatment of anemia in ulcerative colitis with intravenous iron and erythropoietin. Digestion. 1999;60(3):262-7.

GenBank NP_000651.3. Transforming growth factor beta-1 precursor [*Homo sapiens*]. Nov. 13, 2001; online.

Gerbi, S.A. et al., All small nuclear RNAs (snRNAs) of the [U4/U6,U5] Tri-snRNP localize to nucleoli; Identification of the nucleolar localization element of U6 snRNA. Mol Biol Cell. Sep. 2002;13(9):3123-37.

Gershon, P.D., (A)-tail of two polymerase structures. Nat Struct Biol. Oct. 2000;7(10):819-21.

Gierer, A and Schramm, G. Infectivity of ribonucleic acid from tobacco mosaic virus. Nature. Apr. 1956; 177(4511):702-703.

Gilboa, E. et al., Cancer immunotherapy with mRNA-transfected dendritic cells. Immunol Rev. Jun. 2004;199:251-63.

Giljohann, D.A., et al., Gene regulation with polyvalent siRNA-nanoparticle conjugates. J Am Chem Soc. Feb. 2009;131(6): 2072-2073

Gilkeson, G.S. et al., Induction of cross-reactive anti-dsDNA anti-bodies in preautoimmune NZB/NZW mice by immunization with bacterial DNA. J Clin Invest. Mar. 1995;95(3):1398-402.

Ginsberg, S.D. et al., Expression profile of transcripts in Alzheimer's disease tangle-bearing CA1 neurons. Ann Neurol. Jul. 2000;48(1):77-87.

Ginsberg, S.D. et al., Predominance of neuronal mRNAs in individual Alzheimer's disease senile plaques. Ann Neurol. Feb. 1999;45(2):174-81.

Hoerr, I. et al., In vivo application of RNA ads to induction of specific cytotoxic T lymphocytes and antibodies. Eur J Immunol. Jan. 2000;30(1):1-7.

Hoerr, I. et al., Stabilized Messenger RNA (RNActiveTM) as a Tool for Innovative Gene Delivery. Tissue Engineering. Apr. 2007; 13(4): 865-925.

Holcik, M. et al., Four highly stable eukaryotic mRNAs assemble 3'untranslated region RNA-protein complexes sharing cis and trans components. oc Natl Acad Sci U S A. Mar. 18, 1997;94(6):2410-4. Holmes, D. et al., Cell positioning and sorting using dielectrophoresis. Eur Cell Mater. 2002; 4(2):120-2.

Holtkamp, S. et al., Modification of antigen-encoding RNA increases stability, translational efficacy, and T-cell stimulatory capacity of dendritic cells. Blood. Dec. 15, 2006;108(13):4009-17. Houghton, A.N. et al., Cancer antigens: immune recognition of self and altered self. J Exp Med. Jul. 1, 1994;180(1):1-4.

Hsu, F.J. et al., Vaccination of patients with B-cell lymphoma using autologous antigen-pulsed dendritic cells. Nat Med. Jan. 1996;2(1):52-8.

Hu, B., et al., Neural differentation of human induced pluripotent stem cells follows developmental principles but with variable potency. Natl Acad Sci. Mar. 2010; 107(9): 4335-4340.

Hu, S. et al., Codon optimization, expression, and characterization of an internalizing anti-ErbB2 single-chain antibody in Pichia pastoris. Protein Expr. Purif. May 2006;47(1):249-57. Epub Dec. 13, 2005.

Huangfu, D., et al., Induction of pluripotent stem cells by defined factors is greatly improved by small-molecule compounds. Nat Biotech. Jul. 2008; 26(7) 795-797.

Huddleston, J.A. et al., The sequence of the nucleoprotein gene of human influenza A virus, strain A/NT/60/68. Nucleic Acids Res. Feb. 11, 1982;10(3):1029-38.

OTHER PUBLICATIONS

Hue, K.K. et al., A polypurine sequence that acts as a 5' mRNA stabilizer in Bacillus subtilis. J Bacteriol. Jun. 1995;177 (12):3465-71

Hung, C.F. et al., Ovarian cancer gene therapy using HPV-16 pseudovirion carrying the HSV-tk gene. PLoS ONE. Jul. 2012; 7(7):e40983.

Inaba, K. et al., Dendritic cells pulsed with protein antigens in vitro can prime antigen-specific, MHC-restricted T cells in situ. J Exp Med. Aug. 1, 1990;172(2):631-40.

Inaba, K. et al., Direct activation of CD8+ cytotoxic T lymphocytes by dendritic cells. J Exp Med. Jul. 1, 1987;166(1):182-94.

Inaba, K. et al., Generation of large numbers of dendritic cells from mouse bone marrow cultures supplemented with granulocyte/macrophage colony-stimulating factor, J Exp Med. Dec. 1, 1992;176(6):1693-702.

International Search Report from International Application No. PCT/US11/54617 dated Oct. 3, 2011.

International Search Report from International Application No. PCT/US11/54617 dated Feb. 1, 2012.

International Search Report from International Application No. PCT/US2012/031781 dated Jan. 11, 2013.

International Search Report from International Application No. PCT/US12/38028 dated Aug. 14, 2012.

International Search Report from International Application No. PCT/US12/54561 dated Feb. 26, 2013.

International Search Report from International Application No. PCT/US12/58519 dated Feb. 28, 2013.

International Search Report from International Application No PCT/US12/68732 dated Feb. 22, 2013.

International Search Report from International Application No.

PCT/US12/69610 dated Feb. 27, 2013. International Search Report from International Application No.

PCT/US12/71105 dated Mar. 5, 2013. International Search Report from International Application No.

PCT/US13/20921 dated Mar. 26, 2013. International Search Report from International Application No.

PCT/US12/71118 dated Apr. 5, 2013. Ito, M.K., ISIS 301012 gene therapy for hypercholesterolemia:

sense, antisense, or nonsense? Ann Pharmacother. Oct. 2007; 41(10): 1669-78.

Ivanovska, N. et al. Immunization with a DNA chimeric molecule encoding a hemagglutinin peptide and scFv CD21-specific antiobdy fragment induces long-lasting IgM and CTL responses to influenza virus. Vaccine. Mar. 10, 2006;24(11):1830-7. Epub Nov. 2, 2005. Iwasaki, A. et al., Enhanced CTL responses mediated by plasmid DNA immunogens encoding costimulatory molecules and cytokines. J. Immunol. May 15, 1997;158(1):4591-601.

Jady, B.E. et al., A small nucleolar guide RNA functions both in 2'-O-ribose methylation and pseudouridylation of the U5 spliceosomal RNA. EMBO J. Feb. 1, 2001;20(3):541-51.

Janeway, C. et al., Immunobiology: the immune system in health and disease. Garland Publishing, Inc, London. 1997; 13:12-13:21. Jansen, P.L.M., Diagnosis and management of Crigler-Najjar syndrome. Eur J Pediatr. Dec. 1999;158 [Suppl 2]:S89-S94.

Janssens, S. et al., Role of Toll-like receptors in pathogen recognition. Clin Microbiol Rev. Oct. 2003;16(4):637-46.

Jemielity, J. et al., Novel "anti-reverse" cap analogs with superior translational properties. RNA. Sep. 2003;9(9):1108-22.

Jia, F., et al., A nonviral minicircle vector for deriviing human iPS Cells. Nat Methods. Mar. 2010; 7(3): 197-199.

Jia, Z., et al., Long-term correction of hyperbilirubinemia in the Gunn Rat by repeated intravenous delivery of naked plasmid DNA into muscle. Mol Ther. Nov. 2005; 12(5): 860-866.

Jiang, J. et al., Topical application of ketoconazole stimulates hair growth in C3H/HeN mice. J Dermatol. Apr. 2005;32(4):243-7.

Jirikowski, G.F., et al., Reversal of diabetes insipidus in Brattleboro Rats: Intrahypothalamic injection of vasopressin mRNA. Science. Feb. 1992; 255(5047): 996-998.

Johnson, K.M. et al., Role of heparan sulfate in attachment to and infection of the murine female gential tract by human papillomavirus. J Virol. Mar. 2009; 83(5): 2067-2074.

Jones, P.C.T., An Alteration in Cell Morphology under the Influence of a Tumor RNA. Nature, 1964,202:1226-7.

Juliano, R.L., et al., Cell-targeting and cell-penetrating peptides for delivery of therapeutic and imaging agents. Wiley Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology. May/Jun. 2009; 1(3): 324-335.

Kabanov, A V. et al., A new class of antivirals: antisense oligonucleotides combined with a hydrophobic substituent effectively inhibit influenza virus reproduction and synthesis of virus-specific proteins in MDCK cells. FEBS Lett. Jan. 1, 1990;259(2):327-30. Kahan, F.M. et al., The role of deoxyribonucleic acid in ribonucleic acid synthesis. J Biological Chem. Dec. 1962; 287(12): 3778-3785. Kaji, K., et al., Virus free induction of pluripotency and subsequent excision of reprogramming factors. Nature. Apr. 2009; 458(7239): 771-775.

Kalnins, A. et al., Sequence of the lacZ gene of $Escherichia\ coli.$ EMBO J. 1983;2(4):593-7.

Kanaya, S. et al., Codon usage and tRNA genes in eukaryotes: correlation of codon usage diversity with translation efficiency and with CG-dinucleotide usage as assessed by multivariate analysis. J Mol Evol. Oct.-Nov. 2001;53(4-5):290-8.

Kandimalla, E.R. et al., Divergent synthetic nucleotide motif recognition pattern: design and development of potent immunomodulatory oligodeoxyribonucleotide agents with distinct cytokine induction profiles. Nucleic Acids Res. May 1, 2003;31(9):2393-400.

Kandimalla, E.R. et al., Immunomodulatory oligonucleotides containing a cytosine-phosphate-2'-deoxy-7-deazaguanosine motif as potent toll-like receptor 9 agonists. Proc Natl Acad Sci U S A. May 10, 2005:102(19):6925-30. Epub Apr. 28, 2005.

Karande, A.A., et al., In vitro induction of chronic myeloid leukemia associated immune reactivity in normal human lymphocytes by xenogeneic immune RNA. Neoplasma. 1983, 30(4):403-9.

Kuhn, E., et al., Developing multiplexed assays for Troponin I and Interleukin-33 in plasma by peptide immunoaffinity enrichment and targeted mass spectrometry. Clinical Chem. 2009; 55(6): 1108-1117.

Kundu, T.K. et al., CpG islands in chromatin organization and gene expression. J Biochem. Feb. 1999;125(2):217-22.

Kusakabe, K. et al., The timing of GM-CSF expression plasmid administration influences the Th1/Th2 response induced by an HIV-1-specific DNA vaccine. J Immunol. Mar. 15, 2000;164(6):3102-11.

Kvasnica, M. et al., Platinum(II) complexed with steroidal esters of L-methonine and L-histidine: synthesis, characterization and cytotoxic activity. Bioorg Med Chem. Apr. 1, 2008;16(7):3704-13. Epub Feb. 7, 2008.

Kwoh, D.Y. et al., Transcription-based amplification system and detection of amplified human immunodeficiency virus type 1 with a bead-based sandwich hybridization format. Proc Natl Acad Sci U S A. Feb. 1989;86(4):1173-7.

Kwissa, M. et al., Cytokine-facilitated priming of CD8+ T cell responses by DNA vaccination. J Mol Med (Berl). Feb. 2003;81(2):91-101. Epub Nov. 22, 2002.

Lacour, F. et al., Transplantable malignant tumors in mice induced by preparations containing ribonucleic acid extracted from human and mouse tumors. J. Natl Cancer Inst., 1960, 24(2):301-27.

Lai, C.J. et al., Patterning of the neural ectoderm of Xenopus laevis by the amino-terminal product of hedgehog autoproteolytic cleavage. Development. Aug. 1995;121(8):2349-60.

Lai, S.K., et al., Rapid transport of large polymeric nanoparticles in fresh undiluted human mucus. PNAS. Jan. 30, 2007;104(5): 1482-1487.

Lai, S.K., et al., Mucus-penetrating nanoparticles for drug and gene delivery to mucosal tissues. Adv Drug Deliv Rev. Feb. 27, 2009; 61(2): 158-171.

Lange, T.S., et al., Transient nucleolar localization of U6 small nuclear RNA in Xenopus laevis oocytes. Mol Biol Cell. Jul. 2000;11(7):2419-28.

OTHER PUBLICATIONS

Langford, C.J. et al., Evidence for an intron-contained sequence required for the splicing of yeast RNA polymerase II transcripts. Cell. Jun. 1983;33(2):519-27.

Larregina, A.T. et al., Changing paradigms in cutaneous immunology: adapting with dendritic cells. J Invest Dermatol. Jan. 2005;124(1):1-12.

Latarjet, R., Production of multiple cancers in mice having received nucleic acid extract from isologous & homologous leukemic tissues. C.R. Hebd Seances Acad. Sci., 1958, 246(5):853-5.

Lathe, R., Synthetic oligonucleotide probes deduced from amino acid sequence data: Theoretical and practical considerations. J Mol Biol. May 5, 1985;183(1):1-12.

Leader B., et al., Protein therapeutics: a summary and pharmacological classification. Nat Rev Drug Discov. Jan. 2008; 7(1): 21-39. Lee, G. et al., Modeling pathogenesis and treatment of familial dysautonomia using patient-specific iPSCs. Nature. Sep. 17, 2009;461(7262):402-6. Epub Aug. 19, 2009.

Lee, J. et al., Molecular basis for the immunostimulatory activity of guanine nucleoside analogs: activation of Toll-like receptor 7. Proc Natl Acad Sci U S A. May 27, 2003;100(11):6646-51. Epub May 8, 2003.

Lee, J. T., et al., An arginine to glutamine mutation in residue 109 of human ornithine transcarbamylase completely abolishes enzymatic activity in Cos1 cells. J. Clin. Invest. Dec. 1989; 84: 1762-1766

Lehto, T., et al., Cell-penetrating peptides for the delivery of nucleic acids. Expert Opin. Drug Deliv. Jul. 2012; 9(7): 823-836.

Leitner, W.W. et al., DNA and RNA-based vaccines: principles, progress and prospects. Vaccine. Dec. 10, 1999;18(9-10):765-77. Lenz, A. et al., Human and murine dermis contain dendritic cells. Isolation by means of a novel method and phenotypical and functional characterization. J Clin Invest. Dec. 1993;92(6):2587-96.

Lerner, M.R. et al., Are snRNPs involved in splicing? Nature. Jan. 10, 1980;283(5743):220-4.

Lesaffre, B. et al., Direct non-cell autonomous Pax6 activity regulates eye development in the zebrafish. Neural Dev. Jan. 17, 2007;2:2.

Lewandowski, L.J. et al., Separation of the infectious ribonucleic acid of potato spindle tuber virus from double-stranded ribonucleic acid of plant tissue extracts. J Virol. Nov. 1971;8(5):809-12.

Lewis, David, Dynamic Polyconjugates (DPC) Technology: An elegant solution to the siRNA delivery problem. Arrowhead Research Corp (NASDAQ: ARWR). Nov. 2011.

Lewis, J.D. et al., The influence of 5' and 3' end structures on pre-mRNA metabolism. J Cell Sci Suppl. 1995;19:13-9.

Lewis, J.K., et al., Matrix-assisted laser desorption/ionization mass spectrometry in peptide and protein analysis. Enc of Anal Chem. 2000; R.A. Meyers (Ed.) 5880-5894.

Li, L. et al., Preparation and gene delivery of alkaline amino acids-based cationic liposomes. Arch Pharm Res. Jul. 2008;31(7):924-31. Epub Aug. 14, 2008.

Li, L. et al., Overcoming obstacles to develop effective and safe siRNA therapeutics. Expert Opin Biol Ther. May 2009;9(5): 609-19. Li, X. et al., Generation of destabilized green flourescent protein as a transcription reporter. J Biol Chem. Dec. 25, 1998;273(52):34970-5

Lian, T. et al., Trends and developments in liposome drug delivery systems. J Pharm Sci. Jun. 2001;90(6):667-80.

Liang, X.H. et al., The spliced leader-associated RNA is a trypanosome-specific sn(o) RNA that has the potential to guide pseudouridine formation on the SL RNA. RNA. Feb. 2002;8(2):237-46.

Licatalosi, D.D. et al., Splicing regulation in neurologic disease. Neuron. Oct. 5, 2006;52(1):93-101.

Linehan, D.C. et al., Tumor-specific and HLA-A2 restricted cytolysis by tumor-associated lymphocytes in human metastatic breast cancer. J Immunol. Nov. 1, 1995;155(9):4486-91.

Lobenberg, R. et al., Improved body distribution of 14C-labelled AZT bound to nanoparticles in rats determined by radioimmunography. J Drug Target. 1998;5(3):171-9.

Loging, W.T. et al., Identifying potential tumor markers and antigens by database mining and rapid expression screening. Genome Res. Sep. 2000;10(9):1393-402.

Lopez, M.F., et al., Selected reaction monitoring-mass spectrometric immunoassay responsive to parathyroid hormone and related variants. Clinical Chem. 2010; 56(2): 281-290.

Lopez-Berestein, G. et al., Treatment of systemic fungal infections with liposomal amphotericin B. Arch Intern Med. Nov. 1989;149(11):2533-6.

Lorenzi, J.C., et al., Intranasal vaccination with messenger RNA as a new approach in gene therapy: Use against tuberculosis. BMC Biotechnocol. Oct. 2010; 10(77): 1-11.

Lowe, T.M. et al., A computational screen for methylation guide snoRNAs in yeast. Science. Feb. 19, 1999;283(5405):1168-71.

Lowry, W.E., et al., Generation of human induced pluripotent stem cells from dermal fibroblasts. Proc Natl Acad Sci USA. Feb. 2008; 105(8): 2883-2888.

Lukkonen, B.G. et al., A conditional U5 snRNA mutation affecting pre-mRNA splicing and nuclear pre-mRNA retention identifies SSD1/SRK1 as a general splicing mutant suppressor. Nucleic Acids Res. Sep. 1, 1999;27 (17):3455-65.

Lund, P.E., et al., Pseudovirions as vehicles for the delivery of siRNA. Pharm Res. Mar. 2010; 27(3): 400-420. Epub Dec. 9, 2009. Luo, D. et al., Synthetic DNA delivery systems. Nat Biotechnol. Jan. 2000;18(1):33-7.

Ma, X. et al., Pseudouridylation (Psi) of U2 snRNA in *S. cerevisiae* is catalyzed by an RNA-independent mechanism. EMBO J. Apr. 15, 2003;22(8):1889-97.

Mackie, G.A., Vectors for the synthesis of specific RNAs in vitro. Biotechnology. 1988;10:253-67.

Maden, B.E.H. et al., Classical and novel approaches to the detection and localization of the numerous modified nucleotides in eukaryotic ribosomal RNA. Biochimie. 1995;77(1-2):22-9.

Langer, R., New methods of drug delivery. Science. Sep. 28, 1990;249(4976):1527-33.

Magee, W. E. et al., Marked stimulation of lymphocyte-mediated attack on tumor cells by target-directed liposomes containing immune RNA, Cancer Res., 1978, 38(4):1173-6.

Warren, L. et al. Highly efficient reprogramming to pluripotency and directed differentiation of human cells with synthetic modified mRNA. Cell Stem Cell. Nov. 5, 2010;7(5):618-30.

Kormann, M. et al. Expression of therapeutic proteins after delivery of chemically modified mRNA in mice, Nat Biotechnol. Feb. 2011;29(2):154-7.

Mannick, J.A. et al., Transformation of Nonimmune Lymph Node Cells to a State of Transplantation Immunity by RNA. A Preliminary Report, Ann. Surg., 1962, 156:356-66.

Mansour, S.L. et al., Disruption of the proto-oncogene int-2 in mouse embryo-derived stem-cells: a general strategy for targeting mutations to non-selectable genes. Nature, 1988, 336:348-52.

Mansour, et al., Functional Studies with Uterine RNA. PNAS, 1965, 53:764-70.

Marson, A., et al., Wnt signaling promotes reprogramming of somatic cells to pluripotency. Cell Stem Cell. Aug. 2008; 3(2): 132-135.

Martin, S.A. et al., Purification of mRNA guanylyltransferase and mRNA (guanine-7-) methyltransferase from vaccinia virions. J Biol Chem. Dec. 25, 1975;250(24):9322-9.

Martinon, F. et al., Induction of virus-specific cytotoxic T lymphocytes in vivo by liposome-entrapped mRNA. Eur J. Immunol. Jul. 1993;23(7):1719-22.

Massenet, S. et al., Pseudouridine mapping in the *Saccharomyces cerevisiae* spliceosomal U small nuclear RNAs (snRNAs) reveals that pseudouridine synthase pus1p exhibits a dual substrate specificity for U2 snRNA and tRNA. Mol Cell Biol. Mar. 1999;19(3):2142-54.

Mathers, A.R. et al., Professional antigen-presenting cells of the skin. Immunol Res. 2006;36(1-3):127-36.

OTHER PUBLICATIONS

Matray, T.J. et al., Synthesis and properties of RNA analogsoligoribonucleotide N3'→P5' phosphoramidates. Nucleic Acids Res. Oct. 15, 1999;27(20):3976-85.

Maurer, N., et al., Spontaneous entrapment of polynucleotides upon electrostatic interation with ethanol-destabilized cationic liposomes. Biophys J. May 2001; 80(5): 2310-2326.

Mayfield, S.P. et al., Expression and assembly of a fully active antibody in algae. Proc Natl Acad Sci U S A. Jan. 21, 2003;100(2):438-42. Epub Jan. 8, 2003.

McCafferty, J et al., Phage antibodies: filamentous phage displaying antibody variable domains. Nature. Dec. 6, 1990;348(6301):552-4. McCormack, A.L., et al., a-Synuclein suppression by targated small interfering RNA in the primate substantia nigra. PLoS ONE. Aug. 2010; 5(8): e12122.

McCormack, M., et al., Activation of the T-cell oncogene LMO2 after gene therapy for X-linked severe combined immunodeficiency. N Engl J Med. Feb. 2004; 350: 931-922.

McDonald, J.D., et al., Characterization of mutations at the mouse phenylalanine hydroxylase locus. Genomics. 1997; 39: 402-405.

McElwee, K.J. et al., Transfer of CB8(+) cells induces localized hair loss whereas CD4(+)/CD25(-) cells promote systemic alopecia areata and CD4(+)/CD25(+) cells blockade disease onset in the C3H/HeJ mouse model. J Invest Dermatol. May 2005;124(5):947-57

McGee, M. et al., The Quantitative determination of phenylalanine hydroxylase in rat tissues. Biochem. J. 1972; 127:669-674.

McGlynn, R. et al., Differential subcellular localization of cholesterol, gangliosides, and glycosaminoglycans in murine models of mucopolysaccharide storage disorders. J Compl Neurol. Dec. 20, 2004;480(4):415-26.

McKenzie, B.S. et al., Nucleic acid vaccines: tasks and tactics. Immunol Res. 2001:24(3):225-44.

McLean, M.J., et al., Membrane differentiation of cardiac myoblasts induced in vitro by an RNA-enriched fraction from adult heart. Exp Cell Res. Nov. 1977;110(1):1-14.

MEGAscript Kit product Manual, Ambion/Invitrogen website: http://tools.invitrogen.com/content/sfs/manuals/cms_072987.pdf, Publication Date: Oct. 27, 2009 (last accessed Mar. 17, 2013).

Mellits, K.H. et al., Removal of double-stranded contaminants from RNA transcripts: synthesis of adenovirus VA RNAI from a T7 vector. Nucleic Acids Res. Sep. 25, 1990;18(18):5401-6.

Meunier, L. et al., Heterogeneous populations of class II MHC+cells in human dermal cell suspensions. Identification of a small subset responsible for potent dermal antigen-presenting cell activity with features analogous to Langerhans cells. J Immunol. Oct. 15, 1993;151(8):4067-80.

Mignone, F. et al., Untranslated regions of mRNAs. Genome Biol. 2002;3(3):REVIEWS0004. Epub Feb. 28, 2002. pp. 1-10.

Minks, M.A. et al., Structural requirements of double-stranded RNA for the activation of 2',5'-oligo(A) polymerase and protein kinase of interferon-treated HeLa cells. J Biol Chem. Oct. 25, 1979;254(20):10180-3.

Mishra, N.C. et al., Induction by RNA of inositol independence in Neurospora crassa. Proc. Natl Acad. Sci. U.S.A., 1975, 72(2):642-5. Mishra, R.K. et al., Improved leishmanicidal effect of phosphorotioate antisense oligonucleotides by LDL-mediated delivery. Biochim Biophys Acta. Nov. 7, 1995;1264(2):229-37.

Mitchell, D.A. et al., RNA transfected dendritic cells as cancer vaccines. Curr Opin Mol Ther. Apr. 2000;2(2):176-81.

Mitchell, D.A. et al., RNA-transfected dendritic cells in cancer immunotherapy. J Clin Invest. Nov. 2000;106(9):1065-9.

Mitchell, P. et al., mRNA turnover. Curr Opin Cell Biol. Jun. 2001;13(3):320-5.

Miura, K., et al., Variation in the safety of induced pluripotent stem cell lines. Nat Biotechnology. Aug. 2009; 27(8):743-745.

Morinaga, T. et al., Primary structures of human alpha-fetoprotein and its mRNA. Proc Natl Acad Sci U S A. Aug. 1983;80(15):4604-8

Morse, M.A. et al., Generation of dendritic cells in vitro from peripheral blood mononuclear cells with granulocyte-macrophage-colony-stimulating factor, interleukin-4, and tumor necrosis factoralpha for use in cancer immunotherapy. Ann Surg. Jul. 1997;226(1):6-16.

Mount, S.M. et al., A catalogue of splice junction sequences. Nucleic Acids Res. Jan. 22, 1982;10(2):459-72.

Muller, M.R. et al., Transfection of dendritic cells with RNA induces CD4- and CD8-mediated T cell immunity against breast carcinomas and reveals the immunodominance of presented T cell epitopes. J Immunol. Jun. 15, 2003;170(12):5892-6.

Murakawa, G.J. et al., Direct detection of HIV-1 RNA from AIDS and ARC patient samples. DNA. May 1988;7(4):287-95.

Myette, J.R. et al., Domain structure of the vaccinia virus mRNA capping enzyme. Expression in *Escherichia coli* of a subdomain possessing the RNA 5'-triphosphatase and guanylyltransferase activities and a kinetic comparison to the full-size enzyme. J Biol Chem. May 17, 1996;271(2):1936-44.

Nagata, S., et al., Molecular cloning and expression of cDNA for human granulocyte colony-stimumating factor. Nature. Jan. 30-Feb. 5, 1986; 319(6052): 415-8.

Nagata, S., et al., The chromosomal gene structure and two mRNAs for human granulocyte colony-stimulating factor. EMBO J. Mar. 1986; 5(3): 575-81.

Nagata, T. et al., Codon optimization effect on translational efficiency of DNA vaccine in mammalian cells: analysis of plasmid DNA encoding a CTL epitope derived from microorganisms. Biochem Biophys Res Commun. Aug. 2, 1999;261(2):445-51.

Nair, S. et al., Soluble proteins delivered to dendritic cells via pH-sensitive liposomes induce primary cytotoxic T lymphocyte responses in vitro. J Exp Med. Feb. 1, 1992;175(2):609-12.

Nair, S.K. et al., Antigen-presenting cells pulsed with unfractionated tumor-derived peptides are potent tumor vaccines. Eur J Immunol. Mar. 1997;27(3):589-97.

Nair, S.K. et al., Induction of cytotoxic T cell responses and tumor immunity against unrelated tumors using telomerase reverse transcriptase RNA transfected dendritic cells. Nat Med. Sep. 2000;6(9):1011-7.

Nair, S.K. et al., Induction of primary carcinoembryonic antigen (CEA)-specific cytotoxic T lymphocytes in vitro using human dendritic cells transfected with RNA. Nat Biotechnol. Apr. 1998;16(4):364-9.

Nakamura, K. et al., A model for the autosensitization autoantibody production associated with xenogeneic thymic RNA. J Immunol. Aug. 1978;121(2):702-9.

Nakamura, K. et al., Antigen restricted hybridization between antigen primed macrophage and thymic RNA. Immunol Commun. 1981;10(4-5):367-82.

Nakamura, K. et al., Conversion of immune response patterns from high to low and low to high by an RNase-sensitive thymocyte extract. Immunology. Sep. 1980;41(1):25-35.

Nakamura, K. et al., Generation of anti-NZB red blood cell anti-body-forming plasma cells from bone marrow cultures of syngeneic and allogeneic mice: functional modulation of helper T-cell subsets in autosensitization. Immunology. Mar. 1983;48(3):579-86.

Nakamura, K. et al., Itranuclear incorporation of thymic low molecular weight RNA by murine bone marrow immunoblasts and inhibition of plasma cell formation by a derivative of rifampicin. Microbiol Immunol. 1982;26(1):41-57.

Nakamura, K. et al., Mechanism of anti-DNA antibody formation. The functional modulation of helper T-subset plays the key role in both murine and human B-cell autosensitization. Microbiol Immunol. 1986;30(7):703-15.

Ponsaerts, P. et al., Messenger RNA electroporation of human monocytes, followed by rapid in vitro differentiation, leads to highly stimulatory antigen-loaded mature dendritic cells. J Immunol. Aug. 15, 2002;169(4):1669-75.

Porgador, A. et al., Induction of antitumor imminity using bone marrow-generated dendritic cells. J Immunol. Apr. 15, 1996;156(8):2918-26.

Pradilla, G. et al., Prevention of vasospasm following subarachnoid hemorrhage in rabbits by anti-CD11/CD18 monoclonal antibody therapy. J Neurosurg. Jul. 2004;101(1):88-92.

OTHER PUBLICATIONS

Preisler, H.D. et al., Sensitization in vitro to murine myeloblastic leukemia cells by xenogeneic immune RNA. J Natl Cancer Inst. Jan. 1979;62(1):133-7.

Preiss, T. et al., Dual function of the messenger RNA cap structure in poly(A)-tall-promoted translation in yeast. Nature. Apr. 2, 1998;392(6675):516-20.

Probst, J., et al., Spontaneous cellular uptake to exogenous messenger RNA in vivo is nucleic acid-specific, saturable and ion dependent. Gene Therapy. 2007; 14: 1175-1180.

Puga, A., et al., Diference between functional and structural integrity of messenger RNA. Proc Natl Acad Sci U S A. Jul. 1973;70(7):2171-5.

Pulford, B., et al., Liposome-siRNA-peptide complexes cross the

blood-brain barrier and significantly decrease PrP[^]C on neuronal cells and PrPλRES in infected cell cultures. PLoS ONE. 2010; 5(6): e11085.

Purchio, A.F. et al., [24] Methods for molecular cloning in eukaryotic cells. Methods Enzymol. 1979; 68:357-75.

Query, C.C. et al., Branch nucleophile selection in pre-mRNA splicing; evidence for the bulged duplex model. Genes Dev. Mar. 1, 1994;8(85):587-97.

Rabinovich, P.M., et al., Synthetic messenger RNA as a tool for gene therapy. Hum. Gen Ther. Oct. 2006; 17:1027-1035.

Rabinovich, P.M., et al., Chemeric receptor mRNA transfection as a tool to generate Antineoplastic Lymphocytes. Hum. Gene Ther. Jan. 2009; 20: 51-61.

Raff, M., Adult stem cell plasticity: fact or artifact? Annu Rev Cell Biol. 2003;19:1-22.

Rajagopalan, L.E. et al., Turnover and translation of in vitro synthesized messenger RNAs in transfected, normal cells. J Biol Chem. Aug. 16, 1996;271(33):19871-6.

Ramazeilles, C. et al., Antisense phoaphorothioale oligonucleotides: selective killing of the intracellular parasite Leishmania amazonensis. Proc Natl Acad Sci U S A. Aug. 16, 1994;91(17):7859-63.

Rammensee, H.G. et al., Peptides naturally presented by MHC class I molecules. Annu Rev Immunol. 1993;11:213-44.

Rascati, R.J. et al., Characterization of Fv-1 gene-produce-mediated resistance transfer. Intervirology. 1981;15(2):87-96.

Ratajczak, J. et al., Embryonic stem cell-derived microvesicles reprogram hematopoletic progenitors: evidence for horizontal transfer of mRNA and protein delivery. Leukemia. May 2006;20(5):847-56

Ratajczak, J. et al., Membrane-derived microvesicles: important and underappreciated mediators of cell-to-cell communication. Leukemia. Sep. 2006;20(9):1487-95. Epub Jul. 20, 2006.

Read, M.L., et al., A versatile reducible polycation-based system for efficient delivery of a broad range of nucleic acids. Nucleic Acids Res. 2005; 33(9): e86.

Reddy, A. et al., The effect of labour and placental separation on the shedding of syncytiotrophoblast microparticles, cell-free DNA and mRNA in normal pregnancy and pre-eclampsia. Placenta. Nov. 2008;29(11):942-9. Epub Oct. 1, 2008.

Reed, R. et al., Intron sequences involved in lariat formation during pre-mRNA splicing. Cell. May 1985;41(1):95-105.

Regnier, P. et al., Degradation of mRNA in bacteria; emergence of ubiquitous features. Bioessays. Mar. 2000;22(3):235-44.

Rejman, J., et al., mRNA transfection of cervical carcinoma and mesenchymal stem cells mediated by cationic carriers. J Controlled Rel. Nov. 2010; 147(3): 385-391.

Renkvist, N. et al., A listing of human tumor antigens recognized by T cells. Cancer Immunol Immunother. Mar. 2001;50(1);3-15.

Reyes-Sandoval, A. et al., DNA Vaccines. Curr Mol Med. May 2001;1(2):217-43.

Reynolds, B.A. et al., Generation of neurons and astrocytes from isolated cells of the adult mammalian central nervous system. Science. Mar. 27, 1992;255(5052):1707-10.

Ruhnke, M. et al., Long-term culture and differentiation of rat embryonic stem cell-like cells into neuronal, glial, endothelial, and hepatic lineages. Stem Cells. 2003;21(4):428-36.

Richter, J.D., Cytoplasmic polyadenylation in development and beyond. Microbiol Mol Biol Rev. Jun. 1999;63(2):446-56.

Roberts, J.N. et al., Genital transmission of HPV in a mouse model is potentiated by nonoxynol-9 and inhibited by carrageenan. Nat Med. Jul. 2007; 13(7): 857-861.

Robbins, P.F. et al., Human tumor antigens recognized by T cells. Curr Opin Immunol. Oct. 1996;8(5):628-36.

Robinson, F. et al., Expression of human nPTB is limited by extreme suboptimal codon content. PLoS One, Mar. 12, 2008;3(3):e1801.

Robinson, H.L. et al., Protection against a lethal influenza virus challenge by immunization with a heamagglutinin-expressing plasmid DNA. Vaccine. 1993;11(9):957-60.

Robles, A.I. et al., Reduced skin tumor development in cyclin D1-dificient mice highlights the oncogenic ras pathway in vivo. Genes Dev. Aug. 15, 1998;12(16):2469-74.

Rock, K.L. et al., A new foreign policy: MHC class I molecules monitor the outside world. Immunol Today. Mar. 1996;17(3):131-7. Rodriguez, P.L., et al., Minimal "self" peptides that inhibit phagocytic clearance and enhance delivery of nanoparticles. Science. Feb. 22, 2013; 339(6122): 971-975.

Rohloff, C.M., et al., DUROS® Technology delivers peptides and proteins at consistent rate continuously for 3 to 12 months. J Diabetes Sci Technol. May 2008; 2(3): 461-467.

Romani, N. et al., Generation of mature dendritic cells from human blood. An improved method with special regard to clinical applicability. J Immunol Methods. Sep. 27, 1996;196(2):137-51.

Romani, N. et al., Presentation of exogenous protein antigens by dendritic cells to T cell clones. Intact protein is presented best by immature, epidermal Langerhans cells. J Exp Med. Mar. 1, 1989;189(3):1169-78.

Rosa, A., et al., Synthetic mRNAs: Powerful tools for reprogramming and differentiation of human cells. Cell Stem Cell. Nov. 2010; 7: 540-550.

Rosenberg, S.A. et al., Cancer immunotherapy: moving beyond current vaccines. Nat Med. Sep. 2004;10(9):909-15.

Ross, B.S. et al., Synthesis and incorporation of 2'-O-methyl-pseudouridine into oligonucleotides. Nucleosides and Nucleotides. 1997; 16(7/9):1547-9.

Ross, J. Control of messenger RNA stability in higher eukaryotes. Trends Genet. May 1996;12(5):171-5.

Rossi, Derrick, Open letter Entitled "Change to mRNA Reprogramming Protocol" Publication Date: Aug. 13, 2011 (available at Addgene website: http://www.addgene.org/static/data/83/87/3686c0f2-c9a2-11e0-b8a9-003048dd6500.pdf, last retrieved Mar. 17, 2013).

Ryser, M., et al., S1P1 overexpression stimulates S1P-dependent chemotaxis of human CD34+ hematopoietic progenitor cells but strongly inhibits SDF-1/CXCR4-dependent migration and in vivo homing. Mol Immunology. 2008; 46: 166-171.

Saenz-Badillos, J. et al., RNA as a tumor vaccine: a review of the literature. Exp Dermatol. Jun. 2001;10(3):143-54.

Saison-Behmoaras, T. et al., Short modified antisense oligonucleotides directed against Ha-ras point mutation induce selective cleavage of the mRNA and inhibit T24 cells proliferation. EMBO J. May 1991;10(5):1111-8.

Saito, K. et al., Cell participation in immune response by immune ribonucleic acid. I. The role of T lymphocytes in immune response by immune RNA against T-depedent antigens. Immunology. Dec. 1980;41(4):937-45.

Saito, R., et al., Distribution of liposomes into brain and rat brain tumor models by convetion-enhanced delivery monitored with magnetic resonance imaging. Cancer Res. Apr. 2004; 64: 2572-2579

Sakuma, S. et al., Mucaodhesion of polystyrene nanoparticles having surface hydrophilic polymeric chains in the gastrointestinal tract. Int J Pharm. Jan. 25, 1999;177(2):161-72.

Sallusto, F. et al., Dendritic cells use macropinocytosis and the mannose receptor to concentrate macromolecules in the major

OTHER PUBLICATIONS

histocompability complex class II compartment: downregulation by cytokines and bacterial products. J Exp Med. Aug. 1, 1995;182(2):389-400.

Sallusto, F. et al., Efficient presentation of soluble antigen by cultured human dendritic cells is maintained by granulocyte/macrophage colony-stimulating factor plus interleukin 4 and downregulated by tumor necrosis factor alpha. J Exp Med. Apr. 1, 1994;179(4):1109-18.

Veres, G., et al., The molecular basis of the sparse fur mouse mutation. Science. Jul. 1987;237(4813):415-7.

Verheggen, C. et al., Box C/D small nucleolar RNA trafficking involves small nucleolar RNP proteins, nucleolar factors and a novel nuclear domain. EMBO J. Oct. 1, 2001;20(19):5480-90.

Verheggen, C. et al., Mammalian and yeast U3 snoRNPs are matured in specific and related nuclear compartments. EMBO J. Jun. 3, 2002;21(11):2736-45.

Verma, I.M. et al., Gene therapy: promises, problems and prospects. Nature. Sep. 18, 1997;389(6648):239-42.

Verma, I.M. et al., Gene therapy: twenty-first century medicine. Annu Rev Biochem. 2005;74:711-38.

Verma, S. et al., Modified oligonucleotides: synthesis and strategy for users. Annu Rev Biochem. 1998;67:99-134.

Vilee, D.B., Ribonucleic acid; control of steroid synthesis in endocrine tissue. Science. Nov. 3, 1967;158(3801):652-3.

Villaret, D.B. et al., Identification of genes overexpressed in heat and neck squamous cell carcinoma using a combination of complementary DNA subtraction and microarray analysis. Laryngoscope. Mar. 2000;110(3 Pt 1):374-81.

Virovic, L. et al., Novel delivery methods for treatment of viral hepatitis: an update. Exppert Opin Drug Deliv. Jul. 2005;2(4):707-17

Viza, D. et al., Human lymphoblastoid cells in culture replicate immune information carried by xenogeneic RNA. Differentiation. 1978;11(3):181-4.

Wagner, E. Polymers for siRNA delivery: Inspired by viruses to be targeted, dynamic, and precise. Acc Chem Res. 2012; 45(7): 1005-1013

Wahle, E. Poly(A) tail length control is caused by termination of processive synthesis. J Biol Chem. Feb. 10, 1995; 270(6): 2800-2808.

Wang, B. et al., Gene inoculation generates immune responses against human immunodeficiency virus type 1. Proc Natl Acad Sci U S A. May 1, 1993;90(9):4156-60.

Wang, B. et al., Immunization by direct DNA inoculatuion induces rejection of tumor cell challenge. Hum Gene Ther. Apr. 1995;6(4):407-18.

Wang, B.S. et al., Fractionation of immune RNA capable of transferring tumor-specific cellular cytotoxicity. Cell Immunol. May 1978;37(2):358-68.

Wang, S.P. et al., Phylogeny of mRNA capping enzymes. Proc Natl Acad Sci U S A. Sep. 2, 1997;94(18):9573-8.

Wang, Y., et al., Systemic delivery of modified mRNA enoxiing herpes simplex virus 1 thymidine kinase for targeted cancer gene therapy. Mol Therapy. 2012; 11: 1-10.

Warren, T.L. et al., Uses of granulocyte-macrophage colony-stimulating factor in vaccine development. Curr Opin Hematol. May 2000;7(3):168-73.

Weaver, J.C., Electroporation theory. Concepts and mechanisms. Methods Mol Biol. 1995;55:3-28.

Watanabe, T. et al., Induction of wild-type p53 activity in human cancer cells by ribozymes that repair mutant p53 transcripts. Proc Natl Acad Sci U S A. Jul. 18, 2000;97(15):8490-4.

Weber, J. et al., Granulocyte-macrophage-colony-stimulating factor added to a multipeptide vaccine for resected Stage II melanoma. Cancer. Jan. 1, 2003;97(1):186-200.

Weide, B. et al., Results of the first phase I/II clinical vaccination trial with direct injection of mRNA. J Immunother. Feb.-Mar. 2008;31(2):180-8.

Weide, B., et al., Direct injection of protamine-protected mRNA: Results of a phase 1/2 vaccination trial in metastatic melanoma patients. J. of Immunotherapy. Jun. 2009; 32(5): 498-507.

Nakamura, O. et al., Abstract: The Role of Immune RNA in the Immunotherapy of Malignant Brain Tumor. 1982, 34(2);333-9.

Weisberger, A.S., Induction of altered globin synthesis in human immature erythrocytes incubated with ribonucleoprotein. Proc Natl Acad Sci USA. Jan. 1962; 48(1): 68-80.

Weiss, S.B. et al., Pseudouridine Formation: Evidence for RNA as an Intermediate. Science. Jul. 23, 1965: 149(3682): 429-431.

Weissman, D. et al., Dendritic cells express and use multiple HIV coreceptors. Adv Exp Med Biol. 1997;417:401-6.

Weissman, D. et al., HIV GAG mRNA transfection of dendritic cell (DC) delivers encoded antigen to MHC class I and II molecules, causes DC maturation, and induced a potent human in vitro primary immune response. J Immunol. Oct. 15, 2000;165(8):4710-7.

Wels, W., et al., Construction, bacterial expression and characterization of a bifunctional single-chain antibody-phosphate fusion protein targeted to the human erbb-2 receptor. Biotechnology (NY). Oct. 1992; 10(10): 1128-1132.

Wickens, M. et al., A PUF family portraint: 3'UTR regulation as a way of life. Trends Genet. Mar. 2002;18(3):150-7.

Wiehe, J.M. et al., mRNA-mediated gene delivery into human progenitor cells promotes highly efficient protein expression. J Cell Mol Med. May-Jun. 2007;11(3):521-30.

Wilkie, G.S. et al., Regulation of mRNA translation by 5'- and 3'-UTR-binding factors. Trends Biochem Sci. Apr. 2003;28(4):182-8

Wilusz, C.J. et al., Bringing the role of mRNA decay in the control of gene expression into focus. Trends Genet. Oct. 2004;20(10):491-7

Wilusz, J. et al., A B4 kd nuclear protein binds to RNA segments that include the AAUAAA polyadenylation motif. Cell. Jan. 29, 1988;52(2):221-8.

Winnicka, B, et al., CD13 is dispensable for normal hematopoiesis and myeloid cell functions in the mouse. J Leukoc Biol. Aug. 2010: 88(2); 347-359. Epub Apr. 29, 2010.

Wolff, J.A. et al., Direct gene transfer into mouse muscle in vivo. Science. Mar. 23, 1990;247(4949 Pt 1):1465-8.

Woltjen, K. et al., piggyBac transposition reprograms fibroblasts to induced pluripotent stem cells. Nature. Apr. 2009 (458): 10.1038-07863.

Woodberry, T. et al., Immunogenicity of a human immunodeficiency virus (HIV) polytope vaccine containing multiple HLA A2 HIG CD8(+) cytotoxic T-cell epitopes. J Virol. Jul. 1999;73(7):5320-5.

Wu, J. et al., Mammalian pre-mRNA branch site selection by U2 snRNP involves base pairing. Genes Dev. Oct. 1989;3(10):1553-61. Wu, L. et al., Fusion protein vectors to increase protein production and evaluate the immunogenicity of genetic vaccines. Mol Ther. Sep. 2000;2(3):288-97.

Wu, X.C. et al., Engineering a Bacillus subtilis expression-secretion system with a strain deficient in six extracellular proteases. J Bacteriol. Aug. 1991;173(16):4952-8.

Wurm, F. et al., Suppression of melanoma development and regression of melanoma in xiphophorine fish after treatment with immune RNA. Cancer Res. Sep. 1981;41(9 Pt 1):3377-83.

Wyatt, J.R. et al., Site-specific cross-linking of mammalian U5 snRNP to the 5' splice site before the first step of pre-mRNA splicing. Genes Dev. Dec. 1992;6(12B):2542-53.

Xu, C. et al., Feeder-free growth of undifferentiated human embryonic stem cells. Nat Biotechnol. Oct. 2001;19(10):971-4.

Xu, J. et al., Identification of differentially expressed genes in human prostate cancer using subtraction and microarray. Cancer Res. Mar. 15, 2000;60(6):1677-82.

Yamamoto, A., et al., Current prospects for mRNA gene delivery. Eur J Pharm Biopharm. Mar. 2009; 71(3): 484-489.

Yamashita, A. et al., Concerted action of poly(A) nucleases and decapping enzyme in mammalian mRNA turnover. Nat Struct Mol Biol. Dec. 2005;12(12):1054-63. Epub Nov. 13, 2005.

Yang, S.F. et al., Albumin synthesis in mouse uterus in response to liver mRNA. Proc Natl Acad Sci U S A. May 1977;74(5):1894-8.

OTHER PUBLICATIONS

Ye, X., et al., Prolonged metabolic correction in adult ornithine transcarbamylase-deficient mice with adenoviral vectors. Biological Chem. Feb. 1996; 271(7): 3639-3646.

Yi, Y., et al., Current advances in retroviral gene therapy. Current Gene Ther. 2011; 11: 218-228.

Ying, H. et al., Cancer therapy using a self-replicating RNA vaccine. Nat Med. Jul. 1999;5(7):823-7.

Yisraeli, J.K. et al., [4] Synthesis of long, capped transcripts in vitro by SP6 and T7 RNA Polymerases. Methods in Enzymology, vol. 180. 1989; 180, 42-50.

Yokoe, H. et al., Spatial dynamics of GFP-tagged proteins investigated by local fluorescence enhancement. Nat Biotechnol. Oct. 1996;14(10):1252-6.

Yoshida, Y. et al., Hypoxia enhances the generation of induced pluripotent stem cells. Cell Stem Cells 5, Sep. 2009; 5: 237-241. You, Z. et al., A retrogen strategy for presentation of an intracellular tumor antigen as an exogenous antigen by dendritic cells induces potent antitumor T helper and CTL responses. Cancer Res. Jan. 1, 2001;61(1):197-205.

Yu, J. et al., Structural and functional analysis of an mRNP complex that mediates the high stability of human beta-globin mRNA. Mol Cell Biol. Sep. 2001;21(17):5879-88.

Yu, J. et al., Induced pluripotent stem cell lines derived from human somatic cells. Science. Dec. 21, 2007; 318(5856): 1917-1920.

Yu, J. et al., Human induced pluripotent stem cells of vector and transgene sequences. Science. May 8, 2009; 324(5938): 797-801.

Yu, P.W. et al., Sustained correction of B-cell development and function in a murine model of X-linked agammaglobulinemia (XLA) using retroviral-mediated gene transfer. Sep. 1, 2004;104(5):1281-90. Epub May 13, 2004.

Yu, Y.T. et al., Internal modification of U2 small nuclear (sn)RNA occurs in nucleoli of Xenopus oocytes. J Cell Biol. Mar. 19, 2001;152(6):1279-88.

Yu, Y.T. et al., Modifications of U2 snRNA are required for snRNP assembly and pre-mRNA splicing. EMBO J. Oct. 1, 1998;17(19):5783-95.

Zebarjadian, Y. et al., Point mutations in yeast CBF5 can abolish in vivo pseudouridylation of rRNA. Mol Cell Biol. Nov. 1999;19(11):7461-72.

Zeitlin, S. et al., In vivo splicing products of the rabbit beta-globin pre-mRNA. Cell. Dec. 1984;39(3 Pt 2):589-602.

Zelcer, A. et al., The detection and characterization of viral-related double-stranded RNAs in tobacco mosaic virus-infected plants. Virology. Sep. 1981;113(2):417-27.

Zeytin, H.E. et al., Construction and characterization of DNA vaccines encoding the single-chain variable fragment of the anti-idiotype antibody 1A7 mimicking the tumor-associated antigen disialoganglioside GD2. Cancer Gene Ther. Nov. 2000;7(11):1426-36

Zhang, X. et al., Advances in dendritic cell-based vaccine of cancer. Cancer Biother Radiopharm. Dec. 2002;17(6):601-19.

Zhang, Y., et al., In vivo gene delivery by nonviral vectors: overcoming hurdles? Mol. Therapy. Jul. 2012; 20(7): 1298-1304. Zhao, X. et al., Pseudouridines in and near the branch site recognition region of U2 snRNA are required for snRNP biogenesis and pre-mRNA splicing in Xenopus oocytes. RNA. Apr. 2004;10(4):681-90.

Zhigaltsev, I.V., et al., Bottom-Up design and synthesis of limit size lipid nanoparticle systems with aqueous and triglyceride cores using millisecond microfluidic mixing. Langmuir. Feb. 21, 2012; 28(7): 3633-3640.

Zhou, W.Z. et al., RNA melanoma vaccine: induction of antitumor immunity by human glycoprotein 100 mRNA immunization. Hum Gene Ther. Nov. 1, 1999;10(16):2719-24.

Zhou, H., et al., Generation of induced pluripotent stem cells using recombinant proteins. Cell Stem Cell. May 4, 2009(5):381-4.

Zhou, J., et al., Short Communication Bilirubin Glucuronidation Revisited: Proper assay conditions to estimate enzyme kinetics with recombinant UGT1A1. Drug metabolism and Disp. 2010; 38(11): 1907-1911.

Zhuang, Y. et al., A compensatory base change in human U2 snRNA can suppress a branch site mutation. Genes Dev. Oct. 1989;3(10):1545-52.

Zimmermann, E. et al., Electrolyte- and pH-stabilities of aqueous solid lipid nanoparticle (SLNTM) dispersions in artificial gastrointestinal media. Eur J Pharm Biopharm. Sep. 2001;52(2):203-10.

Zitvogel, L. et al., Therapy of murine tumors with tumor peptidepulsed dendritic cells: dependence on T cells, B7 costimulation, and T helper cell 1-associated cytokines. J Exp Med. Jan. 1, 1996;183(1);87-97.

Zohra, F.T., et al., Drastic effect of nanoapatite particles on liposome-mediated mRNA delivery to mammalian cells. Analytical Biochem. Oct. 2005; 345(1): 164-166.

Zohra, F.T., et al., Effective delivery with enhanced translational activity synergistically accelerates mRNA-based transfection. Biochem Biophys Res Comm. Jun. 2007; 358(1): 373-378.

Zonta, S. et al., Uretero-neocystostomy in a swine model of kidney transplantation: a new technique. J Surg Res. Apr. 2005;124(2):250-5

Zorio, D.A. et al., Both subunits of U2AF recognize the 3' splice site in Caenorhabditis elegans . Nature. Dec. 16, 1999;402(6763):835-8. Chang, N. et al., Genome editing with RNA-guided Cas9 nuclease in Zebrafish embryos. Cell Res. Apr. 23, 2013(4):465-472.

Cong, L. et al., Multiplex genome engineering using CRISPR/Cas systems. Science. Feb. 15, 2013; 339(6121): 819-823.

Jinek, M. et al., A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. Science. Aug. 17, 2012; 337(6096): 816-821.

Jinek, M. et al., RNA-programmed genome editing in human cells. Elife. 2013;2:e00471.

Maehr, R. et al., Generation of pluripotent stem cells from patients with type 1 diabetes. Proc Natl Acad Sci USA. Sep. 15, 2009; 106(37): 15768-15773.

Mali, P. et al., RNA-guided human genome engineering via Cas9. Science. Feb. 15, 2013; 339(6121): 823-826.

Qi, L.S. et al., Repurposing CRISPR as an RNA-guided platform for sequence-specific control of gene expression. Cell. Feb. 28, 2013: 152(5): 1173-1183.

Shen, B. et al., Generation of gene-modified mice via Cas9/RNA-mediated gene targeting. Cell Res. Apr. 2, 2013; 1-4.

International Search Report from International Application No. PCT/US10/059317 dated Aug. 22, 2011.

International Search Report from International Application No. PCT/US10/059305 dated Aug. 23, 2011.

Yi, P. et al., Betatrophin: A hormone that control pancreatic beta cell proliferation. Cell. May 9, 2013, 153: 1-12.

Graf, T and Enver T. Forcing cells to change lineages. Nature. Dec. 3, 2009; 462(7273): 587-594.

Ieda, M. et al., Direct reprogramming of fibroblasts into functional cardiomyocytes by defined factors. Cell. Aug. 6, 2010; 142(3);

Huangfu, D. et al., Induction of pluripotent stem cells from primary human fibroblasts with only Oct4 and Sox2. Nat Biotechnol. Nov. 2008; 26(11): 1269-1275.

Dong, X.Y. et al., Identification of genes differentially expressed in human hepatocellular carcinoma by a modified suppression subtractive hybridization method. Int J Cancer. Nov. 1, 2004; 112(2): 239-248.

Okita, K. et al., Generation of mouse induced pluripotent stem cells without viral vectors. Science. 2008; 322: 949-953.

Stadtfeld, M. et al., Induced pluripotent stem cells generated without viral integration. Science. Nov. 7, 2008; 322(5903): 945-949. Aoi, T. et al., Generation of pluripotent stem cells from adult mouse liver and stomach cells. Science. Aug. 1, 2008; 321 (6889): 699-

Feng, R. et al., PU.1 and C/EBPalpha/beta convert fibroblasts into macrophage-like cells. Proc Natl Acad Sci USA. Apr. 22, 2008; 105(16): 6057-6062.

OTHER PUBLICATIONS

Szabo, E. et al., Direct conversion of human fibroblasts to multilineage blood progenitors Nature. Nov. 25, 2010; 468(7323): 521-526.

Gonzalez, F. et al., Generation of mouse-induced pluripotent stem cells by transient expression of a single nonviral polycistronic vector. Proc Natl Acad Sci USA. Jun. 2, 2009; 106(22): 8918-8922. Aasen, T. et al., Efficient and rapid generation of induced pluripotent stem cells from human keratinocytes. Nat Biotechnol. Nov. 2008: 26(11): 1276-1284.

Ebert, A.D. et al., Induced pluripotent stem cells from a spinal muscular atrophy patient. Nature. Jan. 15, 2009; 457 (7227): 277-280.o.

Vierbuchen, T. et al., Direct conversion of fibroblasts to functional neurons by defined factors. Nature. Feb. 25, 2010; 463(7284): 1035-1041.

Racila, D. et al., Transient expression of OCT4 is sufficient to allow human keratinocytes to change their differentiation pathway. Gene Ther. Mar. 2011; 18(3): 294-303.

Nakagawa, M. et al., Generation of induced pluripotent stem cells without Myc from mouse and human fibroblasts. Nat Biotechnol. Jan. 2008: 26(1); 101-106. Epub Nov. 30, 2007.

Haft, D.H. et al., A guild of 45 CRISPR-associated (Cas) protein families and multiple CRISPR/Cas subtypes exist in prokaryotic genomes. PLoS Comput Biol. Nov. 2005; 1(6): e60. Epub Nov. 11, 2005.

Brown, C.E., et al., Poly(A) Tail Lengeth Control in *Saccharomyces cerevisiae* Occurs by Message-Specific Deadenylation. Molecular and Cellular Biology. Nov. 1998 p. 6548-6569.

Gao, G., et al., Erythropoietin gene therapy leads to autoimmune anemia in macaques. 2004 103: 3300-3302.

Liu, C., et al., Peptidoglycan Recognition Proteins. A Novel Family of Four Human Innate Immunity Pattern Recognition Molecules. The Journal of Biological Chemistry. vol. 276, No. 37, Issue of Sep. 14, pp. 686-34694, 2001.

Lu, X., Peptidoglycan Recognition Proteins are a New Class of Human Bactericidal Proteins. The Journal of Biological Chemistry. Mar. 3, 2006, vol. 281, No. 9, pp. 5895-5907.

Ngai, P.H.K., et al., Agrocybin, an antifungal peptide from the edible mushroom. Department of Biochemistry, The Chinese University of Hong Kong. Peptides 26 (2005) 191-196.

Endo, F., et al., A Nonsense Mutation in the 4-Hydroxyphenylpyruvic Acid Dioxygenase Gene (Hpd) Causes Skipping of the Constitutive Exon and Hypertyrosinemia in Mouse Strain III. Genomics 25, 164-169 (1995).

Neve, S., et al. Tissue distribution, intracellular localization and proteolytic processing of rat 4-hydorxyphenylpyruvate dioxygenase. Cell Biology International 27 (2003) pp. 611-624.

Ren, W., et al. Molecular clong and characterization of 4-hydroxyphenylpyruvate dioxygenase gene from Lactuca sativa. Journal of Patent Physiology 168 (2011 pp. 1076-1083).

Ruetschu, U., et al. Human 4-Hydroxyphenylpyruvate Dioxygenase Gene (HPD). Genomics 44, pp. 292-299 (1997).

Seabury, C.M., et al., Analysis of sequence variability and protein domain architectures for bovine peptidoglycan recognition protein 1 and Toll-like receptors 2 and 6. Genomics 92 (2008) pp. 235-245. Sumathipala, N. et al., Involvement of Manduca sexta peptidoglycan recognition protein-1 in the recognition of bacteria and activation of prophenoloxidase system. Insect Biochemistry and Molecular Biology 40 (2010) 487-495.

Wei, X. et al., Molecular cloning and MRNA expression of two peptidogluycan recognition protein (PGRP genes from mollusk Solen grandis. Fish & Shellfish Immunology 32 (2012) 178-185. Anonymous: "Messenger RNA". Internet: Wikipedia. Jun. 19, 2013, XP002699196, Retrieved from the Internet: URL: http://en.wikipedia.org/wiki/Messenger RNA.

Grosjean, H., DNA and RNA Modification Enzymes Structure, Mechanisms, Functions and Evolution. Molecular Biology Intelligence Unit. Estimated Publication Date: May 2009. pp. 1-2.

Grosjean, H., Nucleic Acids are not Boring Long Polymers of Only Four Types of Nucleotides: A Guided Tour. Chapter 1. Landes Bioscience. 2009. pp. 1-18.

Grosjean, H., et al. How Nucleic Acids Cope with High Temperature. Physiology and Biochemistry of Extremophiles. 2007. Chapter 4, pp. 39-58.

Grosjean, H., Modification and editing of RNA: historical overview and important facts to remember. pp. 1-22.

Hunt, D.M., et al., The L Protein of Vesicular Stomatitis Cirus Modulates the Response of the Polyadenylic Acid Polymerase to S-Adenosylhomocysteine. J. gen. Virol. (1988), 69, 2555-2561.

Grosjean, H., et al. Fine-Tuning of RNA Functions by Modification and Editing. Topics in Current Genetics, vol. 12, 2005, XXIV, p. 442.

Bouloy, M., et al., Both the 7-methyl and the 2'-O-methyl groups in the cap of mRNA strongly influence its ability to act as primer for influenza virus RNA transcription. Proc. Natl. Acad. Sci. USA, vol. 77, No. 7, pp. 3952-3956, Jul. 1980.

Fernandez, I., et al. Unusual base pairing during the decoding of a stop codon by the ribosome. vol. 000, 2013. pp. 1-5.

Edelheit, S. et al., Transcriptome-Wide Mapping of 5-methylcytidine RNA Modifications in Bacteria, Archaea, and Yeast Reveals m5C within Archaeal mRNAs. PLOS Genetics, Jun. 2013, vol. 9, Issue 6, pp. 1-14.

Cun, Dongmei, et al., Preparation and characterization of poly(DL-lactide-co-glycolide) nanoparticles for siRNA delivery. International Journal of Pharmaceutics 390 (2010) 70-75.

Oster, C.G., et al., Comparative study of DNA encapsulation into PLGA microparticles using modified double emulsion methods and spray drying techniques. Journal of Microencapsulation, May 2005; 22(3): 235-244.

GenBank: Homo sapiens 15 kDA selenoprotein (SEP 15), transcript variant 1, mRNA, NCBI Reference sequence: NM_004261.3, pp. 1-4.

Thomson A. James., et al. Isolation of a primate embryonic stem cell line. vol. 92, pp. 7844-7848, Aug. 1995. Proc. Natl. Acad. Sci. USA. Tahiliani., et al.Conversion of 5-Methylcytosine to 5-Hydroxymethylcytosine in Mammalian DNA by MLL Partner TET1 Science 323, 930 (2009);www.sciencemag.org.

The Human Embryonic Stem Cell and the Human Embryonic Germ Cell. NIH Stem Cells: Scientific Progress and Future Research Directions, Department of Health and Human Services, Chapter 3, Jun. 2001.

The Stem Cell. NIH Stem Cells: Scientific Progress and Future Research Directions, Department of Health and Human Services, Chapter 1, Jun. 2001.

Morgan, D. Hugh, et al. Molecular Basis of Cell and Developmental Biology: Activation-induced Cytidine Dreaminase Deaminates 5-Methylcytosine in DNA and is Expressed in Pluripotent Tissues: Implications for Epigenetic Reprogramming. J. Biol. Chem. 2004, 279:52353-52360 published online Sep. 24, 2004.

Moore, J.E., et al., The Corneal Epithelial Stem Cell. vol. 21, Nos. 5/6, 2002. Mary Ann Liebert, Inc. pp. 443-451.

Koh, Peng Kian, et.al. Tet1 and Tet2 Regulate 5-Hydroxymethylcytosine Production and Cell Lineage Specification in Mouse Embryonic Stem Cells. 200-213, Feb. 4, 2011 2011 Elsevier Inc.

Kariko, Katalin, et.al. Naturally occurring nucleoside modifications suppress the immunostimulatory activity of RNA: Implication for therapeutic RNA development. Current Opinion in Drug Discovery & Development 2007 10(5): 523-532 The Thomson Corporation ISSN 1367-6733.

Ito, Shinsuke, et.al. Role of Tet proteins in 5mC to 5hmC conversion, ES-cell self-renewal and inner cell mass specification. vol. 466[Aug. 26, 2010] Macmillan Publishers Limited. pp. 1129-1133. Freudenberg, M. Johannes, et.al. Acute depletion of Tet1-dependent 5-hydroxymethylcytosine levels impairs LIF/Stat3 signaling and results in loss of embryonic stem cell identity. Published online Dec. 30, 2011. 3364-3377 Nucleic Acids Research, 2012, vol. 40, No. 8. Published by Oxford University Press 2011.

OTHER PUBLICATIONS

Ficz, Gabriella, et al. Dynamic regulation of 5-hydroxymethylcytosine in mouse ES cells and during differentiation. Nature | vol. 473 | May 19, 2011. pp. 398-401. Macmillian Publishers

Blelloch, Robert, et al. Generation of Induced Pluripotent Stem Cells in the Absence of Drug Selection. Sep. 13, 2007. pp. 245-247. Verma, Sandeep et.al. Modified Oligonucleotides: Synthesis and Strategy for Users. Biochem. 1998. 67:99-134. 1998 by Annual Reviews

Leung W. David. The Structure and Functions of Human Lysophosphatidic Acid Acyltransferases. Frontiers in Bioscience 6. pp. 944-953, Aug. 1, 2001.

Lu, Biao, et al. Cloning and characterization of murine 1-acyl-snglycerol 3-phosphate acyltransferases and their regulation by PPAR in murine heart. Biochem J. (2005) 385, 469-477 (printed in Great Britain).

West, James, et.al. Cloning and Expression of Two Human Lysophosphatidic Acid Acyltransferase cDNAs That Enhance Cytokine-Induced Signialing Responses in Cells. DNA and Cell Biology vol. 16, No. 6, 1997. Mary Ann Liebert, Inc. pp. 691-791. Bionaz, Massimo, et.al. ACSL1, AGPAT6, FABP3, LPIN1, and SLC27A6 are the Most Abundant Isoforms in Bovine Mammary Tissue and Their Expression is affected by Stage of Lactation. The Journal of Nutrition, 2008. pp. 1019-2024.

Sterner, D.E. et al., Acetylation of histones and transcription-related factors. Microbiol Mol Biol Rev. Jun. 2000;64(2):435-59.

Stiles, D.K., et al., Widespread suppression of huntingtin with convection-enhanced delivery of siRNA. Experimental Neurology. Jan. 2012; 233(1): 463-471.

Stinchcomb, D.T. et al., Isolation and characterisation of a yeast chromosomal replicator. Nature. Nov. 1, 1979;282(5734):39-43. Strong, V.T. et al., Incorporation of beta-globin untranslated regions into a Sindbis virus vector for augmentation of heterologous mRNA expression. Gene Ther. Jun. 1997;4(6):624-7.

Studier, F.W. et al., Use of bacteriophage T7 RNA polymerase to direct selective high-level expression of cloned genes. J Mol Biol. May 5, 1986;189(1):113-30.

Studier, F.W. et al., [6] Use of T7 RNA polymerase to direct expression of cloned genes. Methods Enzymol. 1990;185-60-89. Su, Z. et al., Enhanced induction of telomerase-specific CD4(+) T Cells using dendritic cells transfected with RNA encoding a chimeric gene product. Cancer Res. Sep. 1, 2002;62(17):5041-8.

Su, Z. et al., Immunological and clinical responses in metastatic renal cancer ptaients vaccinated with tumor RNA-transfected dendritic cells. Cancer Res. May 1, 2003;63(9):2127-33.

Suda, T. et al., Hydrodynamic gene delivery: its principles and applications. Mol Ther. Dec. 2007;15(12):2063-9. Epub Oct. 2, 2007

Sullenger, B.A. et al., Emerging clinical applications of RNA. Nature. Jul. 11, 2002;418(6894):252-8.

Svinarchuk, F.P. et al., Inhibition of HIV proliferation in MT-4 cells by antisense oligonucleotide conjugated to lipophilic groups. Biochimie. 1993;75(1-2):49-54.

Takahashi, K., et al., Induction of pluriopotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell. Aug. 2006; 126(4): 663-76.

Takahashi, K., et al., Induction of pluripotent stem cells from adult human fibroblasts by defined factors. Cell. Nov. 2007; 131(5): 861-72.

Tam, C., et al., Cytokeratins mediate epithelial innate defense through their antimicrobial properties. J Clin Invest. Oct. 1, 2012; 122(10): 3665-3677.

Tanaka, M. et al., Inhibiton of heart transplant injury and graft coronary artery disease after prolonged organ ischemia by selective protein kinase C regulators. J Thorac Cardiovasc Surg. May 2005;129(6):1160-7.

Tang, D.C. et al., Genetic immunization is a simple method for eliciting an immune response. Nature. Mar. 12, 1992;356(6365):152-4.

Tanguay, R.L. et al., Translational efficiency is regulated by the length of the 3' untranslated region. Mol Cell Biol. Jan. 1996;16(1):146-56.

Taranger, C.K. et al., Induction of dedifferentiation, genomewide transcriptional programming, and epigenetic reprogramming by extracts of carcinoma and embryonic stem cells. Mol Biol Cell. Dec. 2005;16(12):5719-35.

Tavernier, G., et al., mRNA as gene therapeutic: How to control protein expression. J. of Controlled Release. Mar. 2011; 150(3): 238-247

Tazi, J. et al., Alternative chromatin structure at CpG islands. Cell. Mar. 23, 1990;60(6):909-20.

Teufel, R. et al., Human peripheral blood mononuclear cells transfected with messenger RNA stimulate antigen-specific cytotoxic T-lymphocytes in vitro. Cell Mol Life Sci. Aug. 2005;62(15);1755-62.

Thompson, M. et al., Nucleolar clustering of dispersed tRNA genes. Science. Nov. 21, 2003;302(5649):1399-401.

Thurner, B. et al., Vaccination with mage-3A1 peptide-pulsed mature, monocyte-derived dendritic cells expands specific cytotoxic T cells and induces regression of some metastates in advanced stage IV melanoma. J Exp Med. Dec. 6, 1999;190(11):1669-78.

Tourriere, H. et al., mRNA degradation machines in eukaryotic cells. Biochimie. Aug. 2002;84(8):821-37.

Towle, H.C. et al., Purification and characterization of bacteriophage gh-l-induced deoxyribonucleic acid-dependent ribonucleic acid polymerase from Pseudomonas putida. J Biol Chem. Mar. 10, 1975;250(5):1723-33.

Treat, J. et al., In Liposomes in the Therapy of Infectious Disease and Cancer, Lopez-Berestein and Fidler (eds.), Liss, New York, 1989. 353-65.

Trinchieri, G. et al., Cooperation of Toll-like receptor signals in innate immune defence. Nat Rev Immunol. Mar. 2007;7(3):179-90. Trojan, A. et al., Immune reactivity against a novel HLA-A3-restricted influenza virus peptide identified by predictive algorithms and interferon-gamma quantitative PCR. J Immunother. Jan.-Feb. 2003;26(1):41-6.

Tsuchiya, M, et al., Isolation and characterization of the cDNA for murine granulocyte colony-stimumating factor. Proc Natl Acad Sci USA. Oct. 1986; 83(20): 7633-7637.

Tung, T.C. et al., Organ formation caused by nucleic acid from different class.—Urodele DNA mediated balancer formation in goldfish. Sci Sin. Jan.-Feb. 1977;20(1):56-8.

Tung, T.C. et al., The effect of carp EGG-mRNA on the transformation of goldfish tail. Sci Sin. Jan.-Feb. 1977;20(1):59-63.

Tung, T.C. et al., Transmission of the nucleic acid-induced character, caudal fin, to the offspring in goldfish. Sci Sin. Mar.-Apr. 1975;18(2):223-31.

Tuting, T. et al., Gene-based strategies for the immunotherapy of cancer. J Mol Med (Berl). Jul. 1997;75(7):478-91.

Tycowski, K.T. et al., A small nucleolar RNA requirement for site-specific ribose methylation of rRNA in Xenopus. Proc Natl Acad Sci U S A. Dec. 10, 1996;93(25):14480-5.

Udenfriend, S., et al., The enzymatic conversion of phenylalanine to tyrosine. J. Biol. Chem. 1952; 194: 503-511.

Úeda, T. et al., Phosphorothioate-containing RNAs show mRNA activity in the prokaryotic translation systems in vitro. Nucleic Acids Res. Feb. 11, 1991;19(3):547-52.

Ulmer, J.B. et al., Heterologous protection against influenza by injection of DNA encoding a viral protein. Science. Mar. 19, 1993;259(5102):1745-9.

Ulmer, J.B., An update on the state of the art of DNA vaccines. Curr Opin Drug Discov Devel. Mar. 4, 2001(2);192-7.

Utikal, J., et al., Immortalization eliminates a roadblock during cellular reprogramming into iPS cells. Nature. Aug. 2009; 460: 1145-1148.

Uzgun, S., et al., PEGylation improves nanoparticle formation and transfection efficiency of messenger RNA. Pharm Res. Sep. 2011; 28(9): 2223-2232.

Uzri, D., et al., Nucleotide sequences and modifications that determine RIG-I/RNA binding and signaling activities. J. Virol. May 2009; 83 (9): 4174-4184.

OTHER PUBLICATIONS

Vaheri, A. and Pagano, J.S. Infectious poliovirus RNA: a sensitive method of assay. Virology. Nov. 1965; 27(3): 434-436.

Valcarcel, J. et al., The protein Sex-lethal antagonizes the splicing factor U2AF to regulate alternative splicing of transformer premRNA. Nature. Mar. 11, 1993;362(6416):171-5.

Van Den Bosch, G.A., et al., Simultaneous activation of viral Antigen-specific Memory CD4+ and CD8+ T-cells using mRNA-eletroporated CD40-activated autologus B-cells. J Immunother. Sep./Oct. 2006; 29, 512-23.

Van Gelder, R.N. et al., Amplified RNA synthesized from limited quantities of heterogeneous cDNA. Proc Natl Acad Sci U S A. Mar. 1990;87(5):1663-7.

Van Tendeloo, V.F. et al., Highly efficient gene delivery by mRNA electroporation in human hematopoietic cells: superiority to lipofection and passive pulsing of mRNA and to electroporation of plasmid cDNA for tumor antigen loading of dendritic cells. Blood. Jul. 1, 2001;98(1):49-56.

Van Tendeloo, V.F., et al., mRNA-based gene transfer as a tool for gene and cell therapy. Curr Opin Mol Therapeutics. 2007; 9(5): 423-431

Vaquero, C. et al., Transient expression of a tumor-specific single-chain fragment and a chimeric antibody in tobacco leaves. Proc Natl Acad Sci U S A,. Sep. 28, 1999;96(20):11128-33.

Varambally, S. et al., Genomic loss of microRNA-101 leads to overexpression of histone methyltransferase EZH2 in cancer. Science. Dec. 12, 2008;322(5908):1695-9. Epub Nov. 13, 2008.

Vassilev, V.B. et al., Microparticle-mediated RNA immunization against bovine viral diarrhea virus. Vaccine. Feb. 28, 2001;19(15-16):2012-9.

PCT Invitation to pay additional fees and, where applicable, protest fee for International application No. PCT/US2013/030061, dated Aug. 22, 2013.

Tripathy, Sandeep et al., Long-term expression of erythopoietin in the systemic circulation of mice after intramuscular injection of a plasmid DNA vector, Proc. Natl. Acad. Sci. USA 93, 1996, pp. 10670-10880.

Yarovoi, Helen, et al., Factor VIII ectopically expressed in platelets: efficacy in hemophilia A treatment, Blood Journal, Dec. 1, 2003, olume 102, No. 12, pp. 4005-4013.

PCT Invitation to pay additional fees and, where applicable, protest fee for International application No. PCT/US2013/030062, dated Jul. 19, 2013.

PCT Invitation to pay additional fees and, where applicable, protest fee for International application No. PCT/US2013/030064, dated Jul. 5, 2013.

Parker et al., Targeting of Polyelectrolyte RNA Complexes to Cell Surface Integrins as an Efficient, Cytoplasmic Transfection Mechanism, Journal of Bioactive and Compatible Polymers, Jul. 2002, pp. 1-10.

Kenneth Stanley, Design of Randomized Controlled Trials, Circulation, 2007; 115: pp. 1164-1169.

Chen XL, et al., Expression of human factor IX in retrovirustransfected human umbilical cord tissue derived mesenchymal stem cells, PubMed, Feb. 2009; 17 (1): 184-87.

Cowling (Jan. 15, 2010, online Dec. 23, 2009, "Regulation of mRNA cap methylation," Biochemical Journal, 425 (Pt 2): 295-302. Kozak, Marilyn, Regulation of translation via mRNA structure in prokaryotes and eukaryotes, Gene 361 (2005), pp. 13-37.

Fuke, Hiroyuki et al., Role of poly (A) tail as an identity element of mRna nuclear export, Nucleic Acids Research, 2008, vol. 36 No. 3, pp. 1037-1049.

Roger S. Riley, MD, Ph.D., Apr. 2005, http://www.pathology.vcu.edu/clinical/coag/FIX%20Deficiency.pdf, no volume, no pages, no publisher, no journal, 2 pages long.

SEQ Search Result 1(U.S. Appl. No. 13/897,362) dated Oct. 11, 2013.

Tracy, M., "Progress in the Development of LNP Delivery for siRNA Advancing LNPs to the Clinic," International Liposome Research Days Meeting, Vancouver, Canada. Aug. 2010, pp. 1-52.

International Search Report from International Application No. PCT/US2013/030064 dated Oct. 21, 2013.

International Search Report from International Application No. PCT/US2013/030062 dated Oct. 21, 2013.

International Search Report and Written Opinion from International Application No. PCT/US2011/54636 dated Apr. 17, 2013.

International Search Report for related application PCT/US2011/46861, Apr. 13, 2012.

International Preliminary Report on Patentability for related application PCT/US2012/031781, Oct. 1, 2013.

International Search Report and Written Opinion from International Application Serial No. PCT/US13/062943 dated Jan. 7, 2014.

Anderson, B.R., et al., Incorporation of pseudouridine into mRNA enhances translation by diminishing PKR activation, Nucleic Acids Res. vol. 38, No. 17, Sep. 1, 2010, pp. 5884-5892.

Kariko, K. et al, Generating the optimal mRNA for therapy: HPLC purification eliminates immune activation and improves translation of nucleoside-modified, protien-encoding mRNA. Nucleic Acids Res. vol. 39, No. 21, Nov. 1, 2011 pp. e142-1, XP002696190.

International Search Report and Written Opinion from International Application Serial No. PCT/US13/030067 dated Dec. 20, 2013. International Search Report and Written Opinion from International

Application Serial No. PCT/US13/030070 dated Dec. 23, 2013. Love et al., Lipid-like materials for low-dose, in vivo genes silencing, PNAS vol. 107 No. 5, pp. 1864-1869, Feb. 2, 2010.

Mockey et al., mRNA-based cancer vaccine: prevention of B16 melanoma progression and metastasis by systemic injection of MART1 mRNA histidylated lipopolyplexes, Cancer Gene Therapy, 2001, 14, pp. 802-814.

Kwon et al. Molecular Basis for LDL receptor recognition by PCSK9. PNAS. 2008 105(6), 1820-1825.

Bates et al., Detection of Familial Hypercholesterolaemia: A Major Treatment Gap in Preventative Cardiology, Heart, Lung and Circulation 2008;17:411-413.

Garber et al.; A sensitive and convenient method for lipoprotein profile analysis of individual mouse plasma samples. Journal of Lipid Research. 2000. 14: 1020-1026.

Goldstein et al., History of Discovery: The LDL Receptor, Arterioscler Thromb Vasc Biol. Apr. 2009; 29(4): 431-438.

Hovingh et al., Diagnosis and treatment of familial hypercholesterolaemia, European Heart Journal (2013) 34, 962-971. Kobayashi et al., Roles of the WHHL Rabbit in Translational Research on Hypercholesterolemia and Cardiovascular Diseases, Journal of Biomedicine and Biotechnology, vol. 2011, Article ID 406473, pp. 1-10.

Lambert et al., Thernatic Review Series: New Lipid and Lipoprotein Targets for the Treatment of Cardiometabolic Diseases The PCSK9 decade, Journal of Lipid Research vol. 53, 2012 pp. 2515-2524.

Lipari et al., Furin-cleaved Proprotein Convertase Subtilisin/Kexin Type 9 (PCSK9) is Active and Modulates Low Density Lopoprotein Receptor and Serum Cholesterol Levels. J Biol Chem. 2012, 287(52); 43482-43491.

Surdo et al., Mechanistic implications for LDLreceptor degradation from the PCSK9/LDLR structure at neutral pH, European Molecular Biology Organization, vol. 12 | No. 12 | 2011, pp. 1300-1130. McNutt et al., Antagonism of Secreted PCSK9 Increases Low Density Lipoprotein Receptor Expression in HepG2 Cells. J Biol Chem. 2009. 284(16): 10561-10570.

Ni et al., A PCSK9-binding antibody that structurally mimics the EGF(A) domain of LDL-receptor reduces LDL cholesterol in vivo, Journal of Lipid Research vol. 52, 2011.

Rader et al., Monogenic hypercholesterolemia: new insights in pathogenesis and treatment, J. Clin. Invest. 111:1795-1803 (2003). Stein et al., Effect of a Monoclonal Antibody of PCSK9 on LDL Cholesterol, N Engl J Med 2012;366:1108-18.

Watts et al., Familial hypercholesterolemia: a missed opportunity in preventive medicine, Nature Clinical Practice, Cardiovascular Medicine, Aug. 2007, vol. 4, No. 8, pp. 404-405.

Zhang et al., Binding of Proprotein Convertase Subtilisin/Kexin Type 9 to Epidermal Growth Factor-like Repeat A of Low Density Lipoprotein Receptor Decreases Receptor Recycling and Increases Degradation, The Journal of Biological Chemistry, vol. 282, No. 25, pp. 18602-18612, Jun. 22, 2007.

OTHER PUBLICATIONS

Penheiter et al., Type II Transforming Growth Factor-β Receptor Recyclins is Dependent upon the Clathrin Adaptor Protein Dab2, Molecular Biology of the Cell, vol. 21, 4009-4019, Nov. 15, 2010. Mulkearns et al., FCHO2 organizes clathrin-coated structures and interacts with Dab2 for LDLR endocytosis, Molecular Biology of the Cell, 2012, pp. 1-28.

Teckchandani et al., The clathrin adaptor Dab2 recruits EH domain scaffold proteins to regulate integrin β1 endocytosis, Molecular Biology of the Cell, 2012, pp. 1-28.

Stockinger et al., The PX-domain protein SNX17 interacts with members of the LDL receptor family and modulates endocytosis of the LDL receptor, European Molecular Biology Organization, vol. 21 No. 16 pp. 4259-4267.

Song et al., A putative role of micro RNA in regulation of cholesterol 7α -hydroxylase expression in human hepatocytes, Nature Biotechnol. 2005, 23:709-717.

Beigneux et al., Human CYP7A1 deficiency: progress and enigmas; The Journal of Clinical Investigation; Jul. 2002, vol. 110, No. 1, pp. 29-31.

Hofman et al., CYP7A1 A-278C Polymorphism Affects the Response of Plasma Lipids after Dietary Cholesterol or Cafestol Interventions in Humans, The Journal of Nutrition, 2004, pp. 2200-2204.

Pullinger et al., Human cholesterol 7α -hydroxylase (CYP7A1) deficiency has a hypercholesterolemic phenotype, J. Clin. Invest. 110:109-117 (2002).

Akinc et al., Targeted Delivery of RNAi Therapeutics With Endogenous and Exogenous Ligand-Based Mechanisms, Mol Ther. 2009 17:872-879.

Yamamoto et al., Current prospects for mRNA gene delivery, European Journal of Pharmaceutics and Biopharmaceutics 71 (2009) 484-489.

Trollet et al., Delivery of DNA into muscle for treating systemic diseases: advantages and challenges. Methods Mol. Biol. 2008., 423, 199-214

Lorenzi, J.C., et al., Protein expression from exogenous mRNA: Uptake by receptor-mediated endocytosis and trafficking via the lysosomal pathway. RNA Biology, vol. 8, No. 4, Jul. 1, 2011, pp. 252-258.

International Search Report from International Application No. PCT/US13/54635 dated Mar. 3, 2014.

International Search Report from International Application No. PCT/US13/030070 dated Dec. 23, 2013.

Kassim et al., Gene Therapy in a humanized Mouse Model of Familial Hypercholesterolernia Leads to a Marked Regression of Atherosclerosis, PLOS ONE, Oct. 2010, vol. 5, Issue 10, pp. e13424.

Supplementary Data from Zhang et al., (J. Biol. Chem 282(25) 18602-12, 2007.

International Search Report from International Application No. PCT/US12/054574 dated Jul. 1, 2013.

NCBI BLAST (hypp://blast.ncbi.nlm.nih.gov/Blast.cgi;accession No. BE136127, 2007.

Bell et al., Predisposition to Cancer Caused by Genetic and Functional Defects of Mammalian Atad5, PLOS Genetics, Aug. 2011, vol. 7, Issue 8, e1002245 pp. 1-15.

Gupta et al., Project Report Condon Optimization, 2003, pp. 1-13. Whiteside, George, The Orgins and the future of microfluidics, Nature, vol. 442, Jul. 27, 2006 pp. 368-373.

Pridgen, et al.; Transepithelial Transport of Fc-Targeted Nanoparticles by the Neonatal Fc Receptor for Oral Delivery, Sci Translation Med., vol. 5, Issue 213, Nov. 27, 2013, pp. 1-8.

Nguyen, M. et al., Injectable Biodegradable Hydrogels, Macromolecular Bioscience, 2010,10, 563-579.

Morton, S. Scalable Manufacture of Built-to-Order Nanomedicine: Spray-Assisted Layer-by-Layer Functionalization of Print Nanoparticles, Advanced Materials, 2013, 25, 4708-4712.

Li, Z et al., Controlled Gene Delivery System Based pn Thermosensitive Biodegradeable Hydrogel, Pharmaceutical Research, vol. 20, No. 6, Jun. 2003.

Lee, et al.; Thermosensitive Hydrogel as a Tgf- β 1 Gene Delivery Vehicle Enhances Diabetic Wound Healing, Pharmaceutical Research, vol. 20, No. 12, Dec. 2003.

Cu, Y. et al., Enhanced Delivery and Potency of Self-Amplifying mRNA Caccines by Electroporation in Situ, Vaccines, 2013, 1, 367-383.

Chang, C. et al., Non-ionic amphiphilic biodegradable PEG-PLGA-PEG copolymer enhances gene delivery efficiency in rat skeletal muscle; Science Direct, Journal of Controlled Release 118 (2007) 245-253.

Nelson, C. et al., Tunable Delivery of SiRNA from a Biodergradable Scaffold to Promote Angiogenesis In Vivo, Advanced Materials, 2013, pp. 1-8.

Stroock, A. et al., Chaotic Mixer for Microchannels, Science, vol. 295, Jan. 25, 2002, pp. 1-6.

Zangi, L. et al., Modified mRNA directs the fate of heart progenitor cells and indices vasuclar regeneration after myocardial infarction, Nature Biology, Advanced Online Publication, May 10, 2013, pp. 1-9.

Valencia, P. et al., Microfluidic Platform for Combinatorial Synthesis and Optimization of Targeted Nanoparticles for Cancer Therapy, ACS Nano. Dec. 23, 2013; 7(12):10671-80.

Chen, Y., Self-assembled rosette nanotubes encapsulate and slowly release dexamethasone, International Journal of Nanomedicine, 2011:6 pp. 1035-1044.

Mitragotri, S.; Devices for Overcoming Biological Barriers: The use of physical forces to disrupt the barriers, Advance Drug Delivery Reviews. 65 (2013)100-103.

Wang, X.; Re-evaluating the Roles of Proposed Modulators of Mammalian Target of Rapamycin Complex 1 (mTORCI) Signaling, The Journal of Biological Chemistry, Nov. 7, 2008, vol. 283, No. 45, pp. 30482-30492.

Dreyer Hans C., Leucine-enriched essential amino acid and carbohydrate ingestion following resistance exercise chances mTOR signaling and protein synthesis in human muscle, AM J. Physiol Endocrinol Metab,; 294; E392-E400,2008.

Lalatsa, Aikaterini, Amphiphilic poly (I-amino acids)—New materials for drug delivery. Journal of Controlled Release, 161 (2012) 523-536.

Stelic Institute & Co., Contract Research Services Specialized in NASH-HCC, Ver.2012.11, 2012, 99.1-10.

Limberis, M et al., Intranasal Antibody Gene Transfer in Mice and Ferrets Elicits Broad Protection Against Pandemic Influenza, Science Transl Med vol. 5, Issue 187, 99. 1-8.

Wei, et al., Induction of Broadly Neutralizing H1N1 Influenza Antibodies by Vaccination, Science vol. 329, (2010) pp. 1060-1064. Palese, P., Making Better Influenza Virus Vaccines?, Emerging Infectious Diseases, vol. 12, No. 1, Jan. 2006, pp. 61-65.

Kwong, P et al., Broadly Neutralizing Antibodies and the Search for an HIV-1 Vaccine: The End of the Beginning, Nature Reviews, Immunology, vol. 13, Sep. 2013, pp. 693-701.

DeMarco, et al., A Non-VH1-69 Hetetrosubtypic Neutrilizing Human Minoclonal Antibody Protects Mice Against H1N1 and H5N1 Viruses, PLOS One, Apr. 2012, vol. 7, Issue 4, pp. 1-9.

Anderson, et al., The Bridge, National Academy of Engineering of the National Academies, Fall 2006, vol. 36., Nov. 3, pp. 1-55.

EP11830061, Supplementary Search Report, Mar. 18, 2014.

Iwase, Reiko et al., Molecular design of a eukaryotic messenger RNA and its chemical synthesis, Nucleic Acids Research, 1991, vol. 20, No. 7, pp. 1643-1648.

Squires, Jeffrey et al., Widespread occurrence of 5-methylcytosine in human coding an non-coding RNC, Nucleic Acids Research, 2012, vol. 40, No. 11, pp. 5023-5033.

Wyatt, et al., Occurrence of 5-Methyl-Cytosine in Nucleic Acid, 1950, vol. 166, No. 4214, pp. 237-238.

Chen, Chun et al., A Flexible RNA Backbone within the Polypyrimidine Tract Is Required for U2AF65 Binding and PremRNA Splicing In Vivo, Molecular and Cellular Biology, 2010, vol. 30, No. 17, pp. 4108-4119.

OTHER PUBLICATIONS

Wantabe, Hiroshi, et al., Conformational Stability and Warfarin-Binding Properties of Human Serum Albumin Studied by Recombinany Mutants, Biochem. J., 2001, vol. 357, No number, pp. 269-274

Abramova, Tatyana, Frontiers and Approaches to Chemical Synthesis of Oligodeoxyribonucleotides, Molecules 2013, vol. 57, No. 18, 1063-1075.

Bain, J.D. et al., Regioselective ligation of oligoribonucleotides using DNA Splints, Nucleic Acids Research, vol. 20, No. 16, p. 4372.

Bonora, G. et al., HELP (High Efficiency Liquid Phase) new oligonucleotide synthesis on soluble polymeric support, Oxford Journals Life Sciences Nucleic Acids Research vol. 18, Issue 11 pp. 3155-3159.

Borovkov, A. Et al., High-Quality Gene Assembly Directly From Unpurified Mixtures of Microarray-Synthesized Oligonucleotides, Nucleic Acids Research, 2010, vol. 38, No. 19, pp. e180 1-e180 10. Cheng, S. et al. Effective Amplification of Long Targets From Cloned Inserts and Hunam Genomic DNA, Proc. Nati. Acad. Sci. USA, 1994, vol. 91, pp. 5695-5699.

Cleary, Michele et al., Production of Complex nucleic acid libraries using highly parallel in situ oligonucleotide synthesis, 2004, Nature Methods vol. 1 No. 3, Dec. 2004, pp. 241-248.

El-Sagheer, Afaf H. et al., Click Nucleic Acid Ligation: Applications in Biology and Nanotechnology, Accounts of Chemical Research, 2012 vol. 45, No. 8, pp. 1258-1267.

Freeman, Willard M. et al., Quantitative RT-PCR: Pitfalls and Potential, BioTechniques, 1999, vol. 26, No. 1, pp. 112-125.

Gibson, D. et al., Creation of a Bacterial Cell Controlled by a Chemically Synthesized Genome, Science, 2010, vol. 329, No. 52, pp. 51-56.

Gibson, Daniel G., Chemical Synthesis of the Mouse Mitochondrial Genome, Nature Methods, vol. 7., No. 11 Nov. 2010, pp. 901-905. Goodchild, John et al., Conjugates of Oligonucleotides and Modified Oligonucleotides: A Review of Their Synthesis and Properties, Bioconjugate Chemistry, 1990, vol. 1., No. 3., pp. 165-187.

Innis, M., DNA Sequencing with Thermus Aquaticus DNA Polymerase and Direct Sequencing of Polymerase Chain Reaction-Amplified DNA, Proc. Natl. Acad. Sci. USA, 1988, vol. 85, pp. 9436-9440.

Kang, Hyunmin, Inhibition of MDR1 Gene Expression by Chimeric HNA Antisense Oligonucleotides, Nucleic Acids Research, 2004, vol. 32, No. 14, pp. 4411-4419.

Lavrik, Inna N. et al., Translational Properties of mHNA, a Messenger RNA Containing Anhydrohexitol Nucleotides, Biochemistry 2001, vol. 40, No. 39, pp. 11777-11784.

Li, Junjie, et al.; Methylation Protects miRNAs and siRNAs from a 3_-End Uridylation Activity in Arabidopsis, Current Biology, 2005, vol. 15, (no number), pp. 1501-1507.

Lizardi, PM., et al., Mutation Detection and Single-Molecule Counting Using Isothermal Rolling-Circle Amplification, Nat Genetics, 1998, vol. 19, No #, pp. 225-232.

Martinelli, Richard A., Chemiluminescent Hybridization-Ligation Assays for F508 and I507 Cystic Fibrosis Mutations, Clinical Chemistry, 1996, vol. 42., No. 1, pp. 14-18.

Moore, M., Site-Specific Modification of Pre-mRNA: The 2"-Hydroxyl Groups at the Splice Sites, Science, 1992, vol. 256, No #, pp. 992-997.

Nagata, S., Synthesis and Biological Activity of Artificial mRNA Prepared with Novel Phosphorylating Reagents, Nucleic Acids Research, 2010, vol. 38, No. 21, pp. 7845-7857.

Norbury, Chris J., Cytoplasmic RNA: A Case of the Tail Wagging the Dog, Nature Reviews, Molecular Cell Biology, 2013, Advanced Online Publication, No Volume Number, pp. 1-10.

Nwe, K. et al., Growing Applications of "Click Chemistry" for Bioconjugation in Contemporary Biomedical Research, Cancer Biotherapy and Radiopharmaceuticals, 2009, vol. 24., No. 3., pp. 289-301.

Ochman, H., Genetic Applications of an Inverse Polymerase Chain Reaction, Genetics, Washington University School of Medicine, 1988, vol. 120, No #, pp. 621-623.

Polidoros, A. et al., Rolling Circle Amplification—RACE: a method for Simultaneous Isolation of 5" and 3" cDNA ends from Amplified cDNA templates, Benchmarks, Biotechniques, 2006, vol. 41, No. 1, pp. 35-42.

Pon, R., Multiple Oligodeoxyribonucleotide Syntheses on a Reusable Solid-Phase CPG Support Via the Hydroquinone-O, O"-diacetic acid (Q-Linker) linker arm, Nucleic Acids Research, 1999, vol. 27, No. 6, pp. 1531-1538.

Shiba, Y. et al., Chemical Synthesis of a Very Long Oligoribonucleotide with a 2-cyanoethoxymethyl (CEM) as the 2'-O-protecting Group: Structural Identification and Biological Activity of a Synthetic 110mer precursor-microRNA Candidate, Nucleic Acids Research, 2007, vol. 35, No. 10, pp. 3287-3296. Sindelar, L. et al., High-throughput DNA Synthesis in a Multichannel Format, Nucl. Acids Res. 1995, vol. 23, No. 6, pp. 982-987.

Stark, M. et al., An RNA Ligase-mediated Method for the Efficient Creation of Large, Synthetic RNAs, Method, 2006, vol. 12, No Vol. number, pp. 2014-2019.

Walker, T., Isothermal In Vitro Amplification of DNA by a Restriction Enzyme/ DNA Polymerase System, Proc. Natl. Acad. Sci. USA, 1992, vol. 89, No number, pp. 392-396.

Zhu, B., Syn5 RNA Polymerase Synthesizes Precise Run-Off RNA Products, Nucleic Acids Research, 2013, vol. 103, No #, pp. 1-10. Prokazyme Ltd., ThermoPhage, ssDNA ligase,2013, No Vol. pp. 1-3

Prokaria Ltd, Tsc DNA ligase, 2013, No Vol., pp. 1-3.

Bolhassani A., et al., Improvement of Different Vaccine Delivery Systems for Cancer Therapy, Molecular Cancer, Biomed Central, London, GB, 2011, vol. 10, No. 3, pp. 1-20.

Cheng, Ee-chun et al., Repressing the Repressor: A lincRNA as a MicroRNA Sponge in Embryonic Stem Cell Self-Renewal, Developmental Cell, 2013, vol. 25, No number, pp. 1-2.

Memczak, Sebastian et al., Circular RNAs are a large class of animal RNAs with Regulatory Potency, Nature, 2013, vol. 495, no number, pp. 333-343.

Hentze, M., Circular RNAs: Splicing's Enigma Variations, The EMBO Journal, 2013, vol. 32, no number, pp. 923-925.

Ledford, Heidi et al, Circular RNAs Throw Genetics for a Loop, In Focus News, Nature, vol. 494, pp. 291-292.

Salzman, Julia et al., Circular RNAs Are the Predominant Transcript Isoform From Hundreds of Human Genes in Diverse Cell Types, PLOS One, 2012, vol. 7, Issue 2, pp. 1-12.

Ebert, Margaret S., MicroRNA sponges: Competitive Inhibitors of Small RNAs in Mammalian Cells, Nature Methods, 2007, vol. 4, No. 9, pp. 721-726.

Jeck, William et al. Circular RNAs Are Abundant, Conserved, and Associated with ALU Repeats, RNA, 2013, vol. 19, pp. 141-157. Matsuda, V. et al., Determinants of Initiation Codon Selection During Translation in Mammalian Cells, PLOS One, 2010, vol. 5, Issue 11, pp. 1-13.

Mukherji, S. et al., MicroRNAs Can Generate Thresholds in Target Gene Expression, Nature Genetics, 2011, vol. 43, No. 9, pp. 854-860.

Hansen, Thomas et al., Natural RNA Circles Function As Efficient MicroRNA Sponges, Nature, 2013, vol. 495, no number, pp. 384-300

Rose, Jason, MicroRNA "Sponge": Proof of Concept for a Novel MicroRNA Target Identification Technique, A Major Qualifying Project Report, Submitted to the Faculty of Worcester Polytechnic Institute, 2010, No Volume, pp. 1-26.

Touriol, C. et al., Generation of Protein Isoform Diversity by Alternative Initiation of Translation At Non-AUG Codons, Biology of the Cell, 2003, vol. 95, no number, pp. 168-178.

Wang et al., Endogenous miRNA Sponge lincRNA-RoR Regulates Oct4, Nanog, and Sox2 in Human Embryonic Stem Cell Self-Renewal, Developmental Cell, 2013, vol. 25, No #, pp. 69-80.

Goldberg, I.H. et al., The incorporation of 5-ribosyluracil triphosphate into RNA in nuclear extracts of mammalian cells. Biochemical Biophysical Research Communications. 1961; 6(5): 394-398.

OTHER PUBLICATIONS

Goldberg, I.H. et al., Comparative utilization of pseudouridine triphosphate and uridine triphosphate by ribonucleic acid polymerase. J Biological Chem. May 1963; 238(5): 1793-1800.

Gordon, S.N. et al., Targeting the vaginal mucosa with human papillomavirus pseudovirion vaccines delivering SIV DNA. J Immunol. Jan. 15, 2012; 188(2): 714-723.

Grabbe, S. et al., Dendritic cells as initiators of tumor immune responses: a possible strategy for tumor immunotherapy? Immunol Today. Mar. 1995;16(3):117-21.

Grabbe, S. et al., Tumor antigen presentation by epidermal antigenpresenting cells in the mouse: modulation by granulocyte-macrophage colony-stimulating factor, tumor necrosis factor alpha, and ultraviolet radiation. J Leukoc Biol. Aug. 1992;52(2):209-17.

Grabbe, S. et al., Tumor antigen presentation by murine epidermal cells. J Immunol. May 15, 1991;146(10):3656-61.

Graf, M. et al., Codon-optimized genes that enable increased heterologous expression in mammalian cells and elicit efficient immune responses in mice after vaccination of naked DNA. Methods Mol Med. 2004;94:197-210.

Graham, F.L., et al., A new technique for the assay of infectivity of human adenovirus 5 DNA. Virology. Apr. 1973;52(2):456-67.

Gram, G.J. et al., Immunological analysis of a Lactococcus lactis-based DNA vaccine expressing HIV gp120. Genet Vaccines Ther. Jan. 29, 2007;5:3.

Granstein, R.D. et al., Induction of anti-tumor immunity with epidermal cells pulsed with tumor-derived RNA or intradermal administration of RNA. J Invest Dermatol. Apr. 2000;114(4):632-6. Greenblatt, M.S. et al., Mutations in the p53 tumor suppressor gene: clues to cancer etiology and molecular pathogenesis. Cancer Res. Sep. 15, 1994;54(18):4855-78.

Grentzmann, G. et al., A dual-luciferase reporter system for studying recoding signals. RNA. Apr. 1998;4(4):479-86.

Grosjean, H., Modification and editing of RNA: historical overview and important facts to remember. Fine-tuning of RNA functions by modification and editing. Topics Curr Gen. Jan. 2005; 12: 1-22.

Gross, G. et al., Heterologous expression as a tool for gene identification and analysis. J Biol Chem. Jul. 31, 1995;41(2):91-110. Grudzien, E. et al., Novel cap analogs for in vitro synthesis of mRNAs with high translational efficiency. RNA. Sep. 2004;10(9):1479-87.

Grudzien-Nogalska, E. et al., Phosphorothioate cap analogs stabilize mRNA and increase translational efficiency in mammalian cells. RNA. Oct. 2007;13(10):1745-55. Epub Aug. 24, 2007.

Gryaznov, S.M., Oligonucleotide N3'→P5'phosphoramidates as potential therapeutic agents. Biochim Biophys Acta. Dec. 10, 1999;1489(1):131-40.

Guhaniyogi, J. et al., Regulation of mRNA stability in mammalian cells. Gene. Mar. 7, 2001;265(1-2):11-23.

Guo, L. et al., Structure and function of a cap-independent translation element that functions in either the 3' or the 5' untranslated region. RNA. Dec. 2000;6(12):1808-20.

Haas, J. et al., Codon usage limitation in the expression of HIV-1 envelope glycoprotein. Curr Biol. Mar. 1, 1996;6(3):315-24.

Hakelien, A.M., et al., Novel approaches to transdifferentiation. Cloning Stem Cells. 2002;4(4):379-87.

Hakelien, A.M., Reprogramming fibroblasts to express T-cell functions using cell extracts. Nat Biotechnol. May 2002;20(5):460-6.

Hambraeus, G. et al., A 5' stem-loop and ribosome binding but not translation are important for the stability of Bacillus subtilis aprE leader mRNA. Microbiology. Jun. 2002;148(Pt 6):1795-803.

Hancock, J.F., Reticulocyte lysate assay for in vitro translation and posttranslational modification of Ras proteins. Methods Enzymol. 1995;255:60-5.

Hannon, G.J. et al., Trans splicing of nematode pre-messenger RNA in vitro. Cell. Jun. 29, 1990;61(7):1247-55.

Harel, J., Action of polyribonucleotides, extracted by the phenol method, on the growth of mouse tumor cells. C.R. Hebd Seances Acad. Sci., 1962, 254:4390-2.

Harris, J. et al., An improved RNA amplification procedure results in increased yield of autologous RNA transfected dendritic cell-based vaccine. Biochim Biophys Acta. Jun. 20, 2005;1724(1-2):127-36. Epub Apr. 7, 2005.

Hausmann, R., Bacteriophage T7 genetics. Curr Top Microbiol Immunol. 1976;75:77-110.

Hays, E.F. et al., Induction of mouse leukaemia with purified nucleic acid preparations. Nature. Dec. 21, 1957;180(4599):1419-20.

He, K. et al., Synthesis and Separation of Diastereomers of Ribonucleoside 5'-(alpha-P-Borano)triphosphates. J Org Chem. Aug. 21, 1998;63(17):5769-5773.

Hecker, J.G. et al., Non-Viral DNA and mRNA Gene Delivery to the CNS Pre-Operatively for Neuroprotection and Following Neurotrauma. Molecular Therapy. 2004; 9, S258-S258.

Hedman, M, et al., Safety and feasibility of catheter-based local intracoronary vascular endothelial growth factor gene transfer in the prevention of postangioplasty and in-stent restenosis and in the treatment of chronic myocardial ischemia: phase II results of the Kuopio Angiogenesis Trial (KAT). Circulation. Jun. 3, 2003; 107(21): 2677-83. Epub May 12, 2003.

Heidenreich, O. et al., Chemically modified RNA: approaches and applications. FASEB J. Jan. 1993;7(1):90-6.

Heidenreich, O. et al., High activity and stability of hammerhead ribozymes containing 2'-modified pyrimidine nucleosides and phosphorothioates. J Biol Chem. Jan. 21, 1994;269(3):2131-8.

Heil, F. et al., Species-specific recognition of single-stranded RNA via toll-like receptor 7 and 8. Science. Mar. 5, 2004;303(5663):1526-9. Epub Feb. 19, 2004.

Heilman, K.L. et al., Internal 6-methyladenine residues increase the in vitro translation efficiency of dihydrofolate reductase messenger RNA. Int J Biochem Cell Biol. Jul. 1996; 28(7): 823-829.

Heiser, A. et al., Autologous dendritic cells transfected with prostate-specific antigen RNA stimulate CTL responses against metastatic prostate tumors. J Clin Invest. Feb. 2002;109(3):409-17.

Heiser, A. et al., Human dendritic cells transfected with renal tumor RNA stimulate polyclonal T-cell responses against antigens expressed by primary and metastatic tumors. Cancer Res. Apr. 15, 2001;61(8):3388-93.

Heiser, A. et al., Human dendritic cells transfected with RNA encoding prostate-specific antigen stimulate prostate-specific CTL responses in vitro. J Immunol. May 15, 2000;164(10):5508-14.

Heiser, A. et al., Induction of polyclonal prostate cancer-specific CTL using dendritic cells transfected with amplified tumor RNA. J Immunol. Mar. 1, 2001;166(5):2953-60.

Helbock, H.J. et al. N2-methyl-8-oxoguanine: a tRNA urinary metabolite—role of xanthine oxidase. Free Radic Biol Med. 1996;20(3):475-81.

Hemmi, H. et al, A Toll-like receptor recognizes bacterial DNA. Nature. Dec. 7, 2000;408(6813):740-5.

Herweijer, H. et al., Gene therapy progress and prospects: hydrodynamic gene delivery. Gene Ther. Jan. 2007;14(2):99-107. Epub Nov. 30, 2006.

Hess, M. et al., The effects of nucleic acids on pituitary ACTH content. Endocrinology. Mar. 1961;68:548-52.

Higman, M.A. et al., The mRNA (guanine-7-)methyltransferase domain of the vaccinia virus mRNA capping enzyme. Expression in *Escherichia coli* and structural and kinetic comparison to the intact capping enzyme. J Biol Chem. May 27, 1994;269(21):14974-81. Higman, M.A. et al., The vaccinia virus mRNA (guanine-N7-)-

methyltransferase requires both subunits of the mRNA capping enzyme for activity. J Biol Chem. Aug. 15, 1992;267(23):16430-7. Hilleren, P. et al., Mechanisms of mRNA surveillance in eukaryotes. Annu Rev Genet. 1999;33:229-60.

Hillman, N.W. et al., Chick Cephalogenesis, I. The Effect of RNA on Early Cephalic Development. PNAS, 1963, 50:486-93.

Ho, CS., et al., Electrospray ionisation mass spectrometry: Principles and clinical applications. Clin Biochem Rev. Feb. 2003; 24: 3-12.

Hoath, S.B. et al., The organization of human epidermis: functional epidermal units and phi proportionality. J Invest Dermatol. Dec. 2003;121(6):1440-6.

WHO Drug Information, International Nonproprietary Names for Pharmaceutical Substances (INN), 1993, vol. 7, No. 4, pp. 1-16.

OTHER PUBLICATIONS

Eli Lilly and Company, ReoPRo, Abciximab, Product Label, 2005, No volume number, pp. 1-4.

Kempeni, Joachim et al., Preliminary Results of Early Clinical Trials with the Fully Human Anti-TNFa Monoclonal Antibody D2E7, Ann Rheum Dis, 1999, vol. 58, Supp I, pp. 170-172.

Lindner, Heidrun et al., Peripheral Blood Mononuclear Cells Induce Programmed Cell Death in Human Endothelial Cells and May Prevent Repair: Role of Cytokines, 1997, vol. 89, No. 6, pp. 1931-1938.

Crowe, J.S. et al., Humanized Monoclonal Antibody CAMPATH-1H Myeloma Cell Expression of Genomic Constructs, Nucleotide Sequence of cDNA Constructs and Comparison of Effector Mechanisms of Myeloma and Chinese Hamster Ovary Cell-Derived Material, Clinical Exp. Immunol., 1992, vol. 87, No number, pp. 105-110.

Ferrara, James et al., Graft-versus Host Disease, Lancet, 2009, vol. 373, No. 9674, pp. 1550-1561.

Hale, G. et al., Removal of T Cells From Bone Marrow for Transplantation: a Monoclonal Antilyphocyte Antibody That Fixes Human Complement, Blood, 1983, vol. 62, No. 4, pp. 873-882.

Lutz, Riechmann et al., Reshaping Human Antibodies for Therapy, Nature, 1988, vol. 332, No. 24, pp. 323-327.

Novartis, Product Label, Simulect, Basiliximab, 1998, No Vol. pp. 1-7.

Baker, Kevin P. et al., Generation and Charaterization of LymphonStat-B, a Human Monoclonal Antibody That Antagonizes the Bioactivities of B Lymphocyte Stimulator, Arthritis & Rheumatism, 2003, vol. 48, No. 11, pp. 3253-3265.

ADIS R&D Profile, Belimumab, Drugs R D, 2010; vol. 10, No. 1, pp. 55-65.

Avastin, Bevacizumab, Labeling Text, 2013, No Volume, pp. 1-27. Chen, Helen et al., Expanding the Clinical Development of Bevacizumab, The Oncologist, 2004, vol. 9, Supp 1, pp. 27-35.

Herbst, Roy et al., Non-Small Cell Lung Cancer and Antiangiogenic Therapy: What Can Be Expected pf Bevacizumab?, The Oncologist, 2004, vol. 9 Supp. 1, pp. 19-26.

Presta, Leonard G. et al., Humanization of Anti-Vascular Endothelial Growth Factor Monoclonal Antibody for the Therapy of Solid Tumors and Other Disorders, Cancer Research, 1997, vol. 57, pp. 4593-4599.

Bowen, Michael et al., Functional Effects of CD30 on a Large Granular Lymphoma Cell Line, YT, The Journal of Immunology, 1993, vol. 151, No. 11, pp. 1-11.

ADCETRIS, brentuximab vedotin, Product Label, 2011,No Volume, pp. 1-15.

Francisco, Joseph et al., cAc10-vcMMAE, an Anti-CD30-monomethyl Auristatin E Conjugate with Potent and Selective Antitumor Activity, Blood, 2003,vol. 102, No. 4, pp. 1458-1465.

Wahl, Alan F. et al, The Anti-CD30 Monoclonal Antibody SGN-30 Promotes Growth Arrest and DNA Fragmentation in Vitro and Affects Antitumor Activity in Models of Hodgkins's Disease, Cancer Research, 2002, vol. 62, pp. 3737-3742.

Alten, Rieke et al., The Human Anti-IL-1β Monoclonal Antibody ACZ885 is Effective in Joint Inflammation Models in Mice and In a Proof-of-Concept Study in Patients with Rheumatoid Arthritis, Arthritis Research & Therapy, 2008, vol. 10, No. 3, pp. 1-9. Canakinumab FDA Label, 2009, No Volume # pp. 1-11.

Church, L et al., Canakinumab, a Fully Human mAB Against IL-1 β for the Potential Treatment of Inflammatory Disorder, Current Opinion in Molecular Therapeutics, 2009, vol. 11, No. 1, pp. 81-89. Lachmann, Helen et al., In Vivo Regulation of Interleukin 1 β in Patients With Cryopyrin-Associated Periodic Syndromes, The Journal of Experimental Medicine, 2008, vol. 206, No. 5, pp. 1029-1036.

Lachmann, Helen et al., Use of Canakinumab in the Cryopyrin-Associated Periodic Syndrome, The New England Journal of Medicine, 2009, vol. 360, No. 23, pp. 2416-2425.

Rowe, William S. et al., Update on the Pathogenesis and Treatment of Systemic Idiopathic Arthritis, Curr. Opinion Pediat, 2011, vol. 23, No. 6, pp. 640-646.

Wells, Michael J. et al., Pathophysiology and Clinical Implications of Pulmonary Arterial Enlargement in COPD, International Journal of COPD, 2013, vol. 8, No number, pp. 509-521.

ImClone Systems Incorporated and Bristol-Myers Squibb Company, ERBITUX, Cetuximab, 2004, No Vol number, pp. 1-18.

Goldstein, N et al., Biological Efficacy of a Chimeric Antibody to the Epidermal Growth Factor Receptor in a Human Tumor Xenograft Model, Clinical Cancer Research, 1995, vol. 1, No number, pp. 1311-1318.

Mendelsohn, J. et al, Epidermal Growth Factor Receptor Inhibition by a Monoclonal Antibody as Anticancer Therapy, 1997, vol. 3 No #, pp. 2703-2707.

Xiang, Bo et al., Colorectal Cancer Immunotherapy, Discovery Medicine, 2013, No Vol., pp. 1-8.

Chapman, Andrew et al., Therapeutic Antibody Fragments With Prolonged in Vivo Half-Lives, Nature America Inc., 1999, vol. 17, No Number, pp. 780-783.

Choy et al, Efficacy of a Novel PEGylated Humanized Anti-TNF Fragment (CDP870) in patients with Rheumatoid Arthritis: A phase II double-blinded, randomized, Dose-Escalating Trial, Rheumatology 2002; vol. 41, No number, pp. 1133-1137.

CIMZIA, Product Label, Reference ID: 3217327, UCB, Inc., 2008, No. Vol #, pp. 1-26.

Goel, N. et al, Certolizumab pegol, mABS, 2010, vol. 2, No. 2, pp. 137-147.

Mease, PJ et al., Effect of certolizumab pegol on signs and symptoms in patients with psoriatic arthritis: 24-week results of a Phase 3 double-blind randomized placebo-controlled study (RAPID-PsA), Ann Rheum Dis, 2014, vol. 73, No #, pp. 48-55.

Queen, C et al., A humanized antibody that binds to the interleukin 2 receptor, Proc. Natl. Acad. Sci. USA, 1989, vol. 86, pp. 10029-10033.

Jaffers, Gregory et al, Monoclonal Antibody Therapy, Transplantation, 1986, vol. 41, No. 5, pp. 572-578.

Ortho Multicenter Transplant Study Group, A Randomized Clinical Trial of OKT3 Monoclonal Antibody for Acute Rejection of Cadaveric Renal Transplants, The New England Journal of Medicine, 1985, vol. 313, No. 6, pp. 337-342.

Roche, Zenapax (daclizumabl) Sterile Concentrate for Injection, 2013, No Vol., pp. 1-11.

Bekker, Pirow et al., The Effect of a Single Dose of Osteoprotegerin in Postmenopausal Women, Journal of Bone and Mineral Research, 2001, vol. 16, No. 2, pp. 1-13.

Bekker, Prow et al., A single-Dose Placebo-Controlled Study of AMG 162, a Fully Human Monoclonal Antibody to RANKL, in Postmenopausal Women, Journal of Bone and Mineral Research, 2004, vol. 19, No. 7, pp. 1-8.

Body, Jean-Jacques et al., A Study of the Biological Receptor Activator of nuclear Factor-KappaB Ligand inhibitor, Denosumab, in patients with multiple myeloma or bone metastases from Breast Cancer, Clinical Cancer Research, 2006, vol. 12, No #, pp. 1221-1228.

Westenfeld, Ralf et al., Anti-RANKL therapy—implications for the bone-vascular-axis in CKD? Denosumab in post-menopausal women with low bone mineral density, Nephrol Dial Transplant, 2006, vol. 21, pp. 2075-2077.

Xgeva (denosumab) Product Label 2010-2013 pp. 1-16.

Hillmen, Peter et al., Effect of Eculizumab on Hemolysis and Transfusion Requirements in Patients with Paroxysmal Nocturnal Hemoglobinuria, The New England Journal of Medicine, 2004, vol. 350, No. 6, pp. 552-559.

Ministry of Health, Labour and Welfare, Report on the Deliberation Results, Soliris for Intravenous Infusion 300 mg, 2010, No Vol., pp. 1-105.

Golimumbab—Product Label—Janssen Biotech, Inc., 2013, No Volume number, pp. 1-19.

Garcia, Maria et al., Patient Consideration in the Management of Rheumatoid Arthritis: Role of Once-A-Month Golimumab Injection, Clinical Medical Insights: Therapeutics, Libertas Academica, 2011, vol. 3, No #, pp. 415-423.

OTHER PUBLICATIONS

Mazumdar, Sohini et al., Golimumab, mAbs, 2009, vol. 1, No. 5, pp. 422-431.

Shealy, David et al., Characterization of Golimumab, A Human Antibody Specific for Human Tumor Necrosis Factor α, mAbs, 2010, vol. No. 2, No. 4, pp. 428-439.

Hainsworth, John, Monoclonal Antibody Therapy in Lymphoid Malignancies, The Oncologist, 2000, vol. 5, No #, pp. 376-384. FDA Label, Ibritumomab Tiuxetan, ZEVALIN, 2001, IDEC Phar-

maceuticals Corporation, No Vol. pp. 1-38.

Wagner, Henry et al., Admiration Guidelines for Radioimmunotherapy of Non-Hodgkin's Lymphoma with 90Y-Labeled Anti-CD20 Monoclonal Antibody, 90Y Radioimmunotherapy Administration, The Journal of Nuclear Medicine, 2002, vol. 43, No. 2, pp. 267-272.

Who Drug Information, International Nonproprietary Names for Pharmaceutical Substances (INN), Recommended INN, 2000, vol. 14, No. 1, pp. 39-76.

Fellner, Christopher et al., Ipilimumab (Yervoy) Prolongs Survival in Advanced Melanoma, Drug Forecast, 2012, vol. 37, No. 9, pp. 503-530.

Hooks, Michael et al., Muromonab CD-3: A Review of Its Pharmacology, Pharmacokinetics, and Clinical Use in Transplantation, Pharmacotherapy, 1991, vol. 11, No. 1, pp. 26-37.

FDA Guide, TYSABRI, Elan Pharmaceuticals, Inc., Reference ID: 3308057, Biogen Idec, Inc. 2013, No Volume #, pp. 1-6.

Gordon, F.H., A Pilot Study of Treatment of Active Ulcerative Colitis With Natalizumab, a Humanized Monoclonal Antibody to Alpha-4 Integrin, Aliment Pharacol Ther, 2002, vol. 16, No #, pp. 699-705.

Guagnozzi, Danila etal, Natalizumab in the Treatment of Crohn's Disease, Biologics: Targets & Therapy, vol. 2, No. 2, pp. 275-284. Nicholas, J et al., New and Emerging Disease-Modifying Therapies for Relapsing-Remitting Multiple Sclerosis: What is New and What is to Come, Journal of Central Nervous System Disease, 2012, vol. 4, No#, pp. 81-103.

Minagar, Alireza et al., Current and Future Therapies for Multiple Sclerosis, Scientifica, 2012, vol. 2013, Artible ID 249101, pp. 1-11. Cong, Shundong et al., Novel CD20 Monoclonal Antibodies for Lymphoma Therapy, Journal of Hematology & Oncology, 2012, vol. 5, No. 64, pp. 1-9.

FDA Label, ARZERRA, Prescribing Info, 2009, GlaxoSmithKline, No. Vol., pp. 1-13.

Issa, Ghayas et al., Movel Agents in Waldenstrom Macroglobulinemia, Clin Investig, 2011, vol. 1, No. 6, pp. 815-824. Jaglowski, Samantha et al., The clinical application of monoclonal antibodies in chronic lymphocytic leukemia, Blood, 2010, vol. 116, No #, pp. 3705-3714.

Rosman, Ziv et al., Biologic Therapy for Autoimmune Diseases: an update, BMC Medicine, 2013, vol. 11 No. 88 pp. 1-12.

Teeling, Jessica et al., Characterization of New Human CD20 Monoclonal Antibodies with Potent Cytolytic Activity Against Non-Hodgkin Lymphomas, Blood, 2004, vol. 104, No#, pp. 1793-1800

Teeling, Jessica et al., The Biological Activity of Human CD20 Monoclonal Antibodies Is Linked to Unique Epitopes on CD20, The Journal of Immunology, 2006, vol. 177, No #, pp. 362-371.

Zhang, Bodi et al., Ofatumumab, mAbs, 2009, vol. 1, No. 4, pp. 326-331

Vichyanond, Pakit et al., Omalizumab in allergic diseases, a recent review, Asian Pac J Allergy Immunol, 2011, vol. 29, No #, pp. 209-219.

Thomson, Neil et al, Circulatory, Respiratory and Pulmonary Medicine, Clinical Medicine Insights, 2012, vol. 6, No #, pp. 27-40.

FDA, Medication Guide Xolair, (omalizumab), 2013, No Vol. pp. 1-2.

Biopharma, Sample Synagis, MedImmune, Inc., 2013, No Vol. pp. 1-19.

FDA Label—SYNAGIS® (PALIVIZUMAB)—1999, MedImmune, Inc., No. Vol. pp. 1-7.

Huang, Kelly et al., Respiratory Syncytial Virus-Neutralizing Monoclonal Antibodies Motavizumab and Palivizumab Inhibit Fusion, Journal of Virology, Aug. 2010, vol. 84, No. 16, pp. 8132-8140.

FDA Label—Vectibix® (panitumumab), Amgen Inc., 2006-2008, No Vol., pp. 1-13.

Grunwalk, Viktor et al., Developing Inhibitors of the Epidermal Growth Factor Receptor for Cancer Treatment, Journal of the National Cancer Institute, 2003, vol. 95, No. 12, pp. 851-867.

Yang, Xiao-Dong et al., Eradication of Established Tumors by a Fully Human Monoclonal Antibody to the Epidermal Growth Factor Receptor without Concomitant chemotherapy, Cancer Research, 1999, vol. 59, No. #, pp. 1236-1243.

Yang, Xiao-Dong et al., Development of ABX-EGF, A Fully Human anti-EGF Receptor Monoclonal Antibody, for Cancer Therapy, Oncology Hematology, 2001, vol. 38, No. #, pp. 17-23.

FDA, Highlights of Prescribing Information LUCENTIS(ranibizumab injection), Genentech, Inc., 2006, No Vol., pp. 1-9.

Binder, Mascha et al., The Epitope Recognized by Rituximab, Blood, 2006, vol. 108, No. 6, pp. 1975-1978.

FDA Label, Rituxan, Rituximab, IDEC Pharmaceuticals Corporation and Genetech, Inc., No Vol #, pp. 1-2.

FDA Label, ACTEMRA (tocilizumab), Risk Evaluation and Mitigation Strategy (REMS) 2013, Genentech, Inc., Reference ID: 3394610, No Vol. #, pp. 1-53.

FDA Label, BEXXAR, Tositumomab and Iodine I 131 Tositumomab 2003, Corixa Corp. and GlaxoSmithKline, No Vol #, pp. 1-49.

Srinivasan, A. et al., Tositumomab and Iodine I 131 Tositumomab Bexaar, Pharmacology Vignette, 2011, vol. 32, No #, pp. 637-638. FDA Guide, HERCEPTIN (trastuzumab), Highlights of Prescribing Information, 2010, Genentech, Inc., pp. 1-33.

European Public Assessment Report (EPAR), REMOVAB, European Medicines Agency, 2009, No Vol. #pp. 1-2.

Ruf, P. et al., Characterization of the New EpCAM-specific antibody HO-3: Implications for Trifunctional Antibody Immunotherapy of Cancer, British Journal of Cancer, 2007, vol. 97, No. 3, pp. 351 321.

Chelius, Dirk et al., Structural and functional characterization of the trifunctional antibody catumaxomab, mAbs, 2010, vol. 2 No. 3, pp. 309-319

Linke, Rolf et al., Catumazomab Clinical Development and Future Directions, Landes Bioscience, mAbs, vol. 2, No. 2, pp. 129-136. McLean, Leon et al., Vedolizumab for the treatment of ulcerative colitis and Crohn's disease, Immunotherapy, 2012, vol. 4, No. 9, pp. 883-898.

Reichert, Janice M. et al., Which Are the Antibodies to Watch in 2013, mAbs, 2013, vol. 5, No. 1, pp. 1-4.

Rob C. et al., IgG4 Breaking the Rules, Immunology, 2002, vol. 105, No #, pp. 9-19.

Alexandrakis, Michael et al., Relationship Between Circulating BAFF Serum Levels with Proliferating Markers in Patients with Multiple Myeloma, Biomed Research International, 2013, vol. 2013, Article ID. 389579, pp. 1-7.

Alfonso, Mauro et al., An Anti-Idiotype Vaccine Elicits a Specific Response to N-Glycolyl Sialic Acid Residues of Glycoconjugates in Melanoma Patients, The Journal of Immunology, 2002, vol. 168, No #, pp. 3523-2529.

Alonso, Ruby et al., Towards the Definition of a Chimpanzee and Human Conserved CD6 Domain 1 Epitope Recognized by T1 Monoclonal Antibody, Hybridoma, 2008, vol. 27, No. 4, pp. 291-301

Alprolix, Highlights of Prescribing Information, Full Prescribing Information, Biogen Idec, 2013, No Vol, pp. 1-19.

David McAuley, Pharm.D., Alzheimer's Disease—Therapeutic agents, 2012, No Vol. #, pp. 1-3.

Angevin, Eric et al., A Phase I/II, Multiple-Dose, Dose-Escalation Study of Siltuximab, an Anti-Interleukin-6 Monoclonal Antibody, in Patients with Advanced Solid Tumors, Clinical Cancer Research, 2014, vol. 20, No. 8, pp. 1-14.

Micromedex, Antihemophilic Factor Viii and Von Willebrand Factor Complex (Intravenous Route), Mayo Clinic, No. Vol #, pp. 1-3.

OTHER PUBLICATIONS

Armstrong, Deborah, et al., Farletuzumab (MORAb-003) in platinum-sensitive ovarian cancer patients experiencing a first relapse, Community Oncology, 2010, vol. 7, No. 2, Supp 1., pp. 1-4.

Baeten, Dominique et al., Anti-interleukin-17A monoclonal anti-body secukinumab in treatment of ankylosing spondylitis: a randomised, double-blind, placebo-controlled trial, The Lancet, 2013, vol. 382, No #, pp. 1705-1713.

Bai, D.L. et al., Huperzine A, A Potential Therapeutic Agent for Treatment of Alzheimer's Disease, Current Medicinal Chemistry, 2000, vol. 7, No. 3, pp. 355-374.

Ballatore, Carlo et al., Microtubule Stabilizing Agents as Potential Treatment for Alzheimer's Disease and Related Neurodegenerative Tauopathies, J. Med Chem., 2012, vol. 55, No. 21, pp. 8979-8996. Barker, Edward, et al., Effect of a Chimeric Anti-Ganglioside GD2 Antibody on Cell-mediated Lysis of Human Neuroblastoma Cells, Cancer Researchm, 1991, vol. 51, No. #, pp. 144-149.

Bamias, Giorgos, et al., Leukocyte Traffic Blockage in Inflammatory Bowel Disease, Current Drug Targets, 2013, vol. 14, No. 12, pp. 1490-1500.

Blom, Dirk J. et al., A 52-Week Placebo-Controlled Trial of Evolocumab in Hyperlipidemia, The New England Journal of Medicine, 2014, No. Vol #, pp. 1-11.

Bococizumab, Statement on a Nonproprietary Name Adopted by the USAN Council, 2013, No Vol. pp. 1-2.

Bohrmann, Bernd et al., Gantenerumab: A Novel Human Anti-Aβ Antibody Demonstrates Sustained Cerebral Amyloid-β Binding and Elicits Cell-Mediated Removal of Human Amyloid-β, Journal of Alzheimer's Disease, 2012, vol. 28, No. #, pp. 49-69.

Borghaei, Hossein et al., Phase I Dose Escalation, Pharmacokinetic and Pharmacodynamic Study of Naptumomab Estafenatox Alone in Patients With Advanced Cancer and With Docetaxel in Patients With Advanced Non-Small-Cell Lung Cancer, Journal of Clinical Oncology, 2009, vol. 27, No. 25, pp. 4116-4123.

Bottero, Federica et al., GeneTransfection and Expression of the Ovarian Carcinoma Marker Folate Binding Protein on NIH/3T3 Cells Increases Cell Growth in Vitro and in Vivo, Cancer Research, 1993, vol. 53, No. #, pp. 5791-5796.

Bousquet, Jean MD et al, Eosinophilic Inflammation in Asthma, The New England Journal of Medicine, 1990, vol. 323, No. 15, pp. 1033-1039.

Burgess, Teresa et al., Biochemical Characterization of AMG 102: A Neutralizing, Fully Human Monoclonal Antibody to Human and Nonhuman Primate Hepatocyte Growth Factor, Molecular Cancer Therapeutics, 2010, vol. 9, No. 2, pp. 400-409.

Busse, William W. et al., Safety profile, pharmacokinetics, and biologic activity of MEDI-563, an anti-IL-5 receptor a antibody, in a phase I study of subjects with mild asthma, J Allergy Clin Immunol, 2010, vol. 125, No. 6, pp. 1237-1244.

Carnahan, Josette et al., Epratuzumab, a Humanized Monoclonal Antibody Targeting CD22 Characterization of in Vitro Properties, Clinical Cancer Research, 2009, vol. 9, No. #, pp. 1-8.

Castro, Mario et al., Reslizumab for Poorly Controlled, Eosinophilic Asthma, A Randomized, Placebo-controlled Study, American Journal of Respiratory and Critical Care Medicine, 2011, vol. 184, No#, pp. 1125-1132.

Cavelti-Weder, Claudia et al., Effects of Gevokizumab on Glycemia and Inflammatory Markers in Type 2 Diabetes, Diabetes Care, 2012, vol. 35, No number, pp. 1654-1662.

Chou, Hsun-Hua et al., A mutation in human CMP-sialic acid hydroxylase occurred after the Homo-Pan divergence, Proc. Natl. Acad. Sci. USA,1998, vol. 95, No #, pp. 11751-11756.

Grundy, Scott et al., Promise of Low-Density Lipoprotein-Lowering Therapy for Primary and Secondary Prevention, Circulation Journal of the American Heart Association, 2008, vol. 117, No #, pp. 569-573

Raal, Frederick et al., Low-Density Lipoprotein Cholesterol-Lowering Effects of AMG 145, a Monoclonal Antibody to Proprotein Convertase Subtilisin/Kexin Type 9 Serine Protease in Patients With Heterozygous Familial Hypercholesterolemia: The Reduction of LDL-C With PCSK9 Inhibition in Heterozygous Familial Hypercholesterolemia Disorder (Rutherford) Randomized Trial, Circulation, 2012, vol. 126, pp. 2408-2417.

Roche Pharma AG, A Study to Evaluate Two Doses of Ocrelizumab in Patients With Active Systemic Lupus Erythematosus (BEGIN), ClinicalTrials.gov, Apr. 1, 2014, No Vol #, http://clinicaltrials.gov/ct2/show/NCT00539838, pp. 1-4.

Genentech, A Study of the Efficacy and Safety of Ocrelizumab in Patients With Relapsing-Remitting Multiple Sclerosis, ClinicalTrials.gov, Apr. 1, 2014, http://clinicaltrials.gov/ct2/show/NCT00676715, pp. 1-3.

Morphotek, Efficacy and Safety of MORAb-003 in Subjects With Platinum-sensitive Ovarian Cancer in First Relapse, ClinicalTrials.gov, Apr. 2, 2014, http://clinicaltrials.gov/ct2/show/NCT00849667?term=Farletuzumab&rank=4&submit_fld_opt, pp. 1.3

Roche Pharma AG, A Study to Investigate the Efficacy and Safety of Bendamustine Compared With Bendamustine +RO5072759 (GA101) in Patients With Rituximab-Refractory, Indolent Non-Hodgkin's Lymphoma (GADOLIN), ClinicalTrials.gov, Apr. 2, 2014, http://clinicaltrials.gov/ct2/show/NCT01059630?term=Obinutuzumab&rank=20&submit_fld_opt, pp. 1-3.

Eli Lilly and Company, A Study of Ramucirumab (IMC-1121B) Drug Product (DP) and Best Supportive Care (BSC) Versus Placebo and BSC as 2nd-Line Treatment in Patients With Hepatocellular Carcinoma After 1st-Line Therapy With Sorafenib (REACH), ClinicalTrials.gov, Apr. 2, 2014, http://clinicaltrials.gov/ct2/show/NCT01140347?term=ramucirumab&rank=12&submit_fld_opt, pp. 1-4.

Eli Lilly and Company, A Study of Chemotherapy and Ramucirumab vs. Chemotherapy Alone in Second Line Non-small Cell Lung Cancer Participants Who Received Prior First Line Platinum Based Chemotherapy, ClinicalTrials.gov, Apr. 2, 2014, http://clinicaltrials.gov/ct2/show/

NCT01168973?term=ramucirumab&rank=2&submit_fld_opt, pp. 1-4.

Eli Lilly and Company, A Study of Paclitaxel With or Without Ramucirumab in Metastatic Gastric Adenocarcinoma, ClinicalTrials.gov, Apr. 2, 2014, http://clinicaltrials.gov/ct2/show/NCT01170663?term=ramucirumab&rank=5&submit_fld_opt, pp. 1-4.

Eli Lilly and Company, A Study in Second Line Metastatic Colorectal Cancer, ClinicalTrials.gov, Apr. 2, 2014, http://clinicaltrials.gov/ct2/show/NCT01183780?term=ramucirumab&rank=20&submit_fld_opt., pp. 1-4.

Hoffmann-La Roche, A Study of Obinutuzumab (RO5072759) in Combination With CHOP Chemotherapy Versus MabThera/Rituxan (Rituximab) With CHOP in Patients With CD20-Positive Diffuse Large B-Cell Lymphoma (GOYA), ClinicalTrials.gov, Apr. 2, 2014, http://clinicaltrials.gov/ct2/show/

NCT01287741?term=Obinutuzumab&rank=13&submit_fld_opt, pp. 1-3.

Hoffmann-La Roche, A Study of Obinutuzumab (RO5072759) Plus Chemotherapy in Comparison With MabThera/Rituxan (Rituximab) Plus Chemotherapy Followed by GA101 or MabThera/Rituxan Maintenance in Patients With Untreated Advanced Indolent Non-Hodgkin's Lymphoma (GALLIUM), ClinicalTrials.gov, Apr. 2, 2014, http://clinicaltrials.gov/ct2/show/NCT01332968, pp. 1-3.

Avid Radiopharmaceuticals, Dominantly Inherited Alzheimer Network Trial: An Opportunity to Prevent Dementia. A Study of Potential Disease Modifying Treatments in Individuals at Risk for or With a Type of Early Onset Alzheimer's Disease Caused by a Genetic Mutation. (DIAN-TU), ClinicalTrials.gov, Apr. 2, 2014, http://clinicaltrials.gov/ct2/show/NCT01760005, pp. 1-5.

Eli Lilly and Company, Progress of Mild Alzheimer's Disease in Participants on Solanezumab Versus Placebo (Expedition 3), ClinicalTrials.gov, Apr. 2, 2014, http://clinicaltrials.gov/ct2/show/NCT01900665, pp. 1-3.

Eli Lilly and Company, Clinical Trial of Solanezumab for Older Individuals Who May be at Risk for Memory Loss (A4), ClinicalTrials.gov, Apr. 2, 2014, http://clinicaltrials.gov/ct2/show/NCT02008357, pp. 1-3.

OTHER PUBLICATIONS

Cohen, Idan et al., Differential release of chromatin-bound IL-1a Discriminates Between Necrotic and Apoptotic Cell Death by the Ability to Induce Sterile Inflammation, PNAS, 2010, vol. 107, No. 6, pp. 2574-2579.

Conde, Francisco et al., The Aspergillus toxin restrictocin is a suitable cytotoxic agent for generation of immunoconjugates with monoclonal antibodies directed against human carcinoma cells, Eur. J. Biochem, 1989, vol. 178, No #, pp. 795-802.

Coney, Leslie et al., Cloning of Tumor-associated Antigen: MOv18 and MOv19 Antibodies Recognize a Folate-binding Protein, Cancer Research, 1991, vol. 51, No #, pp. 6125-6132.

Corren, Jonathan et al., Lebrikizumab Treatment in Adults with Asthma, The New England Journal of Medicine, 2011, vol. 365, No. 12, pp. 1088-1098.

Daridon, Capucine et al., Epratuzumab Affects B Cells Trafficking in Systemic Lupus Erythematosus, Ann Rheum Dis, 2011, vol. 70, No #, pp. 1-2.

Devine, Peter L. et al., The Breast Tumor-associated Epitope Defined by Monoclonal Antibody 3E1.2 Is an O-linked Mucin Carbohydrate Containing N-Glycolylneuraminic Acid, Cancer Research, 1991, vol. 51, No #, pp. 5826-5836.

DiJoseph, John F. et al., Antibody-targeted chemotherapy with CMC-544: a CD22-targeted immunoconjugate of calicheamicin for the treatment of B-lymphoid malignancies, Blood, 2004, vol. 103, No #, pp. 1807-1814.

Dodart, Jean-Cosme et al., Immunization reverses memory deficits without reducing brain A burden in Alzheimer's disease model, Nature Neuroscience, 2002, vol. 5, No. 5, pp. 452-457.

Doody, Rachelle S. et al., Phase 3 Trials of Solanezumab for Mild-to-Moderate Alzheimer's Disease, NEJM Journal Watch, Apr. 2, 2014, No vol. No #, http://www.nejm.org/doi/full/10.1056/NEJMoa1312889, pp. 1-2.

National Cancer Institute, Drugs Approved for Ovarian Cancer, Aug. 16, 2013, No Vol.,pp. 1-2.

Dumont, Jennifer A. et al., Prolonged activity of a recombinant factor VIII-Fc fusion protein in hemophilia A mice and dogs, Blood, 2012, vol. 119, No. #, pp. 3024-3030.

Ebel, Wolfgang et al, Preclinical Evaluation of MORAb-003, a Humanized Monoclonal Antibody Antagonizing Folate Receptoralpha, Cancer Immunity, 2007, vol. 7 No. #, pp. 1-8.

Eisen, Tim et al., Naptumomab Estafenatox: Targeted Immunotherapy with a Novel Immunotoxin, Curr Oncol Rep, 2014, vol. 16, N. 370 pp. 2-6.

Erlandsson, Eva et al., Identification of the Antigenic Epitopes in Staphylococcal Enterotocins A and E and Design of A Superantigen for Human Cancer Therapy, J. Mol. Biol., 2003, vol. 333, No #, pp. 893-905.

Mayo Clinic, Factor Ix Complex (Intravenous Route, Injection Route) Description and Brand Names—Drugs and Supplements, http://www.mayoclinic.org/drugs-supplements/factor-ix-complex-intravenous-route-injection-route/description/drg-20063804, Apr. 1, 2014, No Vol., pp. 1-3.

Ferrara, Claudia et al., Unique carbohydrate-carbohydrate interactions are required for high affinity binding between FcγRIII and antibodies lacking core fucose, PNAS, 2011, No Vo. #, pp. 1-6. Figini, M. et al., Reversion of transformed phenotype in ovarian cancer cells by intracellular expression of anti folate receptor

antibodies, Gene Therapy, 2003 vol. 10, No #, pp. 1018-1025. Vasquez, Ana et al., Racotumomab: an anti-idiotype vaccine related to N-Glycolyl-containing gangliosides-preclinical and clinical date, Frontiers in Oncology, 2012, vol. 2, Article 150, pp. 1-6.

Forsberg, G. et al., Therapy of Human Non-Small-Cell Lung Carcinoma Using Antibody Targeting of a Modified Superantigen, British Journal of Cancer, 2001, vol. 85, No. 1, pp. 129-136.

Forsberg, G et al., Naptumomab Estafentoz, an Engineered Antibody-superantigen Fusion Protien with Low Toxicity and Reduced Antigenicity, J Immunother, 2010, vol. 33, No. 5, pp. 492-499. Feagan, Brian et al., Vedolizumab as Induction and Maintenance Therapy for Ulcerative Colitis, The New England Journal of Medicine, 2013, vol. 369, No. 8, pp. 699-710.

Furie, Richard et al., A Phase III, Randomized, Placebo-Controlled Study of Belimumab, a Monoclonal Antibody That Inhibits B Lymphocyte Stimulator, in Patients With Systemic Lupus Erythematosus, Arthritis & Rheumatism, 2011, vol. 63, No. 12, pp. 3918.3930.

Garcia, Gilles et al., Anti-interleukin-5 Therapy in Serve Asthma, Rare Diseases and Orphan Drugs, 2013, vol. 22, No. #, pp. 251-257. Garin-Chesa, Pilar et al., Trophoblast and Ovarian Cancer Antigen LK26, American Journal of Pathology, 1993, vol. 142, No. 2, pp. 557-567

Genovese, Mark C et al., Efficacy and safety of secukinumab in patients with rheumatoid arthritis: a phase II, dose-finding, double-blind, randomised, placebo controlled study, Ann Rheum Dis, 2013; vol. 72, No #, pp. 863-869.

Genovese, Mark C et al., A phase 2 dose-ranging study of subcutaneous tabalumab for the treatment of patients with active rheumatoid arthritis and an inadequate response to methotrexate, Ann Rheum Dis 2013; vol. 72, No#, pp. 1453-1460.

Genovese, Mark C et al., Ocrelizumab, a Humanized Anti-CD20 Monoclonal Antibody, in the Treatment of Patients With Rheumatoid Arthritis, Arthritis & Rheumatism, 2008, vol. 58, No. 9, pp. 2652-2661.

Gevaert, Philippe, et al., Mepolizumab, a humanized anti-IL-5 mAb, as a treatment option for severe nasal polyposis, Rhinitis, sinusitis, and upper airway disease, J Allergy Clin Immunol, 2011, vol. 128, No. 5, pp. 989-995.

Ghazi, Aasia et al., Benralizumab—a humanized mAb to IL-5Rα with enhanced antibody-dependent cell-mediated cytotoxicity—a novel approach for the treatment of asthma, Expert Opin Biol Ther. 2012, vol. 12, No. 1, pp. 113-118.

Gillies, Stephen et al., Antibody-targeted interleukin 2 stimulates T-cell killing of Autologous Tumor Cells, Proc. Natl. Acad. Sci., 1992, vol. 89, No #, pp. 1428-1432.

Grant, Ryan W. et al., Mechanisms of disease: inflammasome activation and the development of type 2 diabetes, Frontiers in Immunology, 2013, vol. 4, Article 50, pp. 1-10.

Greenfeder, Scott et al., Th2 cytokines and asthma the role of interleukin-5 in allergic eosinophilic disease, Respiratory Research, 2001, vol. 2, No. 2, pp. 71-79.

Grünig, Gabriele et al., Interleukin 13 and the evolution of asthma therapy, Am J Clin Exp Immunol, 2012;vol. 1, No. 1, pp. 20-27. Hamid, Omid et al., Safety and Tumor Responses with Lambrolizumab (Anti-PD-1) in Melanoma, The New England Journal of Medicine, 2013, vol. 369, No. 2, pp. 134-144.

Hank, Jacquelyn, et al., Immunogenicity of the Hu14.18-IL2 Immunocytokine Molecule in Adults With Melanoma and Children With Neuroblastoma, Clinical Cancer Research, 2009, vol. 15, No. 18, pp. 5923-5930.

Hart, Timothy K. et al., Preclinical efficacy and safety of mepolizumab (SB-240563), a humanized monoclonal antibody to IL-5, in cynomolgus monkeys, J Allergy Clin Immunol, 2001, vol. 108, No. 2, pp. 250-257.

Hedlund, Gunnar et al., The Tumor Targeted Superantigen ABR-217620 Selectively Engages TRBV7-9 and Exploits TCR-pMHC Affinity Mimicry in Mediating T Cell Cytotoxicity, PLOS One, 2013, vol. 8, Issue 10, pp. 1-17.

Hernández, Ana María et al., Anti-NeuGcGM3 Antibodies, Actively Elicited by Idiotypic Vaccination in Nonsmall Cell Lung Cancer Patients, Induce Tumor Cell Death by an Oncosis-Like Mechanism, The Journal of Immunology, 2011, vol. 186, No #, pp. 3735-3744. Humbert, Marc et al., Relationship between IL-4 and IL-5 mRNA Expression and Disease Severity in Atopic Asthma, Am J Respir Crit Care Med, 1997, vol. 156, No #, pp. 704-708.

Hole, N. et al., A 72 kD trophoblast glycoprotein defined by a monoclonal antibody, Br. J. Cancer 1988,vol. 57, No. #, pp. 239-246

Huizinga, Tom W J et al., Sarilumab, a fully human monoclonal antibody against IL-6R α in patients with rheumatoid arthritis and an inadequate response to methotrexate: efficacy and safety results

OTHER PUBLICATIONS

from the randomized SARIL-RA-MOBILITY Part A trial, Ann Rheum Dis, 2013; No Vol. pp. 1-9.

Imbimbo, Bruno P et al., Solanezumab for the treatment of mild-to-moderate Alzheimer's disease, Expert Rev. Clin. Immunol., 2012, vol. 8, No. 2, pp. 135-149.

Ito, Asahi et al., Defucosylated anti-CCR4 monoclonal antibody exercises potent ADCC-mediated antitumor eVect in the novel tumor-bearing humanized NOD/Shi-scid, IL-2R_null mouse model, Cancer Immunol Immunother, 2009, vol. 58, No #, pp. 1195-1206.

Winkler, David G. et al., Noggin and Sclerostin Bone Morphogenetic Protein Antagonists Form a Mutually Inhibitory Complex, J. Biol. Chem., 2004, vol. 279, pp. 36293-36298.

Janssens, Ann et al., Rixuximab for Chronic Lymphocytic Leukemia in Treatment-Naive and Treatment-Experienced, OneLive, Bringing Oncology Together, Apr. 2, 2014, No Vol., pp. 1-7.

Jia, Guiquan et al., Periostin is a systemic biomarker of eosinophilic airway inflammation in asthmatic patients, J Allergy Clin Immunol, 2012, vol. 130, No. 3, pp. 647-654.

Jin, Wei et al., IL-17 cytokines in immunity and inflammation, Emerging Microbes and Infections, 2013, vol. 2, No. #, pp. 1-5. Kappos, Ludwig, et al., Ocrelizumab in relapsing-remitting multiple sclerosis: a phase 2, randomised, placebo-controlled, multicentre trial, The Lancet, 2011, vol. 378, Issue 9805, pp. 1779-1787.

Kaur, Sukhwinder et al., Mucins in pancreatic cancer and its microenvironment, Nature Reviews, 2013, No Vol., pp. 1-14. Kausar, Fariha et al., Ocrelizumab: A Step Forward in the Evolution

Kausar, Fariha et al., Occelizumab: A Step Forward in the Evolution of B-Cell Therapy, Expert Opinion Biol. Ther., 2009, vol. 9, No. 7, pp. 889-895.

Kim, Busun et al., The Interleukin-1a precursor is Biologically Active and Is Likely a Key Alarmin in the IL-1 Family of Cytokines, Frontiers in Immunology, 2013, vol. 4, Article 391, pp. 1-9.

Kips, Johan et al., Effect of SCH55700, a Humanized Anti-Human Interleukin-5 Antibody, in Severe Persistent Asthma, American Journal of Respiratory and Critical Care Medicine, Safety of Anti-IL-5 in Asthma, vol. 167, pp. 1655-1659, 2003.

Koenigsknecht-Talboo, Jessica et al., Rapid Microglial Response Around Amyloid Pathology after Systemic Anti-A_Antibody Administration in PDAPP Mice, The Journal of Neuroscience, 2008, vol. 28, No. 52, pp. 14156-1414.

Kolbeck, Roland et al., MEDI-563, a humanized anti-IL-5 receptor a mAb with enhanced antibody-dependent cell-mediated cytotoxicity function, J Allergy Clin Immunol, vol. 125, No. 6, pp. 1344-1353.

Koren, Michel J. et al., Efficacy and Safety of Longer-Term Administration of Evolocumab (AMG 145) in Patients With Hypercholesterolemia: 52-Week Results From the Open-Label Study of Long-Term Evaluation Against LDL-C (OSLER) Randomized Trial, Circulation, 2013, No Vol., pp. 1-20.

Kreitman, Robert J. et al., Antibody Fusion Proteins: Anti-CD22 Recombinant Immunotoxin Moxetumomab Pasudotox, Clinical Cancer Research, 2011, vol. 17, No #, pp. 6398-6405.

Kreitman, Robert J. et al., Phase I Trial of Anti-CD22 Recombinant Immunotoxin Moxetumomab Pasudotox (CAT-8015 or HA22) in Patients With Hairy Cell Leukemia, Journal of Clinical Oncology, 2012, vol. 30, No. 15, pp. 1822-1826.

Krueger, Gerald G. et al., A Human Interleukin-12/23 Monoclonal Antibody for the Treatment of Psoriasis, The New England Journal of Medicine, 2007,vol. 356, No. 6, pp. 580-592.

Kuenen, Bart et al., A Phase I Pharmacologic Study of Necitumumab (IMC-11F8), a Fully Human IgG 1 Monoclonal Antibody Directed Against EGFR in Patients with Advanced Solid Malignancies, Clinical Cancer Research, 2010, vol. 16, No #, pp. 1915-1923.

Kuijpers, Taco W. et al., CD20 deficiency in humans results in impaired T cell—independent antibody responses, The Journal of Clinical Investigation, 2010, vol. 120, No. 1, pp. 214-222.

Kurzrock, Razelle et al., A Phase I, Open-Label Study of Siltuximab, an Anti-IL-6 Monoclonal Antibody, in Patients with

B-cell Non-Hodgkin Lymphoma, Multiple Myeloma, or Castleman Disease, Clinical Cancer Research, 2013, vol. 19, No #, pp. 3659-3670.

Lach-Trifilieff, Estelle et al., An Antibody Blocking Activin Type II Hypertrophy and Protects from Atrophy Receptors Induces Strong Skeletal Muscle, Molecular and Cellular Biology, 2004, vol. 34, No. 4, pp. 606-618.

Legleiter, Justin et al., Effect of Different Anti-A β Antibodies on A β Fibrillogenesis as AAssessed by Atomic Force Microscopy, J. Mol. Biol, 2004, vol. 335, No #, pp. 997-1006.

Leonard, JP et al., Preclinical and clinical evaluation of epratuzumab (anti-CD22 IgG) in B-cell malignancies, Oncogene, 2007, vol. 26 No #, pp. 3704-3713.

Leonardi, Craig et al., Anti-Interleukin-17 Monoclonal Antibody Ixekizumab in Chronic Plaque Psoriasis, The New England Journal of Medicine, 2012, vol. 366, No. 13, pp. 1190-1199.

Lindén, Ola, et al., Dose-Fractionated Radioimmunotherapy in Non-Hodgkin's Lymphoma Using DOTA-Conjugated, 90Y-Radiolabeled, Humanized Anti-CD22 Monoclonal Antibody, Epratuzumab, Clinical Cancer Research, 2005, vol. 11, No #, pp. 5215-5222.

Braun, Stephen et al., Preclinical Studies of Lymphocyte Gene Therapy for Mild Hunter Syndrome (Mucopolysaccharidosis Type II), Human Gene Therapy, 1996, vol. 7, pp. 283-290.

Liu, Alvin et al, Production of a Mouse-Human Chimeric Monoclonal Antibody to CD20 With Potent Fc-Dependent Biological Activity, The Journal of Immunology, 1987,vol. 139, No. 10, pp. 3521-3526.

Lonial, Sagar, et al., Elotuzumab in Combination With Lenalidomide and Low-Dose Dexamethasone in Relapsed or Refractory Multiple Myeloma, Journal of Clinical Oncology, 2012, vol. 30, No. 16, pp. 1953-1959.

Lu, Dan et al., Tailoring in Vitro Selection for a Picomolar Affinity Human Antibody Directed against Vascular Endothelial Growth Factor Receptor 2 for Enhanced Neutralizing Activity, The Journal of Biological Chemistry, 2003, vol. 278, No. 44, pp. 43496-43507. Lubberts, Erik et al., Treatment With a Neutralizing Anti-Murine Interleukin-17 Antibody After the Onset of Collagen-Induced Arthritis Reduces Joint Inflammation, Cartilage Destruction, and Cone Erosion, Arthritis & Rheumatism, 2004, vol. 50, No. 2, pp. 650-659

MacLean, Catherine et al., Ststematic Review: Comparative Effectiveness of Treatments to Prevent Fractures in Men and Women with Low Bone Density or Osteoporosis, Annals of Internal Medicine, 2008, vol. 148, No. 3, pp. 197-217.

Marquina, Gilda et al., Gangliosides Expressed in Human Breast Cancer, Cancer Res, 1996; vol. 56, No #, pp. 5165-5171.

Matsue, Hiroyuki et al., Folate receptor allows cells to grow in low concentrations of 5-methyltetrahydrofolate, Proc. Natl. Acad. Sci. USA, Cell Biology, 1992, vol. 89, No #, pp. 6006-6009.

McInnes, Iain B et al., Efficacy and safety of secukinumab, a fully human anti-interleukin-17A monoclonal antibody, in patients with moderate-to-severe psoriatic arthritis: a 24-week, randomised, double-blind, placebo-controlled, phase II proof-of-concept trial, Ann Rheum Dis, 2014; vol. 73, No. #, pp. 349-356.

McKenney, James M. et al., Safety and Efficacy of a Monoclonal Antibody to Proprotein Convertase Subtilisin/GKexin Type 9 Serine Protease, SAR236553/REGN727, in Patients With Primary Hypercholesterolemia Receiving Ongoing Stable Atorvastatin Therapy, Journal of the American College of Cardiology, 2012, vol. 59, No. 25, pp. 2344-2353.

Di Meglio, Paola et al., The role of IL-23 in the immunopathogenesis of psoriasis, Biology Reports, 2010, vol. 2, No. 40, pp. 1-5.

Merelli, Barbara et al., Targeting the PD1/PD-L1 axis in melanoma: Biological rationale, clinical challenges and opportunities, Critical Reviews in Oncology/Hematology, 2014, vol. 89, No #, pp. 140-165

Moreaux, Jérôme et al., BAFF and APRIL protect myeloma cells from apoptosis induced by interleukin 6 deprivation and dexamethasone, Blood, 2004, vol. 103, No #, pp. 3148-3157.

Morgan, D., Immunotherapy for Alzheimer's disease, Journal of Internal Medicine, 2011, vol. 269, No #, pp. 54-63.

OTHER PUBLICATIONS

Mujoo, Kalpana et al., Disialoganglioside GD2 on Human Neuroblastoma Cells: Target Antigen for Monoclonal Antibodymediated Cytolysis and Suppression of Tumor Growth, Cancer Research, 1987, vol. 47, No #, 1098-1104.

Mujoo, Kalpana et al., Functional Properties and Effect on Growth Suppression of Human Neuroblastoma Tumors by Isotype Switch Variants of Monoclonal Antiganglioside GD2 Antibody 14.18, Cancer Research, 1989, vol. 49, No #, pp. 2857-2861.

Mössner, Ekkehard, Increasing the efficacy of CD20 antibody therapy through the and immune effector cell-mediated B-cell cytotoxicity engineering of a new type II anti-CD20 antibody with enhanced direct, Blood, 2010, vol. 115, No #, pp. 4393-4402.

Nair, P. et al., CD6 synergistic co-stimulation promoting proinflammatory response is modulated without interfering with the activated leucocyte cell adhesion molecule interaction, Clinical& Experimental Immunology, 2010, vol. 162, No #, pp. 116.130. Experimental Immunology, i_4235.

Neal, Zane C. et al., Enhanced Activity of Hu14.18-1L2 Immunocytokine against Murine NXS2 Neuroblastoma when Combined with Interleukin 2 Therapy, Clinical Cancer Research, 2004, vol. 10, pp. 4839-4847.

Neer, Robert M. et al., Effect of Parathyroid Hormone (1-34) on Fractures and Bone Mineral Density in Postmenopausal Women With Osteoporosis, The New England Journal of Medicine, 2001, vol. 344, No. 19, pp. 1434-1441.

Negrier, Claude et al., Enhanced pharmacokinetic properties of a glycoPEGylated recombinant factor IX: a first human dose trial in patients with hemophilia B, Blood, 2011, vol. 118, No #, pp. 2695-2701.

Neninger, Elia et al., Active Immunotherapy with 1E10 Anti-Idiotype Vaccine in Patients with Small Cell Lung Cancer, Cancer Biology & Therapy, 2007, vol. 6, No. 2., pp. 1-6.

Novakovic, Dijana et al., Profile of Gantenerumab and Its Potential in the Treatment of Alzheimer's Disease, Drug Design, Development and Therapy, 2013, vol. 7, No #, pp. 1359-1364.

Wright, Timothy M.D., Transforming Molecules into Breakthrough Therapies, Novartis, Investor Day, London, 2013, No Vol. pp. 1-16. Oldhoff et al., Anti-IL-5 recombinant Humanized Monoclonal Antibody (Mepolizumab) for the treatment of atopic dermatitis, Allergy, 2005, vol. 60, No # pp. 693-696.

Ostrowitzki, Susanne et al., Mechanism of Amyloid Removal in Patients with Alzheimer Disease Treated with Gantenerumab, Arch Neurol., 2012, vol. 69, No. 2, pp. 1-10.

Ottone, F. et al., Relationship Between folate-binding Protein Expression and Cisplatin Sensitivity in Ovarian Carcinoma Cell Lines, British Journal of Cancer, 1997, vol. 76, No. 1, pp. 77-82. Papp, KA et al., Anti-IL-17 Receptor Antibody AMG 827 Leads to Rapid Clinical Response in Subjects with Moderate to Severe Psoriasis: Results from a Phase I, Randomized, Placebo-Controlled Trial, Journal of Investigative Dermatology, 2012, vol. 132, No #, pp. 2466-2469.

Papp, Kim, et al., Brodalumab, an Anti-Interleukin-17-Receptor Antibody for Psoriasis, The New England Journal of Medicine, 2012, vol. 366, No. 13, pp. 1181-1189.

Papp, KA et al, Efficacy and safety of secukinumab in the treatment of moderate-to-severe plaque psoriasis: a randomized, double-blind, placebo-controlled phase II dose-ranging study, 2013,British Journal of Dermatology, vol. 168, No #, pp. 412-421.

Pasadhika, Sirichai et al., Update on the use of systemic biologic agents in the treatment of oninfectious uveitis, Biologics: Targets and Therapy, 2014, vol. 8 No #, pp. 67-81.

Pavord, Ian D et al., Mepolizumab for severe eosinophilic asthma (DREAM): a multicentre, double-blind, placebo-controlled trial, The Lancet, 2012, vol. 380, No Vol #, 2012, pp. 651-659.

Sanofi, Fact Sheet, PCSK9 and Alirocumab Backgrounder, Regeneron, 2013, No Vol. pp. 1-3.

Peters, R.T. et al., Biochemical and functional characterization of a recombinant monomeric factor VIII-Fc fusion protein, Journal of Thrombosis and Haemostasis, 2012, vol. 11, pp. 132-141.

Powell, Jerry S. et al., Safety and prolonged activity of recombinant factor VIII Fc fusion protein in hemophilia A patients, Blood, 2012, vol. 119, No #, pp. 3031-3037.

Prewett, Marie et al., Kinase 1) Monoclonal Antibody Inhibits Tumor Angiogenesis Antivascular Endothelial Growth Factor Receptor (Fetal Liver Kinase 1) Monoclonal Antibody Inhibits Tumor Angiogenesis and Growth of Several Mouse and Human Tumors, Cancer Res, 1999; vol. 59, No #, pp. 5209-5218.

Raal, Frederick et al., Elevated PCSK9 Levels in Untreated Patients With Heterozygous or Homozygous Familial Hypercholesterolemia and the Response to High-Dose Statin Therapy, Journal of the American Heart Association, 2013, No Vol., pp. 1-8.

Rich, PP. et al., Secukinumab induction and maintenance therapy in moderate-to-severe plaque psoriasis: a randomized, double-blind, placebo-controlled, phase II regimen-finding study, British Journal of Dermatology, Therapeutics, 2013, vol. 168, No #, pp. 402-411. Rossi, Edmund et al., Trogocytosis of Multiple B-cell Surface Markers by CD22 Targeting With Epratuzumab, Blood, 2013, vol. 122, No #, pp. 3020-3029.

Rossjohn, Jamie et al., Structure of the activation domain of the GM-CSF/IL-3/IL-5 receptor common β-chain bound to an antagonist, Blood, 2000, vol. 95, No #, pp. 2491-2498.

Roth, Eli M. et al., Atorvastatin with or without an Antibody to PCSK9 in Primary Hypercholesterolemia, The New England Journal of Medicine, 2012, vol. 367, vol. 20, pp. 1891-1900.

Roufosse, Florence E., et al., Long-term safety of mepolizumab for the treatment of hypereosinophilic syndromes, J Allergy Clin Immunol. 2013; vol. 131, No. 2, pp. 461-467.

Salles, Gilles et al., Phase 1 study results of the type II glycoengineered humanized lymphoma patients anti-CD20 monoclonal antibody obinutuzumab (GA101) in B-cell, Blood, 2012, vol. 119, No #., pp. 5126-5132.

Sandborn, William J. et al., Vedolizumab as Induction and Maintenance Therapy for Crohn's Disease, The New England Journal of Medicine, 2013, vol. 369, No. 8, pp. 711-721.

Schuelke, Markus M.D. et al., Myostatin Mutation Associated With Gross Muscle Hypertrophy in a Child, The New England Journal of Medicine, 2004, vol. 350, No. 26, pp. 2862-2688.

Shusterman, Suzanne et al., Antitumor Activity of Hu14.18-IL2 in Patients With Relapsed/Refractory Neuroblastoma: A Children's Oncology Group (COG) Phase II Study, Journal of Clinical Oncology, 2010, vol. 28, No. 33, pp. 4969-4975.

Hueber, Wolfgang et al., Effects of AIN457, a Fully Human Antibody to Interleukin-17A, on Psoriasis, Rheumatoid Arthritis, and Uveitis, Science Translational Medicine, 2010, vol. 2, Issue 52, pp. 1-9

Scursoni, Alejandra M. Et al., Detection of N-Glycolyl GM3 Ganglioside in Neuroectodermal Tumors by Immunohistochemistry: An Attractive Vaccine Target for Aggressive Pediatric Cancer, Clinical and Developmental Immunology, 2011, vol. 2011, Article ID., 245181, pp. 1-6.

Semënov, Mikhail et al., SOST Is a Ligand for LRP5/LRP6 and a Wnt Signaling Inhibitor, The Journal of Biological Chemistry, 2005, vol. 280, No. 29., pp. 26770-26775.

Shapiro, Amy D. et al., Recombinant factor IX-Fc fusion protein (rFIXFc) demonstrates safety and prolonged activity in a phase 1/2a study in hemophilia B patients, Blood, 2012, vol. 119, No #, pp. 666-672.

Sieger, N. et al., CD22 Ligation Inhibits Downstream B Cell Receptor Signaling and Ca2_Flux Upon Activation, Arthritis & Rheumatism, 2013, vol. 65, No. 3, pp. 770-779.

Simon, Thorsten et al., Consolidation Treatment With Chimeric Anti-GD2-Antibody ch14.18 in Children Older Than 1 Year With Metastatic Neuroblastoma, Journal of Clinical Oncology, 2004, vol. 22, No. 17, pp. 3549-3557.

Spratlin, Jennifer L. et al., Phase I Pharmacologic and Biologic Study of Ramucirumab (IMC-1121B), a Fully Human Immunoglobulin G1 Monoclonal Antibody Targeting the Vascular Endothelial Growth Factor Receptor-2, Journal of Clinical Oncology, 2010, vol. 28, No. 5, pp. 780-787.

Steinfield, Serge et al., Epratuzumab (humanized anti-CD22 anti-body) in autoimmune diseases, Expert Opinion, 2006, vol. 6, No. 9, pp. 943-949.

OTHER PUBLICATIONS

Stevenson, Frazier et al., The N-terminal propiece of interleukin Ia is a transforming nuclear oncoprotein, Proc. Natl. Acad. Sci. USA, 1997, vol. 94, No #, pp. 508-513.

William Stohl et al., Future prospects in biologic therapy for systemic lupus erythematosus, Nature Reviews, Rheumatology, No Vol., pp. 1-16.

Sullivan, David et al., Effect of a Monoclonal Antibody to PCSK9 on Low-Density Lipoprotein Cholesterol Levels in Statin-Intolerant Patients the GAUSS Randomized Trial, JAMA, 2012, vol. 308, No. 23, pp. 1-10.0.

Sun, Jian, et al., B lymphocyte stimulator: a new target for treating B cell malignancies, Chinese Medical Journal, 2008; vol. 12, No. 14, pp. 1319-1323.

Tanaka, Toshio et al., Targeting Interleukin-6: All the Way to Treat Autoimmune and Inflammatory Diseases, International Journal of Biological Sciences, 2012, vol. 8 No. 9, pp. 1227-1236.

Toffoli1, Giuseppe et al., Overexpression of Folate Binding Protein in Ovarian Cancers, 1997, Int. J. Cancer (Pred. Oncol.):vol. 74, No #, pp. 193-198.

Gevokizumab, Statement on a Nonproprietary Name Adopted by the USAN Council, No year no Volume p. 1.

Romosozumab, Statement on a Nonproprietary Name Adopted by the USAN Council, No Year, No Volume, p. 1.

van Bezooijen, Rutger L. et al., Sclerostin Is an Osteocyte-expressed Negative Regulator of Bone Formation, But Not a Classical BMP Antagonist, The Journal of Experimental Medicine, 2004, vol. 199, No. 6, pp. 805-814.

van Bezooijen, Rutger L et al., Wnt but Not BMP Signaling Is Involved in the Inhibitory Action of Sclerostin on BMP-Stimulated Bone Formation, Journal of Bone and Mineral Research, 2007, vol. 22, No. 1, pp. 1-10.

van Cruijsen, Hester et al., Tissue micro array analysis of ganglioside N-glycolyl GM3 expression and signal transducer and activator of transcription (STAT)-3 activation in relation to dendritic cell infiltration and microvessel density in non-small cell lung cancer, BMC Cancer, 2009, vol. 9, No. 180, pp. 1-9.

Wallace, Daniel J. et al., Epratuzumab Demonstrates Clinically Meaningful Improvements in Patients with Moderate to Severe Systemic Lupus Erythematosus (SLE) Results from EMBLEM, a Phase IIB Study, ACR Concurrent Abstract Sessions, Systemic Lupus Enrthematosus—Clinical Aspects and Treatment: New Therapies, 2010, No Vol., pp. 1452.

Wallace, Daniel J et al., Efficacy and safety of epratuzumab in patients with moderate/severe active systemic lupus erythematosus: results from EMBLEM, a phase IIb, randomised, double-blind, placebo-controlled, multicentre study, Ann Rheum Dis, 2014;vol. 73, No #, pp. 183-190.

Wechsler, Michael E. et al., Novel targeted therapies for eosinophilic disorders, J Allergy Clin Immunol., 2012; vol. 130, No. 3, pp. 563-571.

Werman, Ariel et al., The precursor form of IL-1_is an intracrine proinflammatory activator of transcription, PNAS, 2004, vol. 101, No. 8, pp. 2434-2439.

WHO Drug Information, International Nonproprietary Names for Pharmaceutical Substances (INN),2013, vol. 27, No. 4, pp. 1-60. WHO Drug Information, International Nonproprietary Names for Pharmaceutical Substances (INN), 2012, vol. 26, No. 4, pp. 1-71. WHO Drug Information, International Nonproprietary Names for Pharmaceutical Substances (INN), 2011, vol. 25, No. 3, pp. 1-46. WHO Drug Information, International Nonproprietary Names for Pharmaceutical Substances (INN), 2012, vol. 26, No. 2, pp. 1-79. WHO Drug Information, International Nonproprietary Names for Pharmaceutical Substances (INN), 2012, vol. 26, No. 3, pp. 1-36. Winkler, David G. et al. Osteocyte control of bone formation via sclerostin, a novel BMP antagonist, The EMBO Journal, 2003, vol. 22 No. 23 pp. 6267-6276.

Yang, Richard K. et al., Anti-GD2 Strategy in the Treatment of Neuroblastoma, Drugs Future, 2010; vol. 35, No. 8, pp. 1-15. Yu, Alice et al., Phase I Truak of a Human-Mouse Chimeric Ant-Disialoganglioside Monoclonal Antibody ch14.18 in Patients with Refractory Neuroblastoma, and Osteosarcoma, Journal of Clinical Oncology 1998, vol. 16, No. 6, pp. 2169-2180.

Zheng, Yue et al. Intracellular Interleukin-1 Receptor 2 Binding Prevents Cleavage and Activity of Interleukin-1a, Controlling Necrosis-Induced Sterile Inflammation, Immunity,2013, vol. 38, No #, pp. 285-295.

Zhu, Min et al., Population Pharmacokinetics of Rilotumumab, a Fully Human Monoclonal Antibody Against Hepatocyte Growth Factor, in Cancer Patients, Journal of Pharmaceutical Sciences, 2014, vol. 328 No #, pp. 328-336.

Zhu, Zhenping et al., Inhibition of Vascular Endothelial Growth Factor-induced Receptor Activation with Anti-Kinase Insert Domain-containing Receptor Single-Chain Antibodies from a Phage Display Library, Cancer Research, 1998, vol. 58, No # pp. 3209-3214.

Zhu, Z et al, Inhibition of human leukemia in an animal model with human antibodies directed against vascular endothelial growth factor receptor 2. Correlation between antibody affinity and biological activity, Leukemia, 2003), vol. 17, pp. 604-611.

Zia-Amirhosseini, P. et al., Pharmacokinetics and Pharmacodynamics of SB-240563, a Humanized Monoclonal Antibody Directed to Human Interleukin-5, in Monkeys, The Journal of Pharmacology and Experimental Therapeutics, 1999, vol. 291, No. 3, pp. 1060-1067.

Stockinger, Walter et al., The PX-domain Protein SNX17 Interacts With Members of the LDL Receptor Family and Modulates Endocytosis, The EMBO Journal, 2002, vol. 21, No. 16 pp. 4259-4267. Sorrentino, Vincenzo et al., Post-transcriptional regulation of lipoprotein receptors by the E3-ubiquitin ligase inducible degrader of the low-density lipoprotein receptor, Current Opinion, 2012, vol. 23, No. 3, pp. 213-219.

Zelcer, Noam et al., LXR Regulates Cholesterol Uptake through Idol-dependent Ubiquitination of the LDL Receptor, Science, 2009; vol. 325, No. 5936, pp. 100-104.

Zhang , Li et al, Both K63 and K48 ubiquitin linkages signal lysosomal degradation of the LDL receptor, Journal of Lipid Research, 2013, vol. 54, No #, pp. 1410-1420.

Lozier, Jay N , Factor IX Padua: them that have, give , Blood, 2012, vol. 120, No #, pp. 4452-4453.

Simioni, Paolo et al., X-Linked Thrombophilia with a Mutant Factor IX (Factor IX Padua), The New England Journal of Medicine, 2009, vol. 361, No. 17, pp. 1671-1675.

Cornett, Jeff et al. Update of Clinicla Trials to Cure Hemophilia, Hemophilia of Georgia, Dec. 12, 2013, No Vol. pp. 1-2.

Raschke, Silja et al., Adipo-Myokines: Two Sides of the Same Coin—Mediators of Inflammation and Mediators of Exercise, Mediators of Inflammation, 2013, vol. 2013, Article ID 320724, pp. 1-16.

Podbregar, Matej et al., Cytokine Response of Cultured Skeletal Muscle Cells Stimulated with Proinflammatory Factors Depends on Differentiation Stage, The Scientific World Journal, 2013, vol. 2013, Article ID 617170, pp. 1-8.

Guerrero-Ca' zares, Hugo et al. Biodegradable Polymeric Nanoparticles Show High Efficacy and Specificity at DNA Delivery to Human Glioblastoma in Vitro and in Vivo, ACS Nano, 2014, No Vol., No #, pp. 1-14.

Dahlman, James E. et al., In vivo endothelial siRNA delivery using polymeric nanoparticles with low molecular weight, Nature Nanotechnology, 2014, No Vol. #, pp. 1-8.

Kozielski, Kristen L. et al., Bioreducible Cationic Polymer-Based Nanoparticles for Efficient and Environmentally Triggered Cytoplasmic siRNA Delivery to Primary Human Brain Cancer Cells, ACS Nano, 2014, vol. 8, 'No. 4', pp. 3232-3241.

M. Kanapathipillai, et al., Nanoparticle targeting of anti-cancer drugs that alter intracellular signaling or influence the tumor microenvironment, Adv. Drug Deliv. Rev. (2014), , pp. 1-12.

Seldin, Marcus M. et al., Regulation of tissue crosstalk by skeletal muscle-derived myonectin and other myokines, Adipocyte, 2012, vol. 1, No. 4, pp. 200-202.

OTHER PUBLICATIONS

Hamrick, Mark W. et al., The skeletal muscle secretome: an emerging player in muscle—bone crosstalk, BoneKEy Reports, 2012, vol. 1, Article No. 60, pp. 1-5.

Compton, J., Nucleic Acid Sequence-Based Amplification, Nature, 1991, vol. 350, No#, pp. 91-92. (Abstract Only).

International Search Report, PCT/US2014/020206, dated May 23, 2014, pp. 1-9.

Kariko, Katalin, et al., Impacts of Nucleoside Modification on RNA-mediated activation of toll-like receptors, 2008, Nucleic Acides in Innate Immunity, No Vol., pp. 171-188.

Cystic Fibrosis Transmembrane Conductance Regulator; cystic fibrosis transmembrane conductance regulator [*Homo sapiens*]; NCBI, 2010, No Vol., pp. 1-5.

Miotti, S. et al., Characterization of Human Ovarian Carcinoma-Associated Antigens Defined by Novel Monoclonal Antibodies with Tumor-Restricted Specificity, Intl. J. Cancer, 1987, vol. 39, No #, pp. 297-303

Robak, Tadeusz et al., Current and Emerging Treatments for Chrinic Lymphocytic Leukaemia, Drugs, 2009, vol. 69, No. 17, pp. 2415-2449.

Hutas, Ocrelizumab, a humanized monoclonal antibody against CD20 for inflammatory disorders and B-cell malignancies, Curr Opin Investig Drugs, 2008, vol. 11, No #, pp. 1206-1216. (Abstract Only).

Verma, Sandeep, et.al., Functional Tuning of Nucleic Acids by Chemical Modifications: Tailored Oligonucleotides as Drugs, Devices, and Diagnodtics, The Japan Chemical Journal Forum and Wiley Periodicals, Inc., 2003, Chem Rec 3, pp. 51-60.

Argininosuccinate synthetase; argininosuccinate synthetase, isoform CRA_b {Homo sapiens} NCBI, Dec. 18, 2006, No Vol., pp. 1-3.

Bovine Model of Citrullinemia, PNAS, 1999, vol. 96, No #, pp. 3081-3086

Strausberg et al., National Cancer Institute, Cancer Genome Anatomy Project, Tumor Gene Index, gene accession No. BE136127, 1997 pp. ??.

Lysosomal Acid Lipase (lysosomal acid lipase/ cholesteryl ester hydrolase isoform 1 precursor [*Homo sapiens*]; NCBI, 2010, No Vol., pp. 1-3.

Du et al., Lysosomal Acid Lipase Deficiency: Correction of Lipid Storage by Adenovirus-Mediated Gene Transfer in Mice; Human Gene Therapy; vol. 13, No #, pp. 1361-1372.

Gu, Minghao et al., Combinatorial synthesis with high throughput discovery of protein-resistant membrane surfaces, BioMaterials, 2013, vol. 34, No#., pp. 6133-6138.

Glucosylceramidase, glucosylceramidase isoform 1precursor [Homo sapiens]; NCBI, 2010, No Vol., pp. 1-4.

Robbins et al., Retroviral Vectors for Use in Human Gene Therapy for Cancer, Gaucher Disease, and Arthritis; Annals of the New York Academy of Sciences, 2006, vol. 716, No. 1, pp. 72-89.

Bertrand, Edouard et al., The snoRNPs and Related Machines: Ancient Devices That Mediate Maturation of rRNA and Other RNAs, 2004, Chapter 13, pp. 223-257.

Zhao, Xiansi et al., Regulation of Nuclear Receptor Activity by a Pseudouridine Synthase through Posttranscriptional Modification of Steroid Receptor RNA Activator, Molecular Cell, 2004, vol. 15, No #, pp. 549-558.

Zhao, Xinliang, Detection and quantitation of RNA base modifications, RNA, 2004, vol. 10:, pp. 996-1002.

Bosma, Piter Jabik et al., Inherited disorders of bilirubin metabolism, Journal of Hepatology, 2003, vol. 38, No #, pp. 107-117.

Chowdhury, Jayanta R. et al., Bilirubin Mono- and Diglucuronide Formation by Human Liver In Vitro: Assay by High-Pressure Liquid Chromatography, Hepatology, 1981, vol. 1, No. 6, pp. 622-627.

Chowdhury, Jayanta R. et al., Molecular Basis for the Lack of Bilirubin-specific and 3-Methylcholanthrene-inducibleU DP-GlucuronosyltransferaseActivities in Gunn Rats, Thej Ournaofl B Iological Chemistry, 1991, vol. 266, No. 27, pp. 18294-18298.

Chowdhury, Namita et al., Isolation of Multiple Normal and Functionally Defective Forms of Uridine Diphosphate-Glucuronosyltransferase from Inbred Gunn Rats, J. Clin. Invest, 1987, vol. 79, No. #, pp. 327-334.

Crigler, John et al. Society Transactions, Society for Pediatric Research, 31st Annual Meeting, Atlantic City, Congenital Familial Nonhemolytic Jaundice with Kernicterus: A New Clinical Entity, 1951, 3rd session, no Vol. pp. 1-3.

Miyagi, Shogo J. et al., The Development of UDP-Glucuronosyltransferases 1A1 and 1A6 in the Pediatric Liver, Drug Metabolism and Disposition, 2011, vol. 39, No. 5, pp. 912-919. Gunn, Charles, Hereditary Acholuric Jaundice in the Rat, Can M.J.,

1944, vol. 50, No #, pp. 230-237. Brockton, NT et al, UGT1A1 polymorphisms and colorectal cancer

susceptibility, Cancer, Gut, 2002; vol. 50, pp. 747-748. Iyanagi, Takashi et al., Molecular Basis of Multiple UDP-Glucuronosyltransferase Isoenzyme Deficiencies in the Hyperbilirubinemic Rat (Gunn Rat), 1991, vol. 266, No. 35, pp. 24048-24052.

Kadakol, Ajit et al., Genetic Lesions of Bilirubin Uridinediphosphoglucuronate Glucuronosyltransferase (UGT1A1) Causing Crigler-Najjar and Gilbert Syndromes: Correlation of Genotype to Phenotype, Human Mutation, 2000, vol. 16, No #, pp. 297-306.

Miranda, Paula S. Montenegro et al., Towards Liver-Directed Gene Therapy for Crigler-Najjar Syndrome, Current Gene Therapy, 2009, vol. 9, pp. 72-82.

Pastore, Nunzia et al., Sustained Reduction of Hyperbilirubinemia in Gunn Rats After Adeno-Associated Virus-Mediated Gene Transfer of Bilirubin UDP-Glucuronosyltransferase Isozyme 1A1 to Skeletal Muscle, Human Gene Therapy, 2012, vol. 23, No #, pp. 1082-1089.

Schmitt, Françoise et al., Lentiviral Vectors That Express UGT1A1 in Liver and Contain miR-142 Target Sequences Normalize Hyperbilirubinemia in Gunn Rats, Gastroenterology, vol. 139, No #,pp. 999-1007, 2010.

Strassburg, Christian P. et al., Hyperbilirubinemia syndromes (Gilbert-Meulengracht, Crigler-Najjar, Dubin-Johnson, and Rotor syndrome), Best Practice & Research Clinical Gastroenterology, 2010, vol. 24, No. #, pp. 555-571.

Sugatani, Junko et al., Transcriptional Regulation of Human UGT1A1 Gene Expression: Activated Glucocorticoid Receptor Enhances constitutive Androstane Receptor/ Pregnane X Receptor-Mediated UDP-Glucuronosyltransferase 1A1 Regulation with Glucocorticoid Receptor-Interacting Protein 1, Molecular Pharmacology, 2013, vol. 67, No. 3, pp. 845-855.

Batshaw, Mark L. et al., Treatment of Inborn Errors of Urea Synthesis, The New England Journal of Medicine, 1982, vol. 306, No. 23, pp. 1387-1392.

Batshaw, Mark L. Et al., Risk of Serious Illness in Heterozygotes for Ornithine Transcarbamylase Deficiency, J. Pediatr, 1986, vol. 108, No. 2, pp. 236-241.

Braissant, Olivier et al., Current concepts in the pathogenesis of urea cycle disorders, Molecular Genetics and Metabolism, 2010, vol. 100, pp. S3-S12.

Hodges, Peter E. et al., The spf h mouse: A missense mutation in the ornithine transcarbamylase gene also causes aberrant mRNA splicing, Genetics, Proc. Nati. Acad. Sci. USA, 1989,vol. 86, pp. 4142-4146.

Marini, Juan C et al., Phenylbutyrate improves nitrogen disposal via an alternative pathway without eliciting an increase in protein breakdown and catabolism in control and ornithine transcarbamylase-deficient patients, Am J Clin Nutr , 2011, vol. 93, No. #, pp. 1248-1254.

Rosenberg, Leon E., et al., Biogenesis of Ornithine Transcarbamylase in sprsh Mutant Mice: Two Cytoplasmic Precursors, One Mitochondrial Enzyme, Science,1983, vol. 222, No Vol. #, pp. 426-428.

Summar, MD, Marshall et al., Current strategies for the management of neonatal urea cycle disorders, The Journal of Pediatrics, 2001, vol. 138, No. 1, pp. s30-s39.

Walker, V., Ammonia toxicity and its prevention in inherited defects of the urea cycle, Diabetes, Obesity and Metabolism, 2009, vol. 11, No #, pp. 823-835.

OTHER PUBLICATIONS

Whitington, P. F. et al., Liver transplantation for the treatment of urea cycle disorders, J. Inher. Metab. Dis., 1998, vol. 21 (Suppl 1) pp. 112-118.

Wilcken, Bridget et al., Problems in the management of urea cycle disorders, Molecular Genetics and Metabolism, 2004, vol. 81, No #, S86-S91

Cosman, David et al., ULBPs, Novel MHC Class I-Related Molecules, Bind to CMV Glycoprotein UL16 and Stimulate NK Cytotoxicity through the NKG2D Receptor, Immunity,2001, vol. 14, No Vol. pp. 123-133.

Croft, Michael et al., TNF superfamily in inflammatory disease: translating basic insights, Trends Immunol, 2012; vol. 33, No. 3, pp. 144-152

Friese, Manuel A. et al., MICA/NKG2D-Mediated Immunogene Therapy of Experimental Gliomas, Cancer Res, 2003, vol. 63, pp. 8996-9006.

Gomes, Anita Q. et al., Non-classical major histocompatibility complex proteins as determinants of tumour immunosurveillance, 2007, EMBO reports, vol. 8, No. 11, pp. 1024-1030.

Guo, Z Sheng et al., Life after death: targeting high mobility group box 1 in emergent cancer therapies, Am J Cancer Res, 2013;vol. 3, No. 1 pp. 1-20.

Kane, Lawrence P. et al., TIM Proteins and Immunity, J Immunol., 2010; vol. 184, No. (6): 2743-2749.

Lanca, Telma et al., The MHC class Ib protein ULBP1 is a nonredundant determinant of leukemia/lymphoma susceptibility to gd T-cell cytotoxicity, Blood, 2010, vol. 115, No #, pp. 2407-2411. Lee, Sylvia et al., Cytokines in Cancer Immunotherapy , Cancers, 2011, vol. 3, No. #, pp. 3856-3893.

Lee, Judong et al., TIM Polymorphisms—Genetics and Function, Genes Immun. 2011, vol. 12, No. 8, pp. 595-604.

Raghavan, Malini et al., Calreticulin in the immune system: ins and outs, Cell Press, Trends in Immunology, 2013, vol. 34, No. 1, pp. 13, 21

Shin, Jae Hun et al., Positive conversion of negative signaling of CTLA4 potentiates anti-tumor efficacy of adoptive T cell therapy in murine tumor models, Blood, 2012, No Vol., pp. 1-29.

Sutherland, Claire L. et al., ULBPs, human ligands of the NKG2D receptor, stimulate tumor immunity with enhancement by IL-15, 2006, vol. 108, No #, pp. 1313-1319.

Wang, Haichao et al., HMG-1 as a Late Mediator of Endotoxin Lethality in Mice, Science, 1999, vol. 285, No. 284, pp. 248-251. Bikard, David et al., Programmable repression and activation of bacterial gene expression using an engineered CRISPR-Cas system, Nucleic Acids Research Advance, 2013, No Vol. 190, pp. 1-9. Le Cong et al., Multiplex Genome Engineering Using CRISPR/Cas

Systems, Science, 2013, vol. 339, No. 819, pp. 819-823. Ornithine Carbamoyltransferase; ornithine carbamoyltransferase, mitochondrial precursor [*Homo sapiens*]; NCBI, 2010, No Vol., pp. 1-3.

Kiwaki et al., Correction of Ornithine Transcarbamylase Deficiency in Adult spfash Mice and in OTC-Deficient Human Hepatocytes with Recombinany Adenoviruses Bearing the CAG Promoter; Human Gene Therapy, 1996, vol. 7, No #, pp. 821-830.

Hwang, Woong Y et al., Efficient genome editing in zebrafish using a CRISPR-Cas system, Nature Biotechnology, 2013, No Vol. pp. 1-3.

International Search Report, PCT/US2013/75177, dated May 5, 2014, pp. 1-20.

Robbins, Majorie et al., 2'-O-methyl-modified RNAs Act as TLR7 Antagonists, Molecular Therapy, 2007, vol. 15, No. 9, pp. 1663-1669.

Kandimalla, Ekambar R. et al.Design, synthesis and biological evaluation of novel antagonist compounds of Toll-like receptors 7, 8 and 9, Nucleic Acids Research, 2013, vol. 41, No. 6, pp. 3947-3961.

Hochreiter-Hufford, Amelia et al., And Digestion Clearing the Dead: Apoptotic Cell Sensing, Recognition, Engulfment, Cold Spring Harb Perspect Biol, 2013, No Vol #, pp. 1-20.

Kim, Sunjung et al, Transcriptional Suppression of Interleukin-12 Gene Expression following Phagocytosis of Apoptotic Cells, Immunity, 2004, vol. 21, No #, pp. 643-653.

Broz, Petr et al., Newly described pattern recognition receptors team up against intracellular pathogens, Nature Reviews, Immunology, 2013, vol. 13, No. #, pp. 551-565.

Bonham, Kevin S. et al., A Promiscuous Lipid-Binding Protein Diversifies the Subcellular Sites of Toll-like Receptor Signal Transduction, Cell, 2014, vol. 156, No #, pp. 705-716.

Ravichandran, Kodi S., Find-me and eat-me signals in apoptotic cell clearance: progress and conundrums, JEM, 2010, vol. 207, pp. 1807-1817.

Stuart, Lynda M. et al., Cell Maturation upon Endotoxin-Driven Myeloid Dendritic Inhibitory Effects of Apoptotic Cell Ingestion, The Journal of Immunology, 2002, vol. 168, No #, pp. 1627-1635. Wallet, Mark A et al., Immunoregulation of Dendritic Cells, Clinical Medicine & Research, 2005, Vo. 3, No. 3, pp. 166-175.

Williams, Charlotte A. et al, Apoptotic cells induce dendritic cell-mediated suppression via interferon-c-induced IDO, Immunology, 2007, vol. 124, No #, pp. 89-101.

Keegan, Liam P. et al., The Many Roles of an RNA Editor, Nature Reviews, Genetics, 2001, vol. 2, No #, pp. 869-878.

Felden, Brice et al., Presence and location of modified nucleotides in *Escherichia coli*t mRNA: structural mimicry with tRNA acceptor branches, The EMBO Journal, 1998, vol. 17 No. 11 pp. 3188-3196. Doffek, Kara et al., Phosphatidyserine Inhibits NFkB and p38 MAPK Activation in Human Monocyte Derived Dendritic Cells, Molecular Immunology, 2011, vol. 48, No. #, pp. 1771-1777.

Oberg (Aquaporins, Production Optimization and Characterization; Thesis for the Degree of Doctor of Philosophy in Natural Science; University of Gothenburg, Department of Chemistry—Biochemistry; pp. 1-69, published May 27, 2011. No Vol.

by hAQP5 (*Homo sapiens* aquaporin 5 (AQP5) mRNA; NCBI, pp. 1-5, published Dec. 27, 2010, No. Vol.

Iduronate 2-Sulfatase: iduronate 2-sulfatase isofirm a preproprotien [*Homo sapiens*], NCBI, 2010, No Vol., pp. 1-4.

European Supplementary Search Report, EP11815407, Jun. 13, 2014, pp. 1-13.

Bermudez et al., Treatment with Recombinant Granulocyte Colonystimulating Factor (Filgrastin) Stimulates Neutrophils and Tissue /macrophages and Induces an Effective non-specific Response Against *Mycobacterium avium* in Mice, Imnnunology, 1998, vol. 94, No. 3, pp. 297-303.

Sheridan, W. et al., Effects of Peripheral-Blood Progenitor Cells Mobilised by Filgrastim (G-CSF) on Platelet Recovery After High-Dose Chemotherapy, The Lancet, 1992, vol. 339, pp. 640-644.

Alpha Galactosidase A; alpha-galactosidase A precursor [*Homo sapiens*] NCBI, 2010, pp. 1-4.

Ziegler et al., AAV2 Vector Harboring a Liver-Restricted Promoter Facilates Sustained Expression of Therapeutic Levels of a-Galactosidase A and the Induction of Immune Tolerance in Fabry Mice, Molecular Therapy, 2004, vol. 9, No. 2, pp. 231-240.

International Search Report from International Application No. PCT/US2012/068714, dated Aug. 6, 2013.

Iduronate 2-Sulfatase; iduronate 2-sulfatase isofrom a preproprotein [*Homo sapiens*]; NCBI, 2010, pp. 1-4.

Braun et al., Preclinical Studies of Lymphocyte Gene Therapy for Mild Hunter Syndrome (Mucopolysaccharidosis Type II); Human Gene Therapy, 1996, Vol., No #, pp. 283-290.

Desmond Padhi et al., Single-Dose, Placebo-Controlled, Randomized Study of AMG 785, a Sclerostin Monoclonal Antibody, Journal of Bone and Mineral Research, vol. 26, No. 1, 2011, pp. 19-26.

Yu, Alice et al, Anti-GD2 Antibody with GM-CSF, Interleukin-2, and Isotretinoin for Neuroblastoma, The New England Journal of Medicine, 2010, vol. 363; No. 14, pp. 1324-1334.

Carboxypeptidas N, Carboxypeptidas N caralytic Chanin precursor [Homo sapiens] NCBI, 2010, pp. 1-4.

US 20020198163, 12/2002, Felgner et al. (withdrawn).

Abuchowski, A. et al., Immunosuppressive properties and circulating life of Achromobacter glutaminase asparaginase covalently attached to polyethylene glycol in man. Cancer Treat Rep. Nov. Dec. 1981;65(11-12):1077-81.

OTHER PUBLICATIONS

Abuchowski, A. et al., Reduction of plasma urate levels in the cockerel with polyethylene glycol-uricase. J Pharmacol Exp Ther. Nov. 1981;219(2):352-4.

Aduri, R., et al., AMBER force field parameters for the naturally occurring modified nucleosides in RNA. J Chem Theory Comput. 2007; 3: 1464-1475.

Agaisse, H. et al., STAB-SD: a Shine-Dalgarno sequence in the 5' untranslated region is a determinant of mRNA stability. Mol Microbiol. May 1996;20(3):633-43.

Aissani, B. et al., CpG islands, genes and isochores in the genomes of vertebrates. Gene. Oct. 15, 1991;106(2):185-95.

Akashi, H., Gene expression and molecular evolution. Curr Opin Genet Dev. Dec. 2001;11(6):660-666.

Aksenova, N.N. et al., Influence of ribonucleic acids from the liver on implantation and growth of transplantable tumours. Nature. Nov. 3, 1962;196:443-4.

Alberts, et al., Molecular Biology of the Cell, 3rd ed. Garland Publishing, Inc. New York, NY, 1994, pp. 368-369.

Aleku, M., et al., Atu027, a liposomal small interfering RNA formulation targeting protein kinase N3, inhibits cancer progression. Cancer Res. 2008; 68: 9788-9798.

Anderson, B.R., et al., Nucleoside modifications in RNA limit activation of 2'-5'-oligoadenylate synthetase and increase resistance to cleavage by Rnase L. Nucleic Acids Res. 2011; pp. 1-10.

Anderson, D.M. et al., Stability of mRNA/cationic lipid lipoplexes in human and rat cerebrospinal fluid: methods and evidence for nonviral mRNA gene delivery to the central nervous system. Hum Gene Ther. Feb. 10, 2003;14(3):191-202.

Andrews-Pfannkoch, C. et al., Hydroxyapatite-mediated separation of double-stranded DNA, single-stranded DNA, and RNA genomes from natural viral assemblages. pl Environ Microbiol. Aug. 2010;76(15):5039-45. Epub Jun. 11, 2010.

Andries, O., et al., Comparison of the gene transfer efficiency of mRNA/GL67 and pDNA/GL67 complexes in respiratory cells. Mol Pharmaceutics. 2012; 9: 2136-2145.

Anichini, A. et al., Cytotixic T cells directed to tumor antigens not expressed on normal melanocytes dominate HLA-A2.1-restricted immune repertoire to melanoma. J Immunol. Jan. 1, 1996;156(1):208-17.

Aota, S. et al., Diversity in G+C content at the third position of codons in vertebrate genes and its cause. Nucleic Acids Res. Aug. 26, 1986;14(16):6345-55.

Apostolopoulos, V. et al., Cellular mucins: targets for immunotherapy. Crit Rev Immunol. 1994;14(3-4):293-309.

Archer, S.J., Induction of a T-cell specific antigen on bone marrow lymphocytes with thymus RNA. Immunology Jan. 1978;34(1):123-

Ashley, D.M. et al., Bone marrow-generated dendritic cells pulsed with tumor extracts or tumor RNA induce antitumor immunity against central nevrous system tumors. J Exp Med. Oct. 6, 1997;186(7):1177-82.

Ast, G., How did alternative splicing evolve? Nat Rev Genet. Oct. 2004;5(10):773-82.

Aurup, H. et al., Translation of 2'-modified mRNA in vitro and in vivo. Nucleic Acids Res. Nov. 25, 1994;22(23):4963-8.

Austyn, J.M. et al., New insights into the mobilization and phagocytic activity of dendritic cells. J Exp MNed. Apr. 1, 1996;183(4):1287-92.

Babich, F.R. et al., Cross-species transfer of learning: effect of ribonuclec acid from hamsters on rat behavior. Proc Natl Acad Sci U S A. Nov. 1965;54(5):1299-302.

Bachellerie, J.P. et al., Antisense snoRNAs: a family of nucleolar RNAs with long complementarities to rRNA. Trends Biochem Sci. Jul. 1995;20(7):261-4.

Bag, J., Recovery of normal protein synthesis in heat-shocked chicken myotubes by liposome-mediated transfer of mRNAs. Can. J. Biochem. Cell Biol. 1985: 63(3): 231-235.

Bagnall, et al., Rat strain difference on performance in the Morris water maze. Animal Technology. 1999. 50(2):69-77.

Baker, D.L., et al., RNA-guided RNA modification: functional organization of the archaeal H/ACA RNP. Genes Dev. May 15, 2005;19(10):1238-48. Epub May 3, 2005.

Bakker, J.M. et al, Therapeutic antibody gene transfer: an active approach to passive immunity. Mol Ther. Sep. 2004;10(3):411-6. Balakin, A.G. et al., The RNA world of the nucleolus: two major families of small RNAs defined by different box elements with related functions. Cell. Sep. 6, 1996:86(5):823-34.

Bandbon Balenga, N.A. et al., Bicistronic expression plasmid encoding allergen and anti-IgE single chain variable fragment antibody as a novel DNA vaccine for allergy therapy and prevention. Med Hypotheses. 2006;67(1):71-4. Epub Mar. 2, 2006.

Banerjee, A.K., 5'-terminal cap structure in eucaryotic messenger ribonucleic acids. Microbiol Rev. Jun. 1980;44(2):175-205.

Barber, R., The chromatographic separation of ribonucleic acids. Biochim Biophys Acta. Feb. 21, 1966;114(2):422-4.

Bargmann, C.I. et al., The neu oncogene encodes an epidermal growth factor receptor-related protein. Nature. Jan. 16-22, 1986;319(6050):226-30.

Barlow, P.G., et al., The human cathelicidin LL-37 preferentially promotes apoptosis of infected airway epithelium. Am J Respir Cell Mol Biol. Dec. 2010; 43(6): 692-702.

Basarkar, A. et al., Nanoparticulate systems for polynucleotide deliver. Int J Nanomedicine. 2007; 2(3): 353-360.

Basha, G. et al., Influence of cationic lipid composition on gene silencing properties of lipid nanoparticle formulations of siRNA in antigen-presenting cells. Mol Ther. Dec. 2011; 19(12): 2186-2200. Bechler, K., Influence of capping and polydenylation on mRNA expression and on antisense RNA mediated inhibition of gene expression. Biochem Biophys Res Commun. Dec. 8, 1997;241(1):193-9.

Beljanski, et al., Iron stimulated RNA-dependent DNA polymerase activity from goldfish eggs. Cell Mol Biol. 1988;34(1):17-25.

Belliveau, N.M., et al., Microfluidic synthesis of highly potent limit-size lipid nanoparticles for in viro delivery of siRNA. Mol Ther Nucleic Acids. Aug. 2012; 1(8): e37.

Bernardi, G. et al., The vertebrate genome: isochores and evolution. Mol Biol Evol. Jan. 1993;10(1):186-204.

Bernhard, H. et al., Generation of immunostimulatory dendritic cells from human CD34+ hematopoietic progenitor cells of the bone marrow and peripheral blood. Cancer Res. Mar. 1, 1995;55(5):1099-104

Bernstein, E. et al., Role for a bidentate ribonuclease in the initiation step of RNA interference. Nature Jan. 18, 2001;409(6818):363-6. Bernstein, P. et al., Poly(A), poly(A) binding protein and the regulation of mRNA stability. Trends Biochem Sci. Sep. 1989;14(9):373-7.

Bertolini, M.C., et al., Fractionation of immune RNA isolated from the spleens of mice infected with Trypanosoma cruz. J Infect Dis. Jun. 1981;143(6):827-31.

Bertolini, In vitro effect of 18S immune RNA on macrophage resistance to Trypanosoma cruzi. Cell Mol Biol. 1986;32(2):167-71. Bertolini, The protective effect of the 4-5S immune RNA against Trypanosoma cruzi infection in mice. Trop Med Parasitol. Sep. 1985;36(3):131-4.

Bertrand, E. et al., Assembly and traffic of small nuclear RNPs. Prog Mol Subcell Biol. 2004;35:79-97.

Bettinger, T. et al., Peptide-mediated RNA delivery: a novel approach for enhanced transfection of primary and post-mitotic cells. Nucleic Acids Res. Sep. 15, 2001;29(18):3882-91.

Bevan, M.J. et al., Antigen presentation to cytototix T lymphocytes in vivo. J Exp Med. Sep. 1, 1995;182(3):639-41.

Bevilacqua, A. et al., Post-transcriptional regulation of gene expression by degradation of messenger RNAS. J Cell Physiol. Jun. 2003;195(3):356-72.

Bieler, K. et al., Plasmids for Therapy and Vaccination. Wiley-VCH GmbH. Weinheim, Feb. 2001.

Kariko, K. et al., Incorporation of pseudouridine into mRNA yields superior nonimmunogenic vector with increased translational capacity and biological stability. Mol Ther. Nov. 2008;16(11):1833-40. Epub Sep. 16, 2008.

OTHER PUBLICATIONS

Kariko, K. et al., Phosphate-enhanced transfection of cationic lipid-complexed mRNA and plasmid DNA. Biochim Biophys Acta. Mar. 2, 1998;1369(2):320-34.

Kariko, K., et al., In vivo protein expression from mRNA delivered into adult rat brain. J. of Neuroscience Methods. Jan. 2001; 105(1): 77-86.

Kariko, K. et al., mRNA is an endogenous ligant for Toll-like receptor 3. J Biol Chem. Mar. 26, 2004;279(13):12542-50. Epub Jan. 16, 2004.

Kariko, K. et al., Suppression of RNA recognition by Toll-like receptors: the impact of nucleoside modification and the evolutionary origin of RNA. Immunity. Aug. 2005;23(2):165-75.

Kariko, K., et al., Increased erythropoiesis in mice injected with submicrogram quantities of pseudouridine-containing mRNA encoding erythropoietin. Mol Ther. May 2012; 20(5): 948-853.

Karlin, S. et al., Applications and statistics for multiple high-scoring segments in molecular sequences. Proc Natl Acad Sci U S A. Jun. 15, 1993;90(12):5873-7.

Katre, N.V. et al., Chemical modification of recombinant interleukin 2 by polyethylene glycol increases its potency in the murine Meth A sarcoma model. Proc Natl Acad Sci U S A. Mar. 1987;84(6):1487-91

Katz, N., et al., Rapid onset of cutaneous anesthesia with EMLA cream after pretreatment with a new ultrasound-emitting device. Anesth Analg. 2004; 98: 371-376.

Kawai, T., et al., Antiviral signaling through pattern recognition receptors, J. Biochem. 2007; 141(2): 137-145.

Kawamura, T., et al., Linking the p53 tumor suppressor pathway to somatic cell reprogramming. Nature. Aug. 2009; 460(7259): 1140-1144.

Kazmierczak, K.M. et al., The phage N4 virion RNA polymerase catalytic domain is related to single-subunit RNA polymerases. EMBO J. Nov. 1, 2002;21(21):5815-23.

Keith, B., et al., HIF1a and HIF1a: sibling rivairy in hypoxic tumor growth and progression. Nat Rev Cancer. Jul. 2012; 12(1): 9-22. Keller, E.B. et al., Intron splicing: a conserved internal signal in introns of animal pre-mRNAs. Proc Natcl Acad Sci U S A. Dec. 1984;81(23):7417-20.

Keown, W.A., et al., [41] Methods for Introducing DNA into Mammalian Cells Methods in Enzymology, 1990, 185:527-37.

Keshishian, H., et al., Quantification of cardiovascular biomarkers in patient plasma by targeted mass spectrometry and stable isotope dilution. Mol Cell Proteomics. Oct. 2009; 8(10): 2339-2349.

Kesselheim, A.S., An empirical review of major legistation affecting drug developmement: Past experiences, effects, and unintended consequences. The Milbank Quarterly. 2011; 89(3): 450-502.

Khare, P.D. et al., Tumor growth suppression by a retroviral vector displaying scFv antibody to CEA and carrying the iNOS gene. Anticancer Res. Jul.-Aug. 2002;22(4):2443-6.

Khullar, N. et al., Comparative evaluation of the protective effect of immune spleen cells and immune RNA against Plasmodium berghei. Ann. Trop. Med. Parasitol., 1988, 82(6):519-26.

Kim, C.H. et al., Codon optimization for high-level expression of human erythropoietin (EPO) in mammalian cells. Gene. Oct. 15, 1997;199(1-2):293-301.

Kim, D., et al., Generation of human induced pluripotent stem cells by direct delivery of reprogramming proteins. Cell Stem Cell. Jun. 2009; 4(6): 472-476.

Kim, S.H., et al., Opsonized erythrocyte ghosts for liver-targeted delivery of antisense oligodeoxynucleotides. Biomaterials. Feb. 2009; 30(5): 959-967. Epub Nov. 22, 2008.

Kines, R.C. et al., The initial steps leading to papillomavirus infections occur on the basement membrane prior to cell surface binding. PNAS. Dec. 1, 2009; 106(48): 20458-20463.

Kinosita, K. Jr. et al., Formation and resealing of pores of controlled sizes in human erythrocyte membrane. Nature. Aug. 4, 1977;268(5619):438-41.

Kirby, K.S., A New Method for the Isolation of Ribonucleic Acids from Mammalian Tissues. J. Biochem., 1956, 64:405.

Kirshenbaum, et al., Designing polymers that mimic biomolecules. Curr Opin Struct Biol. 1999. 9:530-5.

Kirpotin, D.B., et al., Antobody targeting of long-circulating lipidic nanoparticles does not incease tumor localization but does increase internalization in animal models. Cancer Res. 2006; 66: 6732-6740. Kiss, T., Small nucleolar RNA-guided post-transcriptional modification of cellular RNAs. EMBO J. Jul. 16, 2001;20(14):3617-22. Kiss, T., Small nucleolar RNAs an abundant group of noncoding RNAs, with a divisor cellular functions.

Kiss, T., Small nucleolar RNAs: an abundant group of noncoding RNAs with diverse cellular functions. Cell. Apr. 19, 2002;109(2):145-8.

Kitaguchi, K. et al., Immune deficiency enhances expression of recombinant human antibody in mice after nonviral in vivo gene transfer. Int J Mol Med. Oct. 2005;16(4):683-8.

Klinman, D.M. et al., DNA vaccines: safery and efficacy issues. Springer Semin Immunopathol. 1997;19(2)2:245-56.

Koch, G. and Bishop, J.M. The effect of polycations on the interaction of viral RNA with mammalian cells: Studies on the infectivity of single- and double-stranded poliovirus RNA. Virology. May 1968; 35(1): 9-17.

Koch, G., et al., Quantitative Studies on the Infectivity of ribonucleic acid from partially purified and high purified poliovirus preparations, Virology. Mar. 1960; 10(3): 329-343.

Koch, G., et al., An agar cell-suspension plaque assay for isolated viral RNA. Biochem and Biophys Res. Comm. 1966; 24(3): 304-309

Kohler, G. et al., Continuous cultures of fused cells secreting antibody of predefined specificity. Nature. Aug. 7, 1975;256(5617):495-7.

Koide, Y. et al., DNA vaccines. Jpn J Pharmacol. Jul. 2000;83(3):167-74.

Koido, S. et al., Induction of antitumor immunity by vaccination of dendritic cells transected with MUC1 RNA. J. Immunol. Nov. 15, 2000;165(10):5713-9.

Kolb, A.F. et al., A virus-neutralising antibody is not cytotoxic in vitro. Mol Immunol. Feb. 2006;43(6):677-89.

Komar, A.A. et al., Synonymous codon substitutions affect ribosome traffic and protein folding during in vitro translation. FEBS Lett. Dec. 3, 1999;462(3):387-91.

Kontermann, R.E. et al., Recombinant bispecific antibodies for cancer therapy. Acta Pharmacol Sin. Jan. 2005;26(1):1-9.

Korsten, K.H. et al., The strategy of infection as a criterion for phylogenetic relationships of non-coli phages morphologically similar to phage T7. J Gen Virol. Apr. 1979;43(1):57-73.

Koski, G.K. et al., Cutting edge innate immune system discriminates between RNA containing bacterial versus eukaryotic structural features that prime for high-level IL-12 secretion by dendritic cells. J Immunol. Apr. 1, 2004;172(7):3989-93.

Krieg, P.A. et al., Functional messenger RNAs are produced by SP6 in vitro transcription of cloned cDNAs. Nucleic Acids Res. Sep. 25, 1984;12(18):7057-70.

Krieg, P.A. et al., In vitro RNA synthesis with SP6 RNA polymerase. Methods Enzymol. 1987;155:397-415.

Kreiter, S., et al., Intranodal vaccination with naked antigen-encoding RNA elicits potent prophylactic and therapeutic antitumoral immunity. Cancer Res. 2010; 70: 9031-9040.

Kreiter, S., et al., Tumor vaccination using messenger RNA: prospect of a future therapy. Curr Opinion in Immun. Jun. 23, 2011(3): 399-406.

Kudla, G. et al., High guanine and cytosine content increases mRNA levels in mammalian cells. PLoS Biol. Jun. 2006;4(6):e180. Epub May 23, 2006.

Kufe, D.W. et al., Holland-Frei cancer medicine, 6th edition. Hamilton (ON): BC Decker; 2003; Table 12-1.

Kugler, A. et al., Regression of human metastatic renal cell carcinoma after vaccination with tumor cell-dendritic cell hybrides. Nat Med. Mar. 2000;6(3):332-6.

Kuhn, A.N., et al., mRNA as a versatile tool for exogenous protein expression. Current Gene Therapy. Oct. 2012; 12(5): 347-361.

Nakamura, K. et al., The proliferation of plasma cells from mouse bone marrow in vitro. III. Primary and secondary immune responses associated with thymic RNA. Immunol Commun. 1979;8(5-6):511-29.

OTHER PUBLICATIONS

Nakamura, K., The proliferation of plasma cells from mouse bone marrow in vitro. II-Stimulation of IgG-producing cells by a RNasesensitive thymocyte homogenate. Cell Immunol. Aug. 1976:25(2):163-77.

Nallagatla, S.R. et al., A brilliant disguise for self RNA: 5'-end and internal modifications of primary transcripts suppress elements of innate immunity. RNA Biol. Jul.-Sep. 2008;5(3):140-4. Epub Jul. 20, 2008.

Narayanan, A. et al., Role of the box C/D motif in localization of small nucleolar RNAs to coiled bodies and nucleoli. Mol Biol Cell. Jul. 1999;10(7):2131-47.

Naz, R.K. et al., Novel human prostate-specific cDNA: molecular cloning, expression, and immunobiology of the recombinant protein. Biochem Biophys Res Commun. Oct. 11, 2002;297(5):1075-84.

Needleman, S.B. et al., A general method applicable to the search for similarities in the amino acid sequence of two proteins. J Mol Biol. Mar. 1970;48(3):443-53.

Nestle, F.O. et al., Vaccination of melanoma patients with peptideor tumor lysate-pulsed dendritic cells. Nat Med. Mar. 1998;4(3):328-32.

Neumann, E. et al., Fundamentals of electroporative delivery of drugs and genes. Bioelectrochem Bioenerg. Feb. 1999;48(1):3-16. Newby, M.I. et al., Sculpting of the spliceosomal branch site recognition motif by a conserved pseudouridine. Nat Struct Biol. Dec. 2002;9(12):958-65.

Newman, A. et al., Mutations in yeast U5 snRNA alter the specificity of 5' splice-site cleavage. Cell. Apr. 5, 1991;65(1):115-23.

Newman, A.J. et al., U5 snRNA interacts with exon sequences at 5' and 3' splice sites. Cell. Feb. 21, 1992;68(4):743-54.

Newmark, J. et al., Preparation and properties of adducts of streptokinase and streptokinase-plasmin complex with poly ethylene glycol and pluronic polyol F38. J Appl Biochem. 1982; 4:185-9. Ni, J. et al., Small nucleolar RNAs direct site-specific synthesis of pseudouridine in ribosomal RNA. Cell. May 16, 1997;89(4):565-73.

Nicholson, A.W. et al., Accurate in vitro cleavage by RNase III of phosphorothioate-substituted RNA processing signals in bacteriophage T7 early mRNA. Nucleic Acids Res. Feb. 25, 1988;16(4):1577-91.

Nielsen, D.A. et al., Preparation of capped RNA transcripts using T7 RNA polymerase. Nucleic Acids Res. Jul. 25, 1986;14(14):5936. Nielsen, P.E., Peptide nucleic acids as therapeutic agents. Curr Opin Struct Biol. Jun. 1999;9(3):353-7.

Nikolin, V.P. et al., Resistance of Mice Exposed to Whole-Body Irradiation to Transplanted Hemopoietic Cells Modified with RNA Preparations. Bull. Exp. Biol. Med., 2000, 129:5571-4.

Niu, M.C. et al., Genetic Manipulation in Higher Organisms; III. Detection of Soya Protein in Seeds Derived from Soya mRNA-Treated Rice. Scientia Sinica. 1980, 23:119-23.

Niu, M.C. et al., Ribonucleic acid-induced changes in mammalian cells. Proc Natl Acad Sci U S A. Oct. 15, 1961;47:1689-700.

Matsuda, A. et al., Nucleosides. 120. Synthesis of 2'-Deoxy-?-isocytidine and 2'-Deoxy-1-methyl-?-uridine from ?-Uridine^1. J Org Chem. 1981; 46:3603-3609.

Matsuda, A. et al., Synthesis of 3-Methylpseudouridine and 2'-Deoxy-3-Methyl-pseudouridine. Carbohydr Res. Mar. 1, 1982; 100: 297-302.

Bhattacharya, B.K. et al., A practical synthesis of N1-Methyl-2'-deoxy-?-uridine (?-Thymidine) and its incorporation into G-rich triple helix forming oligonucleotides. Nucleosides & Nucleotides. 1995; 14(6): 1269-1287.

Desaulniers, J.P. et al., Synthesis of 15N-enriched pseudouridine derivatives. Org Lett. Oct. 30, 2003; 5(22): 4093-4096.

Jachertz, D. et al., Treatment of P815 mastocytoma in DBA/2 mice with RNA. J Immunogen. 1974; 1: 355-362.

McGary, E.C. et al., Post-transcriptional regulation of erythropoietin mRNA stability by erythropoietin mRNA-binding protein. J Biologic Chem. Mar. 28, 1997; 272(13): 8628-8634. Hornung, V. et al., 5'-triphosphate RNA is the ligand for RIG-I.

Hornung, V. et al., 5'-triphosphate RNA is the ligand for RIG-I Science. Nov. 10, 2006; 314(5801): 994-997.

Davis, D.R. Stabilization of RNA stacking by pseudouridine. Nucleic Acids Res. 1995; 23(24): 5020-5026.

Monobe, M. et al., Beta-pseudouridine, a beer component, reduces radiation-induced chromosome aberrations in human lymphocytes. Mutat Res. Jul. 8, 2003; 538(1-2): 93-99.

Hanessian, S. et al., A highly stereocontrolled and efficient synthesis of alpha- and beta-pseudouridines. Tetrahedron Letters. 2003; 44: 8321-8323.

Shi, Y. et al., Identification and characterization of pancreatic eukaryotic initiation factor 2 alpha-subunit kinase, PEK, involved in translational control. Mol Cell Biol. Dec. 1998; 18(12): 7499-7509. Nguyen, A. et al., Quantitative assessment of the use of modified nucleoside triphosphates in expression profiling: differential effects on signal intensities and impacts on expression ratios. BMC Biotechnol. Jul. 31, 2002; 2:14.

Carrington, J.C. et al., Cap-independent enhancement of translation by a plant potyvirus 5' nontranslated region. J Virol. Apr. 1990; 64(4): 1590-1597.

Gallie, D. R. The 5'-leader of tobacco mosaic virus promotes translation through enhanced recruitment of elF4F. Nuc Acids Res. 2002; 30(15): 3401-3411.

Decatur, W. A. et al., RNA-guided nucleotide modification of ribosomal and other RNAs. J Biologic Chem. Jan. 10, 2003; 278(2): 695-698.

Badis, G. et al., A snoRNA that guides the two most conserved pseudouridine modifications within rRNA confers a growth advantage in yeast. RNA. Jul. 2003; 9(7): 771-779.

Nitin, N. et al., Peptide-linked molecular beacons for efficient delivery and rapid mRNA detection in living cells. Nuc Acids Res. 2004; 32(6): e58.

Cho, E.J. et al., mRNA capping enzyme is recruited to the transcription complex by phosphorylation of the RNA polymerase II carboxy-terminal domain. Genes Dev. Dec. 15, 1997; 11(24): 3319-3326.

Santi, D.V. Mechanistic studies of RNA modifying enzymes. RNA pseudouridine synthase and m5Cytosine methyl transferase. Nucleic Acids Symp Ser. 2000; 44: 147-148.

Strobel, I. et al., Human dendritic cells transfected with either RNA or DNA encoding influenza matrix protein M1 differ in their ability to stimulate cytotoxic T lymphocytes. Gene Ther. Dec. 2000; 7(23): 2028-2035.

Takahashi, T.T. et al., mRNA display: ligand discovery, interaction analysis and beyond. Trends in Biochem Sci. Mar. 2003; 28(3): 159-165.

Niu, M.C. et al., The Developmental Potentiality of the Liver-RNA-Treated Posterior Primitive Streak in the Chick Embryo. Biol. Bull, 1968, 135:200-7.

Niu, M.C. et al., The Entrance of Exogenous RNA into the Mouse Ascites Cell. Proc. Soc. Exp. Biol. Med., 1968, 128 (2):550-5.

Niu, M.C., RNA-Induced Biosynthesis of Specific Enzymes. PNAS, 1962, 48:1964-9.

Niu, M.C., Antagonistic Action of Exogenous Histone and RNA in Mouse Ascites Cells. Proc. Soc. Exp. Biol. Med., 1972, 140:256-62. Niu, M.C., Causal Analysis of Embryonic Differentiation; I. Responsiveness of Presumptive Ectoderm as a Regulating Factor in RNA Function. Exp. Cell Res., 1971, 64:57-64.

Niu, M.C., Causal Analysis of Embryonic Differentiation; II. Dual Function of Exogenous RNA in differentiation of Presumptive Ectoderm. Exp. Cell Res., 1971, 64:65-76.

Niu, M.C., Current Evidence Concerning Chemical Inducers. Evolution of Nervous Control from Primitive Organisms. 1959, 7-30. Niu, M.C., Functional Potentiality of Ribonucleic Acid. Acta. Unio. Int. Contra. Cancrum, third meeting Philadelphia, 1964, 20:995-6. Niu, M.C., Genetic manipulation in higher organisms; I. Goldfish ova as materials of operation, mRNA mediated alteration of the liver specific isozymes. Scientia Sinica, 1977, 20(6):803-8.

Ma, B. et al., HPV pseudovirions as DNA delivery vehicles. Ther Deliv. Apr. 2011; 2(4): 427-430.

OTHER PUBLICATIONS

Samarsky, D.A. et al., The snoRNA box C/D motif directs nucleolar targeting and also couples snoRNA synthesis and localization. EMBO J. Jul. 1, 1998;17(13):3747-57.

Santini, S.M. et al., Type I interferon as a powerful adjuvant for monocyte-derived dendritic cell development and activity in vitro and in Hu-PBL-SCID mice. J Exp Med. May 15, 2000;191(10):1777-88.

Sanyal, S. et al., Effects of RNA on the developmental potentiality of the posterior primitive streak of the chick blastoderm. Proc Natl Acad Sci U S A. Apr. 1966;55(4):743-50.

Saponara, A.G. et al., The isolation from ribonucleic acid of substituted uridines containing alpha-aminobutyrate moieties derived from methionine. Biochem Biophys Acta. Apr. 27, 1974;349(1):61-77.

Satoh, M. et al., X-linked immunodeficient mice spotaneously produce lupus-related anti-RNA helicase A autoantibodies, but are resistant to pristane-induced lupus. Int Immunol. Sep. 2003;15(9):1117-24.

Satthaporn, S. et al., Dendritic cells (II): Role and therapeutic implications in cancer. J R Coll Surg Edinb. Jun. 2001;46(3):159-67.

Satz, M.L. et al., Mechanism of immune transfer by RNA extracts. Immune RNA induces the synthesis of idiotype-bearing antigen receptors in noncommitted cells. Mol Cell Biochem. Dec. 16, 1980;33(3):105-13.

Scheel, B. et al., Immunostimulating capacities of stabilized RNA molecules. Eur J Immunol. Feb. 2004;34(2):537-47.

Schirrmacher, V. et al., Intra-pinna anti-tumor vaccination with self-replicating infectious RNA or with DNA encoding a model tumor antigen and a cytokine. Gene Ther. Jul. 2000;7(13):1137-47 Schmidt, W.M. et al., CapSelect: a highly sensitive method for 5' CAP-dependent enrichment of full-length cDNA in PCR-mediated analysis of mRNAs. Nucleic Acids Res. Nov. 1, 1999;27(21):e31. Schmitt, W.E. et al., In vitro induction of a bladder cancer-specific T-cell response by mRNA-transfected dendritic cells. J Cancer Res Clin Oncol. 2001;127(3):203-6.

Scholte, B.J. et al., Animal models of cystic fibrosis. J Cyst Fibros. Aug. 2004;3 Suppl 2:183-90.

Schott, J.W., et al., Viral and non-viral approaches for transient delivery of mRNA and proteins. Current Gene Ther. 2011; 11(5): 382-398.

Schuler, G. et al., Murine epidermal Langerhans cells mature into potent immunostimulatory dendritic cells in vitro. J Exp Med. Mar. 1, 1985;161(3):526-46.

Schuler-Thurner, B. et al., Mage-3 and influenza-matrix peptide-specific cytotoxic T cells are inducible in terminal stage HLA-A2.1+ melanoma patients by mature monocyte-derived dendritic cells. J Immunol. Sep. 15, 2000;165(6):3492-6.

Segura, J., et al., Monitoring gene therapy by extranal imaging of mRNA: Pilot study on murine erythropoietin. Ther Drug Monit. Oct. 2007; 29(5): 612-8.

Semple, S.C., et al., Efficient encapsulation of antisense oligonucleotides in lipid vesicles using ionizable aminolipids: formation of novel small multilamellar vesicle structures. Biochim Biophys Acta. Feb. 9, 2001; 1510(1-2): 152-166.

Semple, S.C., et al., Rational design of cationic lipids for siRNA delivery. Nat Biotechnol. Feb. 2010; 28(2): 172-176.

Serrate, S. et al., Transfer of cellular immunity in vivo with immune RNA in an allogeneic murine model. Clin Immunol Immunopathol. Jan. 1982;22(1):75-82.

Sharp, J.S. et al., Effect of translational signals on mRNA decay in Bacillus subtilis. J Bacteriol. Sep. 2003;185(18)5372-9.

Sharp, P.M. et al., DNA sequence evolution: the sounds of silence. Philos Trans R Soc Lond B Biol Sci. Sep. 29, 1995;349(1329):241-7.

Shea, R.G. et al., Synthesis, hybridization properties and antiviral activity of lipid-oligodeoxynucleotide conjugates. Nucleic Acids Res. Jul. 11, 1990;18(13):3777-83.

Shi, Y., et al., A combined chemical and genetic approach for the generation of induced pluripotent stem cells. Cell Stem Cell. Jun. 2008; 2: 525-528.

Shingo, T. et al., Erythropoietin regulates the in vitro and in vivo production of neuronal progenitors by mammalian forebrain neural stem cells. J Neurosci. Dec. 15, 2001;21(24):9733-43.

Shuman, S. et al., Purification and characterization of a GTP-pyrophosphate exchange activity from vaccinia virions. Association of the GTP-pyrophosphate exchange activity with vaccinia mRNA guanylyltransferase. RNA (guanine-7-) methyltransferase complex (capping enzyme). J Biol Chem. Dec. 10, 1980;256(23):1588-98. Shuman, S., Capping enzyme in eukaryotic mRNA synthesis. Prog

Nucleic Acid Res Mol Biol. 1995;50:101-29. Shuman, S., Structure, mechanism, and evolutuion of the mRNA capping apparatus. Prog Nucleic Acid Res Mol Biol. 2001;66:1-40.

Siena, S. et al., Expansion of Immunostimulatory Dendritic Cells from Peripheral Blood of Patients with Cancer. Oncologist. 1997;2(1):65-69.

Simonaro, C.M. et al., Joint and bone disease in mucopolysac-charidoses VI and VII: identification of new therapeutic targets and biomarkers using animal models. Pediatr Res. May 2005;57(5 Pt 1):701-7. Epub Mar. 3, 2005.

Slapikoff, S. et al., Mechanism of ribonucleic acid polymerase action. Effect of nearest neighbors on competition between uridine triphosphate and uridine triphosphate analogs for incorporation into ribonucleic acid. Biochemistry. Dec. 1967; 6(12): 3654-3658.

Sleeman, J. et al., Dynamic interactions between splicing snRNPs, coiled bodies and nucleoli revealed using snRNP protein fusions to the green fluorescent protein. Exp Cell Res. Sep. 15, 1998;243(2):290-304.

Smith, C.M. et al., Sno storm in the nucleolus: new roles for myriad small RNPs. Cell. May 30, 1997;89(5):669-72.

Smith, J.P., et al., Drug retention and distribution after intratumoral chemotherapy with fluorouracil/epinephrine injectable gel in human pancreatic cancer xenografts. Cancer Chemother Pharmacol. 1999; 44: 267-274.

Smith, K.P. et al., Interactions of U2 gene loci and their nuclear transcripts with Cajal (coiled) bodies: evidence for PreU2 within Cajal bodies. Mol Biol Cell. Sep. 2000;11(9):2987-98.

Smith, W.S. et al., RNA modified uridines: VI: Conformations of 3-[3-(S)-Amino-3-Carboxypropyl]Uridine (acp3U) from tRNA and 1-Methyl-3-[3-(S)-Amino-3-Carboxypropyl]Pseudouridine

(m1acp3?) from rRNA. Nucleosides and Nucleotides. 1992; 11(10):1683-94.

Smits, E., et al., RNA-based gene transfer for adult stem cells and T cells. Leukemia. 2004; 18: 1898-1902.

Smull, C.E., and Ludwig, E.H. Enhancement of the plaque-forming capacity of poliovirus ribonucleic acid with basic proteins. Journal of Bacteriology. 1962; 84(5): 1035-1040.

Sohn, R.L., et al., In-vivo particle mediated delivery of mRNA to mammalian tissues: ballistic and biological effects. Wound Rep and Regen. Jul.-Aug. 2001; 287-296.

Soll, D. Enzymatic modification of transfer RNA. Science. Jul. 23, 1971; 173(3994): 293-299.

Sontheimer, E.J. et al., The U5 and U6 small nuclear RNAs as active site components of the spliceosome. Science. Dec. 24, 1993;262(5142):1989-96.

Sousa, R. et al., T7 RNA polymerase. Prog Nucleic Acid Res Mol Biol. 2003;73:1-41.

Sousa, R., Use of T7 RNA polymerase and its mutants for incorporation of nucleoside analogs into RNA. Methods Enzymol. 2000:317:65-74.

Spooner, R.A. et al., DNA vaccination for cancer treatment. Gene Ther. May 1995;2(3):173-80.

Sproat, B.S., Chemistry and applications of oligonucleotide analogues. J Biotechnol. Jul. 31, 1995;41(2-3):221-38.

Staley, J.P. et al., Mechanical devices of the spliceosome: motors, clocks, springs, and things. Cell. Feb. 6, 1998;92(3):315-26.

Stanek, D. et al., Detection of snRNP assembly intermediates in Cajal bodies by fluorescence resonance energy transfer. J Cell Biol. Sep. 27, 2004;166(7):1015-25.

Steege, D.A., Emerging features of mRNA decay in bacteria. RNA. Aug. 2000;6(8):1079-90.

OTHER PUBLICATIONS

Steinman, R.M. et al., Dendritic cells: antigen presentation, accessory function and clinical relevance. Adv Exp Med Biol. 1993;329:1-9.

Steinman, R.M., The dendritic cell system and its role in immunogenicity. Annu Rev Immunol. 1991;9:271-96.

Stepinski, J. et al., Synthesis and properties of mRNAs containing the novel "anti-reverse" cap analogs 7-methyl(3'-O-methyl)GpppG and 7-methyl (3'-deoxy)GpppG. RNA. Oct. 2001;7(10):1486-95.

Malone, R.W. et al., Cationic liposome-mediated RNA transfection. Proc Natl Acad Sci U S A. Aug. 1989;86(16):6077-81.

Niu, M.C., Glucose-6-Phosphate: Re-examination of the RNA-Induced Activity in Mouse Ascites Tumor Cells. Science. 1965, 148:513-6.

Niu, M.C., Mode of Action of the Exogenous Ribonucleic Acid in Cell Function. Natl Cancer Inst. Monogr. 1964, 13:167-77.

Niu, M.C., et al., Poly(A)-attached RNA as activator in embryonic differentiation. Proc. Soc Exp Biol Med. Oct. 1974;147(1):318-22. Niu, M.C., et al., Presence of liver-forming fraction in fish egg mRNAs detected by its ability to encode albumin syntehsis. Scientia Sinica. 1980, 23(4):510-6.

Niu, M.C., et al., Re-examination of the DNA-mediated transformation in goldfish. Scientia Sinica, 1983, 24(7):700-7.

Niu, M.C., The Development of Tubular heart in RNA-Treated Post-Nodal pieces of Chick Blastoderm. J Embryol. Exp. Morphol., 1973, 29:485-501.

Niu, M.C., The Effect of mRNA on Nuclear Activity in Developing Systems. 1980, 415-33.

Niu, M.C., The role of Exogenous Heart-RNA in Development of the Chick Embryo Cultivated In Vitro. J Embryol. Exp. Morphol., 1970, 64:57-64.

Niu, M.C., Thymus Ribonucleic Acid and Embryonic Differentiation. PNAS, 1958, 44:1264-1274.

Niu, M.C. et al., Transfer of information from mRNA to chromosomes by reverse transcription in early development of goldfish eggs. Cellular and Molecular Biology, 1989, 35(3):333-45.

Niu, M.C., VII. New Approaches to the Problem of Embryonic Induction. Cellular Mechanisms, Differentiation and Growth. 1956, 155-71.

Oberhauser, B. et al., Effective incorporation of 2'-O-methyloligoribonucleotides into liposomes and enhanced cell association through modification with thiocholesterol. Nucleic Acids Res. Feb. 11, 1992;20(3):533-8.

Occhiogrosso, G., et al., Prolonged convection-enhanced delivery into the rat brainstem. Neurosurgery. Feb. 2003; 52(2): 388-394.

Odens, M., Prolongation of the Life Span in Rats. Journal of the American Geriatrics Soc. Oct. 1973; 11(1):450-1.

O'Doherty, U. et al., Human blood contains two subsets of dendritic cells, one immunologically mature and the other immature. Immonology. Jul. 1994;82(3):487-93.

Ofengand, J. et al., The function of pseudouridylic acid in transfer ribonucleic acid: II. Inhibition of amino acyl transfer ribonucleic acid-ribosome complex formation by ribothymidylyl-pseudouridylyl-cytidylyl-guanosine 3'-phosphate. J Biol Chem. Nov. 25, 1969; 244(22): 6241-6253.

Ohashi, H. et al., Efficient protein selection based on ribosome display system with purified components. Biochem Biophys Res Commun. Jan. 5, 2007;352(1):270-6. Epub Nov. 13, 2006.

Ohmichi, T. et al., Efficient bacterial transcription of DNA nanocircle vectors with optimized single-stranded promoters. Ohmichi T, Maki A, Kool ET. Proc Natl Acad Sci U S A. Jan. 8, 2002;99(1):54-9. Epub Dec. 18, 2001.

Okumura, K. et al., Bax mRNA therapy using cationic liposomes for human malignant melanoma. J Gene Med. 2008; 10: 910-917.

Owen, M. et al., Stromal stem cells; marrow derived osteogenic precursors. CIBA Foundation Symposium, 1988, 136:42-60.

Ozawa, T. et al., Amplification and analysis of cDNA generated from a single cell by 5'-RACE: application to isolation of antibody heavy and light chain variable gene sequences from single B cells. Biotechniques. Apr. 2006;40(4):469-70.

Padilla, R. et al., A Y639F/H784A T7 RNA polymerase double mutant displays superior properties for synthesizing RNAs with non-canonical NTPs. Nucleic Acids Res. Dec. 15, 2002;30(24):e138.

Paglia, P. et al., Murine dendritic cells loaded in vitro with soluble protein prime cytotoxic T lymphocytes against tumor antigen in vivo. J Exp Med. Jan. 1, 1996;183(1):317-22.

Painter, H., et al., 494. Topical delivery of mRNA to the murine lung and nasal epithelium. Mol Ther. 2004; 9: S187.

Palu, G. et al., In pursuit of new developments for gene therapy of human diseases. J Biotechnol. Feb. 5, 1999:68(1):1-13.

Palucka, A.K. et al., Taming cancer by inducing immunity via dendritic cells. Immunol Rev. Dec. 2007;220:129-50.

Papapetrou, E., et al., Stoichiometric and temporal requirements of Oct4, Sox2, Klf4, and c-Myc expression for efficient human IPSC induction and differentiation. Natl. Acad. Sci USA. Aug. 2009; 106: 12759-12764.

Paradi, E. et al., Changes in the content of modified nucleotides in wheat rRNA during greening. Biologia Plantarum. Apr. 2003; 47(1):33-8.

Park, I., et al., Reprogramming of human somatic cells to pluripotency with defined factors. Nature. Jan. 2008; 451(10): 141-146.

Parker, R. et al., Recognition of the TACTAAC box during mRNA splicing in yeast involves base pairing to the U2-like snRNA. Cell. Apr. 24, 1987;49(2):229-39.

Pascolo, S. Vaccination with messenger RNA (mRNA). Handb Exp Pharmacol. 2008; 183:221-235.

Passini, M.A. et al., AAV vector-mediated correction of brain pathology in a mouse model of Niemann-Pick A disease. Mol Ther. May 2005;11(5):754-62.

Passos, G.A. et al., In vivo induction of immunological memory to human tumor extract with poly (A)-containing immune RNA. Cell Mol Biol. 1988;34(2):157-64.

Paul, S., et al., How to improve R&D productivity: the pharmaceutical industry's grand challenge. Nat Reviews Drug Discovery. Mar. 2010; 9: 203-214.

Pays, E., Characterization of double-stranded ribonucleic acid sequences present in the initial transcription products of rat liver chromatin. Biochem J. Aug. 1, 1977;165(2):235-45.

Pearson, W.R. et al., Improved tools for biological sequence comparison. Proc Natl Acad Sci U S A. Apr. 1988;85(8):2444-8.

Peculis, B. RNA processing: pocket guides to ribosomal RNA. Curr Biol. Aug. 1, 1997;7(8):R480-2.

Peng, Z.H. et al., Synthesis and application of a chain-terminating dinucleotide mRNA cap analog. Org Lett. Jan. 24, 2002;4(2):161-4. Peoples, G.E. et al., Breast and ovarian cancer-specific cytotoxic T lymphocytes recognize the same HER2/neu-derived peptide. Proc Natl Acad Sci U S A. Jan. 17, 1995;92(2):432-6.

Perche, F., et al., Enhancement of dedritic cells transfection in vivo and of vaccination against B16F10 melanoma with mannosylated histidylated lipopolyplexes loaded with tumor antigen messenger RNA. Nanomed: Nanotech, Bio, and Med. Aug. 2011; 7(4): 445-453

Pesole, G. et al., Structural and functional features of eukaryotic mRNA untranslated regions. Gene. Oct. 3, 2001;276(1-2):73-81. Pesole, G. et al., UTRdb and UTRsite: specialized databases of sequences and functional elements of 5' and 3' untranslated regions of eukaryotic mRNAs. Update 2002. Nucleic Acids Res. Jan. 1,

2002;30(1):335-40. Petit, I. et al. G-CSF induces stem cell mobilization by decresing bone marrow SDF-I and up-regulating CXCR4, Nature Immunology, Jul. 2002; 3(7): 687-694.

Phillips, J. et al., Antisense RNA Amplification: A Liner Amplification Method for Analyzing the mRNA Population from single Living Cells. Methods. Dec. 1996;10(3):283-8.

Phizicky, E.M. et al., [31] Biochemical genomics approach to map activities to genes. Methods Enzymol. 2002;350:546-59.

Pollard, C., et al., Type I IFN counteracts the induction of antigenspecific immune responses by lipid-based delivery of mRNA vaccines. Mol Ther. Jan. 2013: 21(1): 251-259.

Ponsaerts, P. et al., Cancer immunotherapy using RNA-located dendritic cells. Clin Exp Immunol. Dec. 2003;134(3):378-84.

OTHER PUBLICATIONS

Ponsaerts, P. et al., Messenger RNA electroporation is highly efficient in mouse embryonic stem cells: successful FLPe- and Cre-mediated recombination. Gene Ther. Nov. 2004;11(21):1606-10

Ponsaerts, P., et al., Highly efficient mRNA-based gene transfer in feeder-free cultured H9 human embryonic stem cells. Cloning and Stem Cells. 2004; 6(3): 211-216.

Agadjanyan, M., Prototype Alzheimer's Disease Vaccine Using the Immunodominany B Cell Type from β —Amloid and Promiscuous T Cell Epitope Pan HLA DR-Binding Peptide, J Immunol, 2005, vol. 174, no number, pp. 1580-1586.

Cribbs, David H., Adjuvant-dependent Modulation of Th1 and Th2 Responses to Immunization with β-amyloid, International Immunology, vol. 15, No. 4, pp. 505-514.

Davtyan, H. et al., Immunogenicity, Efficacy, Safety, and Mechanism of Action of Epitope Vaccine (Lu AF20513) for Alzheimer's Disease: Prelude to a Clinical Trial, The Journal of Neuroscience, Mar. 2013, vol. 33, No. 11, pp. 4923-4934.

Zwick, M. et al., Identification and Characterization of a Peptide That Specifically Binds the Human, Broadly Neutralizing Anti-Human Immunodeficiency Virus Type 1 Antibody b12, Journal of Virology, Jul. 2001, vol. 75, No. 14, pp. 6692-6699.

Zwick, M. et al., Molecular Features of the Broadly Neutralizing Immunoglobulin G1, b12 Required for Recognition of Human Immunodeficiency Virus Type 1 gp120, Journal of Virology, 2003, vol. 77, No. 10, pp. 5863-5876.

Wilkinson, R. et al., Structure of the Fab Fragment of F105, a Broadly Reactive Anti-Human Immunodeficiency Virus (HIV) Antibody that Recognizes the CD4 Binding Site of HIV type 1 gp120, Journal of Virology, 2005, vol. 79, No. 20, pp. 13060-13069. Julien, Jean-Philippe et al., Broadly Neutralizing Antibody PGT121 Allosterically Modulates CD4 Binding via Recognition of the HIV-1 gp120 V3 Base and Multiple Surrounding Glycans, PLOS Pathogens, 2013, vol. 9, Issue 5, pp. 1-15.

Laursen, N. et al., Broadly Neutralizing Antibodies Against Influenza Viruses, Antiviral Research, 2013, vol. 98, no number, pp. 476-483

Barouch, Dan et al., Therapeutic Efficacy of Potent Neutralizing HIV-1-specific monoclonal Antibodies in SHIV-infected Rehesus Monkeys, Nature, 2013, vol. 503, No. 7475, pp. 224-228.

Shingai, M. et al., Antibody-mediated Immunotherapy of Macaques Chronically Infected with SHIV Suppresses Viraemia, Nature, 2013, vol. 503, No. 7475, pp. 277-280.

Balaza, Alejandro et al., Vectored Immunoprophylaxis Protects Humanized Mice from Mucosal HIV Transmission, Nature Medicine, 2014, vol. 3, pp. 296-300.

Burton, Dennis et al., A Large Array of Human Monoclonal Antibodies to Type 1 Human Immunodefiency Virus From Combinatorial Libraries of Asymptomatic Seropositive Individuals, Proc. Natl Acad., USA, 1991, vol. 88, No Number, pp. 10134-10137. Burton, Dennis et al., Efficient Neutralization of Primary Isolates of HIV-1 by a Recombinant Human Monoclonal Antibody, Science, 1994, vol. 266, No Number, pp. 1024-1027.

Scheid, Johannes et al., Sequence and Structural Convergence of Broad and Potent HIV Antibodies That Mimic CD4 Binding, Science, 2011, vol. 333, No Number, 1633-1637.

Ledford, H., Supercharged Antibodies Fight HIV-Related Virus in Monkeys, Nature, 2013, No Volume, pp. 1-2.

Delehanty, James B., Peptides for Specific Intracellular Delivery and Targeting of Nanoparticles: Implications for Developing Nanoparticle-Mediated Drug Delivery, Future Science, Therapeutic Delivery, 2010, vol. 1, No. 3, pp. 411-433.

Dharap, S.S., et al., Tumor-specific Targeting of an Anticancer Drug Delivery System by LHRH Peptide, PNAS, 2005, vol. 102, No. 36, pp. 12962-12967.

Du, L. et al., Arginine-rich cell-penetrating peptide dramatically enhances AMO-mediated ATM Aberrant Splicing Correction and Enables Delivery to Brain and Cerebellum, Human Molecular Genetics, 2011, vol. 20, No. 16, pp. 3151-3160.

Ezzat, Kariem et al. PepFect 14, a Novel Cell-penetrating Peptide for Oligonucleotide Deliver in Solution and As Solid Formulation, Nucleic Acids Research, 2011, vol. 39, No. 12, pp. 5284-5298.

Fang, Shun-lung et al., A Novel Cell-Penetrating Peptide Derived from Human Eosinophil Cationic Protein, PLOS One, 2013, vol. 8, Issue 3, pp. 1-13.

Giblin, M. et al., Selective Targeting of *E. coli* Heat-stable Enterotoxin Analogs to Human Colon Cancer Cells, Anticancer Research, 2006,vol. 26, No number, pp. 3243-3252.

Kelly, Kimberley et al., Isolation of a Colon Tumor Specific Binding Peptide Using Phage Display Selection, Neoplasia, 2003, vol. 5, No. 5, pp. 437-444.

Knowles, Lynn et al., CLT1 Targets Angiogenic Endothelium through CLIC1 and Fibronectin, Angiogenesis, 2012, vol. 15, No. 1, pp. 115-129.

Laakkonen, Pirjo et al., Homing Peptides as Targeted Delivery Vehicles, Interactive Biology, 2010, vol. 2, No number, pp. 326-337.

Li, Zhi Jie, et al. Peptides as Targeting Probes Against Tumor Vasculature for Diagnosis and Drug Delivery, Journal of Translational Medicine, 2012, vol. 10, Supp 1, No. s1, pp. 1-9.

Lin, Jieru et al., Bacterial Heat-Stable Enterotoxins: Translation of Pathogenic Peptides into Novel Targeted Diagnostics and Therapeutics, Toxins, 2010, vol. 2, No number, pp. 2028-2054.

Lo, Albert et al., Hepatocellular Carcinoma Cell-Specific Peptide Ligand for Targeted Drug Delivery, Molecular Cancer Therapeutics, 2008, vol. 7, No. 3, pp. 579-589.

Lu, Ruei-Min et al., Targeted Drug Delivery Systems Mediated by a Novel Peptide in Breast Cancer Therapy and Imaging, PLOS One, 2013, vol. 8, Issue 6, pp. 1-13.

Pangburn, Todd et al., Peptide- and Aptamer-Functionalized Nanovectors for Targeted Delivery of Therapeutics, Journal of Biomedical Engineering, 2009, vol. 131, No number, pp. 1-20.

Phelan, Anne et al., Intercellular Delivery of Functional p53 by the Herpesvirus Protein VP22, Nature Biotechnology, 1998, vol. 16, pp. 440-443.

Laakkonen, Pirjo et al., Homing Peptides as Targeted Delivery Vehicles, Integrative Biology, 2010, vol. 2, no number, pp. 326-337. Regberg, Jakob et al., Applications of Cell-Penetrating Peptides for Tumor Targeting and Future Cancer Therapies, Pharmaceuticals, 2012, vol. 5, No number, pp. 991-1007.

Suchanek, Gerda et al., Amino Acid Sequence of Honeybee Prepromelittin Synthesized in Vitro, Proc. Natl. Acad. Sci. USA,1978, vol. 75, No. 2, pp. 701-704.

Torchilin, Vladimir et al., Multifunctional and Stimuli-Sensitive Pharmaceutical Nanocarriers, Eur J. Pharm Biopharm, 2009, vol. 71, No. 3, pp. 431-444.

Yang, Xiaoming, et al., Effect of CD44 Binding Peptide Conjugated to an Engineered Inert Matrix on Maintenance of Breast Cancer Stem Cells and Tumorsphere Formation, PLOS One, 2013, vol. 8, Issue 3, pp. 1-15.

Zou, Li-li et al., Cell-Penetrating Peptide-Mediated Therapeutic Molecule Delivery Into the Central Nervous System, Current Neuropharmacology, 2013, vol. 11, No. 2, pp. 197-208.

Baars, A. et al., A Phase II Study of Active Specific Immunotherapy and 5-FU/Leucovorin as Adjuvant Therapy for Stage III Colon Carcinoma, British Journal of Cancer, 2002, vol. 86, No. 8, pp. 1230-1234.

Badawi, Ahmed, et al., Immune Modulating Peptide for the Treatment and Suppression of Multiple Sclerosis, Clin Immunol, 2012, vol. 144, No. 2, pp. 127-138.

Bandala-Sanchez, Esther et al., T cell Regulation Mediated by Interaction of Soluble CD52 with the Inhibitory Receptor Siglec-10, Nature Immunology, 2013, vol. 14, No. 7, pp. 741-751.

Lu, Changming et al., miR-221 and miR-155 Regulate Human Dendritic Cell Development Apoptosis, and IL-12 Production Through Targeting of p27kip1, KPC1 and SOCS-1, Blood, 2011, vol. 117, No. 16, pp. 4293-4303.

Chang, C et al., Tolerization of Dendritic Cells by Ts cells: The Crucial Role of Inhibitory Receptors ILT3 and ILT4, Nature Immunology, 2002, vol. 3, No. 3, pp. 237-243.

OTHER PUBLICATIONS

Cheng, Guotan et al., T Cell Tolerance and the Multi-Functional Role of IL-2R Signalling in T Regulatory Cells, Immunol Rev., 2011, vol. 241, No. 1, pp. 63-76.

Cools, Nathalie, et al., Balancing Between Immunity and Tolerance: an Interplay Between Dendritic Cells, Regulatory T Cells, and Effector T Cells, Journal of Leukocyte Biology, 2007, vol. 82, pp. 1365-1374.

Cousens, Leslie et al., Tregitope Update: Mechanism of Action Parallels IVIg, Autoimmunity Reviews, 2012, No Volume, pp. 1-8. Cousens, L. et al., In Vitro and In Vitro Studies of IgC-derived Treg Epitopes (Tregitopes): A Promising New Tool for Tolerance Induction and Treatment of Autoimmunity, J. Clin. Immunol, 2013, vol. 33, Supp 1, pp. 43-49.

Cousens, Leslie et al., Application of IgC-Derived Natural Treg Epitopes (IgG Tregitopes) to Antigen-Specific Tolerance Induction in a Murine Model of Type 1 Diabetes, Journal of Diabetes, vol. 2013, Article ID 621693, pp. 1-17.

Danke, Nancy et al., Comparative Study of GAD65-specific CD4+ T cells in healthy and Type 1 Diabetic Subjects, Journal of AutoImmunity, 2005, vol. 25, No Number, 303-311.

DeGroot, Anne S. et al., Activation of Natural Regulatory T cells by IgG F-derived peptide "Tregitopes", 2008, vol. 112, No. 8, pp. 3303-3311.

DiCaro, Valentina, et al., In Vivo Delivery of Nucleic Acid-Formulated Microparticles as a Potential Tolerogenic Vaccine for Type 1 Diabetes, 2012, vol. 9, No. 4, pp. 348-356.

EMEA, Committee for Medicinal Products for Human Use, European Medicines Agency, 2008, No Vol. pp. 1-13.

European Search Report in Application No. 12831509.0, dated Apr. 8, 2015.

Kariko, K. et al., Incorporation of pseudouridine into mRNA yields superior nonimmunogenic vector with increased translational capacity and biological stability. Molecular Therapy, Nature Publishing Group, GB, vol. 16, No. 11, Nov. 1, 2008, pp. 1833-1840. Kariko, K. et al, Generating the optimal mRNA for therapy: HPLC purification eliminates immune activation and improves translation of nucleoside-modified, protien-encoding mRNA, Nucleic Acids Research, Oxford University Press, GB, vol. 39, No. 21, Sep. 2, 2011, pp. e142-1.

Kariko, K., et al., Impacts of nucleoside modification on RNA-mediated activation of toll-like receptors, Jan. 1, 2008, Nucleic Acids in Innate Immunity, CRC Press-Taylor & Francis Group, 6000 Broken Sound Parkway NW, STE 300, Boca Raton, FL 33487-2742 USA, pp. 171-188.

Conry, R.M. et al., Characterization of a messenger RNA polynucleotide vaccine vector. Cancer Res. Apr. 1, 1995;55(7):1397-400.

Conry, R.M. et al., Immune response to a carcionoembryonic antigen polynucleotide vaccine. Cancer Res. Mar. 1, 1994;54(5):1164-8.

Conry, R.M. et al., A carcinoembryonic antigen polynucleotide vaccine has in viro antitumor activity. Gene Ther. Jan. 1995;2(1):59-65

Copreni, E. et al., Lentivirus-mediated gene transfer to the respiratory epithelium: a promising approach to gene therepy of cystic fibrosis. Gene Ther. Oct. 2004;11 Suppl 1:S67-75.

Cortes, J.J. et al., Mutations in the conserved loop of human U5 snRNA generate use of novel cryptic 5' splice sites in vivo. EMBO J. Dec. 15, 1993;12(13):5181-9.

Coughlin, C.M. et al., Targeting adult and pediatric cancers via cell-based vaccines and the prospect of activated B lymphocytes as a novel modality. Cancer Biol Ther. Sep.-Oct. 2003;2(5):466-70. Cox, G.J. et al., Bovine herpesvirus 1: immune responses in mice

Cox, G.J. et al., Bovine herpesvirus 1: immune responses in mice and cattle injected with plasmid DNA. J Virol. Sep. 1993;67(9):5664-7.

Craig, J.M. et al., The distribution of CpG islands in mammalian chromosomes. Nat Genet. Jul. 1994;7(3):376-82.

Cramer, P. et al., Functional association between promoter structure and transcript alternative splicing. Proc Natl Acad Sci U S A. Oct. 14, 1997;94(21):11456-60.

Cree, B. et al., Tolerability and effects of rituxamab (anti CD20 antibody) in neuromyelitis optica (NMO) and rapidly worsening multiple sclerosis (MS). Neurology. 2004; 62(S5):A492.

Cuburu, N. et al., Intravaginal immunization with HPV vectors induces tissue-resident CD8+ T cell responses. J Clin Invest. Dec. 3, 2012; 122(12): 4606-4620.

Culver, K.W. et al., Gene Therapy. A Handbook for Physicians. Mary Ann Lieber, Inc. New York. 1994; 63-77.

Cunningham, S. et al., AAV2/8-mediated correction of OTC deficiency is robust in adult but not neonatal Spf^ ash Mice. Mol Ther. Aug. 2009; 17(8): 1340-1346.

Daguer, J.P. et al., Increasing the stability of sacB transcript improves levansucrase production in Bacilius subtilis. Lett Appl Microbiol. 2005;41(2):221-6.

Dai, M.S. et al., Introduction of human erythropoietin receptor complementary DNA by retrovirus-mediated gene transfer into murine embryonic stem cells enhances erythropoiesis in developing embryoid bodieds. Biol Blood Marrow Transplant. 2000;6(4):395-407

Davidson, E.H., An Analysis of Niu Menchang's Research on Transformation by RNA. Biotechnology in China, 1989, 92-102.

Davis, H.L. et al., DNA-based immunization induces continuous secretion of hepatitis B surface antigen and high levels of circulating antibody. Hum Mol Genet. Nov. 1993;2(11):1847-51.

De Carvalho, S. et al., Biologic properties of human leukemic and tumoral RNA. IV. Leukemia and neoplasms induced in mice with human leukemic RNA carried in tissue culture. J Lab Clin Med. May 1960;55:706-14.

De Carvalho, S. et al., Comparative effects of liver and tumour ribonucleic acids on the normal liver and the Novikoff hepatoma cells of the rat. Nature. Mar. 11, 1961;189:815-7.

De Carvalho, S. et al., Differences in information content of ribonucleic acids from malignant tissues and homologous organs as expressed by their biological activities. Exp Mol Pathol. Apr. 1962;1:96-103.

De Carvalho, S., Angiokines, angiogenesis and angiolymphoproliferative syndromes (ALPS). Angiology. Apr. 1983;34(4):231-43.

De Carvalho, S., Biologic properties of human leukemic and tumoral RNA, III. The effect of different media on the cytopathogenicity in tissue culture. J Lab Clin Med. May 1960:55:694-705.

De Carvalho, S., Cancer 1974: an analytical vademecum of oncologic relevance. Oncology. 1973;28(4):289-98.

De Carvalho, S., Effect of RNA from normal human bone marrow on leukaemic marrow in vivo. Nature. Mar. 16, 1963;197:1077-80. De Carvalho, S., Epigenetic transformation by RNA from human neoplastic cells. Oncology. 1973;27(1):3-29.

De Carvalho, S., In vitro angiogenic activity of RNA from leukemic lymphocytes. Angiology. Jul. 1978;29(7):497-505.

De Carvalho, S., Natural history of cogenital leukemia. An experiment of nature revealing unexplored features of fetal-material isoimmunity, longest recorded survival following use of leukemostatic maternal isoantibody. Oncology. 1973;27(1):52-63.

De Lucca, F.L., et al., Effect of the calcium phosphate-mediated RNA uptake on the transfer of cellular immunity of a synthetic peptide of HIV-1 to human lymphocytes by exogenous RNA. Mol Cell Biochem. Dec. 2001;228(1-2):9-14.

Delafontaine, P. et al., Regulation of vascular smooth muscle cell insulin-like growth factor I receptors by phosphorothioate oligonucleotides. Effects on cell growth and evidence that sense targeting at the ATG site increases receptor expression. J Biol Chem. Jun. 16, 1995;270(24):14383-8.

Deres, K. et al., In vivo priming of virus-specific cytotoxic T lymphocytes with synthetic lipopeptide vaccine. Nature. Nov. 30, 1989;342(6249):561-4.

Deshayes, S. et al., Cell-penetrating peptides: tools for intracellular delivery of therapeutics. Cell Mol Life Sci. Aug. 2005;62(16):1839-49

OTHER PUBLICATIONS

Desrosiers, R. et al., Identification of methylated nucleosides in messenger RNA from Novikoff hepatoma cells. Proc Natl Acad Sci U S A. Oct. 1974;71(10):3971-5.

Diebold, S.S. et al., Innate antiviral responses by means of TLR7-mediated recognition of single-stranded RNA. Science. Mar. 5, 2004;303(5663):1529-31. Epub Feb. 19, 2004.

Dimari, J.F. et al., Initiation of mRNA decay in Bacillus subtilis. Mol Bicrobiol. Mar. 1993;7(5):705-17.

Ding, Z., et al., State-of-the-art 2003 on PKU gene therapy. Mol Genet Metab. Jan. 2004;81(1):3-8.

Dingman, W. et al., Molecular theories of memory. Science. Apr. 3, 1964;144(3614):26-9.

Disbrow, G.L. et al., Codon optimization of the HPV-16 E5 gene enhances protein expression. Virology. Jun 20, 2003;311(1):105-14. Dong, Y. et al., Poly(d.l-lactide-co-glycolide)/montmorillonite nanoparticles for oral delivery of anticancer drugs. Biomaterials. Oct. 2005;26(30):6068-76.

Donnelly, J. et al., Technical and regulatory hurdles for DNA vaccines. Int J Parasitol. May 2003;33(5-6):457-67.

Dubes, G.R. and Klingler, E.A. Jr. Facilitation of infection of monkey cells with poliovirus "ribonucleic acid." Science. Jan. 1961; 133(3446): 99-100.

Dunham, S.P. et al., The application of nucleic acid vaccines in veterinary medicine. Res Vet Sci. Aug. 2002;73(1):9-16.

Dunn, J.J. et al., Different template specificities of phage T3 and T7 RNA polymerases. Nat New Biol. Mar. 17, 1971;230(11):94-6.

Duret, L. et al., Expression pattern and surprisingly, gene length shape codon usage in Caenorhabditis, *Drosophila*, and Arabidopsis. Proc Natl Acad Sci U S A. Apr. 13, 1999;96(8):4482-7.

Duret, L., Evolution of synonymous codon usage in metazonas. Curr Opin Genet Dev. Dec. 2002;12(6):640-9.

Earl, R.A., et al., A chemical synthesis of the nucleoside 1-Methylpseudouridine. A facile chemical synthesis of 1-methylpseudouridine has been accomplished by direct methylation of pseudouridine. J Heterocyclic Chem. Jun. 1977;14:699-700.

Easton, L.E. et al., Rapid, nondenaturing RNA purification using weak anion-exchange fast performance liquid chromatography. RNA. Mar. 2010;16(3):647-53. Epub Jan. 25, 2010.

Eberwine, J. et al., Analysis of gene expression in single live neurons. Proc Natl Acad Sci U S A. Apr. 1, 1992;89(7):3010-4. Edelstein, M. L. et al., Gene therapy clinical trials worldwide

1989-2004—an overview. J Gene Med. Jun. 2004;6(6):597-602. Edery, I. et al., An efficient strategy to isolate full-length cDNAs based on an mRNA cap retention procedure (CAPture). Mol Cell Biol. 1995; 15(6): 3363-3371.

Edmonds, M., Polyadenylate polymerases. Methods Enzymol. 1990:181:161-70.

Biocca, S., et al., Intracellular expression of anti-p21[^] ras single chain Fv fragments inhibits meiotic maturation of xenopus oocytes. Biochem Biophys Res Comm. Dec. 15, 1993; 197(2): 422-427.

Bird, A.P. et al., CpG-rich islands and the function of DNA methylation. Nature. May 15-21, 1986;321(6067):209-13.

Black, D.D. et al., Similarity of the transfer factors in Novikoff ascites tumor and other amino acid-incorporating systems. Cancer Res. May 1970;30(5):1281-6.

Bloch, G. et al., Sequence-dependence of the conformational changes induced by the 5-methyl cytosine in synthetic RNA oligomers. FEBS Lett. Jul. 27, 1987;219(2):464-8.

Boczkowski, D. et al., Dendritic cells pulsed with RNA are potent antigen-presenting cells in vitro and in vivo. J Exp Med. Aug. 1, 1996;184(2):465-72.

Boczkowski, D. et al., Induction of tumor immunity and cytotoxic T lymphocyte responses using dendritic cells transfected with messenger RNA amplified from tumor cells. Cancer Res. Feb. 15, 2000;60(4):1028-34.

Bonehill, A., et al., Single-step antigen loading and activation of dendritic cells by mRNA electroporation for the purpose of therapeutic vaccination in melanoma patients. Clin Cancer Res. May 2009; 15(10): 3366-3375.

Boon, T. et al., Genes coding for tumor rejection antigens: perspectives for specific immunotherapy. Important Adv Oncol. 1994:53-

Bose, S. et al., Role of nucleolin in human parainfluenza virus type 3 infection of human lung epithelial cells. J Virol. Aug. 2004;78(15):8146-58.

Bouxsein, N.F., et al., Structure and gene silencing activities of monovalent and pentavalent cationic lipid vectors complexed with siRNA†. Biochem. 2007; 46(16): 4785-4792.

Brandt, B. et al., Detection of the metastatic potential of bloodborne and immunomagnetically enriched epithelial cells by quantitative erbB-2 RT-PCR. Clin Exp Metastasis. Sep. 1996;14(4):399-408

Brieba, L.G., et al., Role of T7 RNA polymerase His784 in start site selection and initial transcription. Biochem. 2002; 41: 5144-5149. Brossart, P. et al., Her-2/neu-derived peptides are tumor-associated antigens expressed by human renal cell and colon carcinoma lines and are recognized by in vitro induced specific cytotoxic T lymphocytes. Cancer Res. Feb. 15, 1998;58(4):732-6.

Brossart, P. et al., Identification of HLA-A2-restricted T-cell epitopes derived from the MUC1 tumor antigen for broadly applicable vaccine therapies. Blood. Jun. 15, 1999;93(12):4309-17.

Brossart, P. et al., Induction of cytotoxic T-lymphocyte responses in vivo after vaccinations with peptide-pulsed dendritic cells. Blood. Nov. 1, 2000;96(9):3102-8.

Brossart, P. et al., Virus-mediated delivery of antigenic epitopes into dendritic cells as a means to induce CTL. J Immunol. Apr. 1, 1997;158(7):3270-6.

Buccoliero, R. et al., Elevation of lung surfactant phosphatidylcholine in mouse models of Sandhoff and of Niemann-Pick A disease. J Inherit Metab Dis. 2004;27(5):641-8.

Burke, B. et al., Microinjection of mRNA coding for an anti-Golgi antibody inhibits intracellular transport of a viral membrane protein. Cell. Apr. 1984;36(4):847-56.

Burks, E.A. et al, In vitro scanning saturation mutagenesis of an antibody binding pocket. Proc Natl Acad Sci U S A. Jan. 21, 1997;94(2):412-7.

Butler, E.T. et al., Bacteriophage SP6-specific RNA polymerase. I. Isolation and characterization of the enzyme. J Biol Chem. May 25, 1982;257(10):5772-8.

Cannon, G. et al., RNA based vaccines. DNA Cell Biol. Dec. 2002;21(12):953-61.

Capoccia, B.J., et al., G-CSF and AMD3100 mobilize monocytes into the blood that stimulate angiogenesis in vivo through a paracrine mechanism. Blood Oct. 1, 2006; 108(7): 2438-2445.

Caput, D. et al., Identification of a common nucleotide sequence in the 3'-untranslated region of mRNA molecules specifying inflammatory mediators. Proc Natl Acad Sci U S A. Mar. 1986;83(6):1670-4.

Caron, H. et al., The human transcriptome map: clustering of highly expressed genes in chromosomal domains. Science. Feb. 16, 2001;291(5507):1289-92.

Carralot, J.P. et al., Polarization of immunity induced by direct injection of naked sequence-stabilized mRNA vaccines. Cell Mol Life Sci. Sep. 2004;61(18):2418-24.

Carralot, J.P. et al., Production and characterization of amplified tumor-derived cRNA libraries to be used as vaccines against metastatic melanomas. Genet Vaccines Ther. Aug. 22, 2005;3:6.

Caudy, A.A. et al., Fragile X-related protein and VIG associate with the RNA interference machinery. Genes Dev. Oct. 1, 2002;16(19):2491-6.

Cavaille, J. et al., Identification of brain-specific and imprinted small nucleolar RNA genes exhibiting an unusual genomic organization. Proc Natl Acad Sci U S A. Dec. 19, 2000;97(26):14311-6. Cavaille, J. et al., Targeted ribose methylation of RNA in vivo directed by tailored antisense RNA guides. Nature. Nov. 24, 1996;383(6602):732-5.

OTHER PUBLICATIONS

Celluzzi, C.M. et al., Peptide-pulsed dendritic cells induce antigenspecific CTL-mediated protective tumor immunity. J Exp Med. Jan. 1, 1996;183(1):283-7.

Chan, E. et al., Live cell imaging distinguishes bona fide human iPS cells from partially reprogrammed cells. Nat Biotech. Nov. 2009: 27(11): 1033-1037.

Chappell, S.A. et al., Ribosomal tethering and clustering as mechanisms for translation initiation. Proc Natl Acad Sci U S A. Nov. 28, 2006;103(48):18077-82. Epub Nov. 16, 2006.

Charette, M. et al., Pseudouridine in RNA: what, where, how, and why. IUBMB Life. May 2000;49(5):341-51.

Chen, D., et al., Rapid discovery of potent siRNA-containing lipid nanoparticles enabled by controlled microfluidic formulation. J Am Chem Soc. 2012; 134: 6948-6951.

Chen, H., et al., TGF-beta 1 attenuates myocardial ischemiareperfusion injury via inhibition of upregulation of MMP-1. Am J Physiol Heart Circ Physiol. May 2003; 284(5): H1612-7.

Chen, Z. et al., Enhanced protection against a lethal influenza virus challenge by immunization with both hemagglutinin- and neuraminidase-expressing DNAs. Vaccine. Feb. 26, 1999;17(7-8):653-9.

Cheng, C., et al., Multifunctional triblock copolymers for intracellular messenger RNA delivery. Biomaterials. Oct. 2012; 33(28): 6868-6876.

Cheng, W.F. et al., Enhancement of Sindbis virus self-replicating RNA vaccine potency by linkage of herpes simplex virus type 1 VP22 protein to antigen. J Virol. Mar. 2001;75(5):2368-76.

Cheng, W.F. et al., Enhancement of Sindbis virus self-replicating RNA vaccine potency by linkage of *Mycobacterium tuberculosis* heat shock protein 70 gene to an antigen gene. J Immunol. May 15, 2001;166(10):6218-26.

Cho, J.H. et al., Enhanced cellular immunity to hepatitis C virus nonstructural proteins by codelivery of granulocyte macrophage-colony stimulating factor gene in intramuscular DNA immunization. Vaccine. Mar. 5, 1999;17(9-10):1136-44.

Chui, H.M. et al., Synthesis of helix 69 of *Escherichia coli* 23S rRNA containing its natural modified nucleosides, m(3) Psi and Psi. J Org Chem. Dec. 13, 2002;67(25):8847-54.

Clawson, G.A. et al., Increased amounts of double-stranded RNA in the cytoplasm of rat liver following treatment with carcinogens. Cancer Res. Aug. 1982;42(8):3228-31.

Cohen, P.J. et al., Murine epidermal Langerhans cells and splenic dendritic cells present tumor-associated antigens to primed T cells. Eur J Immunol. Feb. 1994;24(2):315-9.

Collas, P. et al., Epigenetic reprogramming of nuclei using cell extracts. Stem Cell Rev. 2006;2(4):309-17.

Binder, R. et al., Evidence that the pathway of transferrin receptor mRNA degradation involves an endonucleolytic cleavage within the 3' UTR and does not involve poly(A) tail shortening. EMBO J. Apr. 15, 1994;13(8):1969-80.

Collas, P., Dedifferentiation of cells: new approaches. Cytotherapy. 2007;9(3):236-44.

Colter, J.S., et al., Infectivity of ribonucleic acid isolated from virus-infected tissues. Virology. 1957; 4(3): 522-532.

Colot, V. et al., Eukaryotic DNA methylation as an evolutionary device. Bioessays. May 1999;21(5):402-11.

Colter, J.S., et al., Infectivity of ribonucleic acid from Ehrlich Ascites tumour cells infected with Mengo Encephalitis. Nature. Apr. 1957; 179(4565): 859-860.

Condon, C. et al., DNA-based immunization by in vivo transfection of dendritic cells. Nat Med. Oct. 1996;2(10):1122-8.

International Search Report from International Application No. PCT/US2012/054561 dated Feb. 26, 2013.

Barlow, et al., The Human Cathelicidin LL-37 Preferentially Promotes Apoptosis of Infected Airway Epithelium. Am J Respir Cell Mol Biol. Dec. 2010, vol. 43, No. 6, pp. 692-702, entire document. Evel-Kabler, Kevin et al., SOCS1 Restricts Dendritic Cells' Ability to Break Self Tolerance and Induce Antitumor Immunity by Regu-

lating IL-12 Production and Signaling, The Journal of Clinical Investigation, 2006, vol. 116, No. 1, pp. 90-100.

Finn, Jonathan et al., Eradication of Neutralizing Antibodies to Factor VIII in Canine Hemophila A After liver Gene Therapy, Blood, 2010, vol. 116, No. 26, pp. 5842-5848.

Han, Shuhong et al., Novel Autoantigens in Type 1 Diabetes, Am J Transl Res, 2013, vol. 5, No. 4, pp. 379-392.

High, Katherine, et al. The Gene Therapy Journey for Hemophilia: Are We There Yet?, Blood, 2012, vol. 120, No. 23, pp. 4482-4487. Hoffman, Brad et al., Nonredundany Roles of IL-10 and TGF-β in Supression of Immune Responses tp Hepatic AAV-Factor IX Gene Transfer, The American Society of Gene and Cell Therapy, 2011, vol. 19, No. 7, pp. 1263-1272.

Hopkins, Benjamin et al., A Secreted PTEN Phosphatase That Enters Cells to Alter Signaling and Survival, Science, 2013,vol. 341, No. 399, pp. 399-341.

Takahashi, R. et al., SOCS1 is Essential for Regulatory T Cell Functions by Preventing Loss of Foxp3 Expression as Well AsIFN-y and IL-17A Production, The Journal of Experimental Medicine, 2011, vol. 208, No. 10, pp. 2055-2067.

Piganis, R. et al., Suppressor of Cyokine Signaling (SOCS) 1 Inhibits Type 1 Interferon (IFN) Signaling via the Interferon a Receptor (IFNAR1)-associated Tyrosine Kinase Tyk2, The Journal of Biological Chemistry, vol. 286, No. 39, pp. 33811-33818.

Jacobsen, Lars et al., Allergen-specific Immunotherapy Provide Immediate, Long-Term and Preventive Clinical Effects in Children and Adults: The Effects of Immunotherapy Can be Categorised by Level of Benefit—the centenary of Allergen Specific Subcutaneous Immunotherapy, Clinical and Translational Allergen, 2012, vol. 2, No. 8, pp. 1-11.

Kinjyo, Ichiko et al., SOCS1/JAB is A Negative Regulator of LPD-Induced Macrophage Activation, Immunity, 2002, vol. 17, No number, pp. 583-591.

LoDuca, Paul et al., Hepatic Gene Transfer as a Means of Tolerance Induction to Transgene Products, Curr Gene Ther. 2009, vol. 9, No. 2, pp. 104-114.

Lu, Li-Fan et al., Foxp3-Dependent MicroRNA 155 Confers Competitive Fitness to Regulatory T Cells by Targeting SOCS1 Protein, CellPress, Immunity, 2008, No Volume Number, pp. 80-91.

Luo, Xunrong et al., Dendritic Cells with TGF-B1 Differentiate naïve CD4=CD25-T Cells Into Islet-Protective Foxp3+ Regulatory T Cells, PNAS, 2007, vol. 104, No. 8, pp. 2821-2826.

Mingozzi, Federico, et al., Pharmacological Modulation of Humoral Immunity in a Nonhuman Primate Model AAV Gene Transfer for Hemophilia B, The American Society of Gene & Cell Therapy, 2012, vol. 20, No. 7, pp. 1410-1416.

Peakman, Mark et al., Can We Vaccinate Against Type 1 Diabetes, F1000Reports Biology, 2012, No Volume no., pp. 1-8.

Roep, Bart et al., Antigen Targets of Type 1 Diabetes Autoimmunity, Cold Spring Harbor Perspectives in Medicine, 2013, No Vol., pp. 1-15

Suciu-Foca, Nicole et al., Soluble IG-Like Transcript 3 Inhibits Tumor Allograft Rejection in Humanized SCID Mice and T Cell Responses in Cancer Patients, The Journal of Immunology, 2007, vol. 178, pp. 4732-7441.

Vlad, George et al., Immunoglobulin-Like Transcript 3-FC Suppresses T-Cell Responses to Allogeneic Human Islet Transplants in hu-NOD/SCID Mice, Diabetes, 2006, vol. 57, No number, pp. 1-9. Wantabee, Hisayo et al., Experimental Autoimmune Thyroiditis Induced b Thyroglobulin-Pulsed Dendritic Cells, 1999, vol. 31, No. 4, pp. 273-282.

Wing, Kajsa et al., Regulatory T Cells Exert Checks and Balances on Self Tolerance and Autoimmunity, Nature Immunology, 2010, vol. 11, No. 1, pp. 1-8.

Yang, Junbao et al., CD+Tcells from Type 1 Diabetic and Healthy Subjects Exhibit Different Thresholds of Activation to a Naturally Processed Proinsulin Epitope, Journal of Autoimmunity, 2008, vol. 31, No Vol. number, pp. 30-41.

Taniguchi, Takashi et al., Serum Levels of Galectin-3: Possible Association with Fibrosis, Aberrant Angiogenesis, and Immune Activation in Patients with Systemic Sclerosis, The Journal of Rheumatology, 2012, vol. 39, No. 3, pp. 539-544.

OTHER PUBLICATIONS

Chen, Juine-Ruey, et al., Vaccination of Monoglycosylated Hemagglutinin Induces Cross-Strain Protection Against Influenza Virus Infection, PNAS, 2013, No Volume Number, pp. 1-6.

Apostolopoulos, Vasso et al., Targeting Antigens to Dendritic Cell Receptors for Vaccine Development, Hindawi Publishing Corporation Journal of Drug Delivery, 2013, vol. 201, Article ID 869718, pp. 1-22.

Deering, Raquel et al., Nucleic Acid Vaccines: Prospects for Non-Viral Delivery of mRNA Vaccines, Expert Opinion, 2014, vol. 11, No. 6, pp. 1-15.

Falugi, Fabiana et al., Role of Protien A in the Evasion of Host Adaptive Immune Responses by *Staphylococcus aureus*, mBio, 2014, vol. 4, Issue 5, pp. 1-10.

Geijtenbeek, Teunis et al., Identification of DC-SIGN, A Novel Dendritic Cell-Specific ICAM-3 Receptor That Supports Primary Immune Responses, Cell, 2000, vol. 100, pp. 575-585.

World Health Organization, Department of Communicable Disease Surveillance and Response, WHO/CSR, 2000, Chapter 7, pp. 1-7. Gupta, Shivali et al., TcVac3 Induced Control of Trypanosoma Cruzi Infection and Chronic Myocarditis in Mice, PLOS One, 2013, vol. 8, Issue 3, pp. 1-16.

Nogueira, Raquel et al., Recombinant Yellow Fever Viruses Elicit CD8+ T Cell Responses and Protective Immunity Against Trypanosoma Cruzi, PLOS One, 2013, vol. 8, Issue 3, pp. 1-13. Barr, Ian et al., Epidemiological, Antigen and Genetic Characteristics of Seasonal Influenza a(H1N1), A (H3N2) and B Influenza Virus: Basis for WHO Recommendation on the Competition of Influenza Vaccines for Using in the 2009-2010 Northern Hemisphere Season, Vaccine, 2010, vol. 28, No number, pp. 1156-1167. Kim, Hwan Keun et al., Nontoxigenic Protein A Vaccine for Methicillin-Resistant Staphylococcus Aureus Infections in Mice, The Journal of Experimental Medicine, 2010, vol. 207, No. 9, pp. 1863-1870.

Lee, Justin B. et al., Lipid Nanoparticle siRNA Systems for Silencing the Androgen Receptor in Human Prostate Cancer in Vivo, International Journal of Cancer, 2012, vol. 131, pp. 781-790.

Brandenburg, Boerries et al., Mechanisms of Hemagglutinin Targeted Influenza Virus Neutralization, PLOS One, 2013, vol. 8, Issue 12, pp. 1-14.

Messer, William B. et al., Dengue Virus Envelope Protein Domain I/II Hinge Determines long-livid Serotype-Specific Dengue Immunity, PNAS, 2014, vol. 111, No. 5, 1939-1944.

Metz, Bernard et al, Identification of Formaldehyde-induced Modifications in Proteins, The Journal of Biological Chemistry, 2004,vol. 279, No. 8, pp. 6235-6243.

Mohamadzadeh, M et al., Dendritic Cell Targeting of Bacillus Anthracis Protective Antigen Expressed by Lactobacillus Acidophilus Protects Mice From Lethal Challenge, PNAS, 2009, vol. 106, No. 11, pp. 4331-4336.

Perez-Velez, Mariel et al., Induction of Neutralization Antibodies in Mice by Dengue-2 Envelope DNA Vaccines, National Institutes of Health, PR Health Sci, 2009, vol. 28, No. 3, pp. 239-250.

Ramanathan, Mathura et al., Development of Novel DNA SynCon Tetravalent Dengue Vaccine That Elicits Immune Responses Against Four Serotypes, Vaccine, 2009, vol. 27, No Number, pp. 6444-6453.

Schroeder, Ulrich et al., Peptide Nanoparticles Serve as a Powerful Platform for the Immunogenic Display of Poorly Antigenic Actin Determinants, Science Direct, J. Mol. Biol., 2009, vol. 386, No vol. Number, pp. 1368-1381.

Arce-Fonseca, Minerva et al., Specific Humoral and Cellular Immunity Induced by Trypanosoma cruzi DNA Immunization in a Canine Model, Veterinary Research, 2013, vol. 44, No. 15, pp. 2-9.

Steel, John et I., Influenza Virus Vaccine Based on the Conserved Hemagglutinin Stalk Domain, mBio, 2010, vol. 1, Issue 1, pp. 1-10. Walker, Andreas et al., SplitCore: An Exceptionally Versatile Viral NanoParticles for Native Whole Protein Display Regardless of 3D Structure, Scientific Reporters, 2011, vol. 1, No. 5, pp. 1-8.

World Health Organization, WHO Manual on Animal Influenza Diagnosis and Surveillance, WHO Global Influenza Programme, CDS, CSR, NCS, 2002, vol. 5, No Number, pp. 1-99.

World Health Organization, Serological Diagnosis of Influenza by Microneutralization Assay, 2010, No Vol., pp. 1-25.

Coller, Barry S. et al, A New Murine Monoclonal Antibody Reports an Activation-Dependent Change in the Confirmation and/or Microenvironment of the Platelet Glycoprotein IIb/IIIa Complex, The American Society for Clinical Investigation, Inc., 1985, vol. 76, No Volume number, pp. 101-108.

Coller, BS et al., Inhibition of Dog Platelet Function by Vivo Infusion of F (ab')2 Fragments of a Monoclonal Antibody to Platelet Glycoprotien IIb/IIIa Receptor, Blood, 1985, vol. 66, No. 6, pp. 1456-1459.

Ellis, SG et al., Safety and Antiplatelet Effect of Murine Monoclonal Antibody 7E3 Fab Directed Against Platelet Glycoprotein IIb/IIIA in Patients Undergoing Elective Coronary Angioplasty, Coron Artery Dis., 1993, vol. 4, No. 2, pp. 167-175.

Abciximab (ReoPro)FDA Description, Jan. 4, 1997, No Volume number, pp. 1-17.

Califf, Robert et al., Use of a Monoclonal Antibody Directed Against the Platelet Glycoprotein IIB/IIIa Receptor in High-Risk Coronary Angioplasty, 1994, The New England Journal of Medicine, vol. 330, No. 14, pp. 1-6.

Bolukbasi et al., miR-1289 and "Zipcode"-like Sequence Enrich mRNAs in Microvesicles. Mol Ther Nucleic Acids. Feb. 7, 2012;1:e10. doi: 10.1038/mtna.2011.2.

Chappell et al., Biochemical and functional analysis of a 9-nt RNA sequence that affects translation efficiency in eukaryotic cells. Proc Natl Acad Sci U S A. Jun. 29, 2004;101(26):9590-4. Epub Jun. 21, 2004.

Fan et al., Overexpression of HuR, a nuclear-cytoplasmic shuttling protein, increases the in vivo stability of ARE-containing mRNAs. EMBO J. Jun. 15, 1998;17(12):3448-60.

Karijolich et al., Converting nonsense codons into sense codons by targeted pseudouridylation. Nature. Jun. 15, 2011;474(7351):395-8. doi: 10.1038/nature10165.

Kedde et al., A Pumilio-induced RNA structure switch in p27-' UTR controls miR-221 and miR-222 accessibility. Nat Cell Biol. Oct. 2010;12(10):1014-20. doi: 10.1038/ncb2105. Epub Sep. 5, 2010.

Kore et al., Synthesis and biological validation of N^7 -(4-chlorophenoxyethyl) substituted dinucleotide cap analogs for mRNA translation. Bioorg Med Chem. Aug. 1, 2013;21(15):4570-4. doi: 10.1016/j.bmc.2013.05.041. Epub Jun. 1, 2013.

Leppek et al., Roquin promotes constitutive mRNA decay via a conserved class of stem-loop recognition motifs. Cell. May 9, 2013;153(4):869-81. doi: 10.1016/j.cel1.2013.04.016.

Matsuda et al., Determinants of initiation codon selection during translation in mammalian cells. PLoS One. Nov. 24, 2010;5(11):e15057. doi: 10.1371/journal.pone.0015057.

Meijer et al., Translational repression and eIF4A2 activity are critical for microRNA-mediated gene regulation. Science. Apr. 5, 2013;340(6128):82-5. doi: 10.1126/science.1231197.

Parisien et al., Rationalization and prediction of selective decoding of pseudouridine-modified nonsense and sense codons. RNA. Mar. 2012;18(3):355-67. doi: 10.1261/rna.031351.111. Epub Jan. 26, 2012.

Pánek et al., An evolutionary conserved pattern of 18S rRNA sequence complementarity to mRNA 5' UTRs and its implications for eukaryotic gene translation regulation. Nucleic Acids Res. Sep. 2013;41(16):7625-34. doi: 10.1093/nar/gkt548. Epub Jun. 26, 2013. Peart et al., Non-mRNA 3' end formation: how the other half lives. Wiley Interdiscip Rev RNA. Sep.-Oct. 2013;4(5):491-506. doi: 10.1002/wrna.1174. Epub Jun. 10, 2013. Review.

Ray et al., A compendium of RNA-binding motifs for decoding gene regulation. Nature. Jul. 11, 2013;499(7457):172-7. doi: 10.1038/nature12311.

Wellensiek et al., Genome-wide profiling of human cap-independent translation-enhancing elements. Nat Methods. Aug.

OTHER PUBLICATIONS

2013;10(8):747-50. doi: 10.1038/nmeth.2522. Epub Jun. 16, 2013. Wellensiek et al., Supplementary Information for Genome-wide profiling of human cap-independent translation-enhancing elements. Nat Methods. 2013. 30 pages.

Wilusz et al., 3' end processing of a long nuclear-retained noncoding RNA yields a tRNA-like cytoplasmic RNA. Cell. Nov. 28, 2008;135(5):919-32. doi: 10.1016/j.cel1.2008.10.012. Zhou et al., A positive feedback vector for identification of nucleotide sequences that enhance translation. Proc Natl Acad Sci U S A. May 3, 2005;102(18):6273-8. Epub Apr. 21, 2005.

* cited by examiner

ENGINEERED NUCLEIC ACIDS AND METHODS OF USE THEREOF

CROSS REFERENCE TO RELATED APPLICATIONS

This application is a continuation of U.S. application Ser. No. 14/342,905, filed Mar. 5, 2014, now abandoned, which is a national phase filing under 35 U.S.C. §371 of International Application No. PCT/US2012/54561, filed Sep. 11, 10 2012, and claims priority to U.S. Provisional Application No. 61/533,537, filed Sep. 12, 2011, entitled Engineered Nucleic Acids and Methods of Use Thereof, the contents of each which are incorporated by reference in their entireties.

REFERENCE TO THE SEQUENCE LISTING

The present application is being filed along with a Sequence Listing in electronic format. The Sequence Listing is provided as a file entitled M006USCONSEQLST.txt 20 created on Nov. 4, 2014 which is 1,397,330 bytes in size. The information in electronic format of the sequence listing is incorporated herein by reference in its entirety.

REFERENCE TO LENGTHY TABLE

The specification includes a lengthy Table 1. Lengthy Table 1 has been submitted via EFS-Web in electronic format as follows: File name: Table.txt, Date created: Nov. 4, 2014; File size: 368,283 Bytes and is incorporated herein by reference in its entirety. Please refer to the end of the specification for access instructions.

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disclosure belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present disclosure, suitable methods and materials are described herein. The materials, methods, and examples are illustrative only and not intended to be limiting. Other features of the disclosure are apparent from the following detailed description and the claims.

SUMMARY OF THE INVENTION

Provided herein are modified nucleic acids encoding anti-microbial polypeptides (AMPs) (e.g., anti-bacterial polypeptides), e.g., anti-microbial polypeptides described herein, precursors thereof, or partially or fully processed forms of these precursors. In certain embodiments, the anti-microbial polypeptide is an anti-bacterial polypeptide. In certain embodiments, the anti-microbial polypeptide is an anti-fungal polypeptide. In certain embodiments, the antimicrobial polypeptide is an anti-viral polypeptide. In certain embodiments, the anti-microbial polypeptide is an antiprotozoal polypeptide. In certain embodiments, the antimicrobial polypeptide is an anti-tumor/cancer polypeptide. In certain embodiment, the anti-microbial polypeptide is an anti-parasitic polypeptide. In certain embodiment, the antimicrobial polypeptide is an anti-prion polypeptide. In certain embodiments, the anti-microbial polypeptide has one or more of anti-bacterial, anti-fungal, anti-viral, anti-protozoal, anti-tumor/cancer, anti-parasitic, or anti-prion activity. In certain embodiments, the modified nucleic acid comprises mRNA. In particular embodiments, the modified mRNA (mmRNA) is derived from cDNA. In certain embodiments,

LENGTHY TABLES

The patent contains a lengthy table section. A copy of the table is available in electronic form from the USPTO web site (http://seqdata.uspto.gov/?pageRequest=docDetail&DocID=US09464124B2). An electronic copy of the table will also be available from the USPTO upon request and payment of the fee set forth in 37 CFR 1.19(b)(3).

BACKGROUND OF THE INVENTION

Naturally occurring RNAs are synthesized from four basic ribonucleotides: ATP, CTP, UTP and GTP, but may contain post-transcriptionally modified nucleotides. Further, over one hundred natural nucleotide modifications have been identified in all RNA species (Rozenski, J, Crain, P, and McCloskey, J. (1999). The RNA Modification Database: 1999 update. Nucl Acids Res 27: 196-197). Nucleotides are modified in RNA to alter functional, structural, or catalytic roles of the parent RNA molecule. More recently, nucleotide modifications have been described to play a role in differentiating host cell RNA species from invading pathogenic RNA species. However, the precise mechanism by which nucleotide modifications alter the host immune response machinery and subsequently affect the translation efficiency of mRNA is unclear.

There is a need in the art for biological modalities to address the modulation of intracellular translation of nucleic acids.

Unless explained otherwise, all technical and scientific 65 terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which this

the mmRNA comprises at least two nucleoside modifications. In certain embodiments, these nucleoside modifications are 5-methylcytosine and pseudouridine.

Provided herein are isolated nucleic acids (e.g., modified mRNAs encoding an anti-microbial polypeptide described herein) comprising a translatable region and at least two different nucleoside modifications, wherein the nucleic acid exhibits reduced degradation in a cell into which the nucleic acid is introduced, relative to a corresponding unmodified nucleic acid. For example, the degradation rate of the nucleic acid is reduced by at least 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, or 90%, compared to the degradation rate of the corresponding unmodified nucleic acid. In certain embodiments, the nucleic acid comprises RNA, DNA, TNA, GNA, or a hybrid thereof. In certain embodiments, the nucleic acid comprises messenger RNA (mRNA). In certain embodiments, the mRNA does not substantially induce an innate immune response of the cell into which the mRNA is introduced. In certain embodiments, the mRNA comprises at least one nucleoside selected from the group consisting of pyridin-4-one ribonucleoside, 5-aza-uridine, 2-thio-5-aza-2-thiouridine, 4-thio-pseudouridine, pseudouridine, 5-hydroxyuridine, 3-methyluridine, 5-car-

1-carboxymethyl-pseudouridine, boxymethyl-uridine, 5-propynyl-uridine, 1-propynyl-pseudouridine, 5-taurinomethyluridine, 1-taurinomethyl-pseudouridine, 5-taurinom-1-taurinomethyl-4-thio-uridine, ethyl-2-thio-uridine, 5-methyl-uridine, 1-methyl-pseudouridine, 4-thio-1-methyl-5 pseudouridine, 2-thio-1-methyl-pseudouridine, 1-methyl-1deaza-pseudouridine, 2-thio-1-methyl-1-deaza-pseudouridine, dihydrouridine, dihydropseudouridine, 2-thiodihydrouridine, 2-thio-dihydropseudouridine, 2-methoxyuridine, 2-methoxy-4-thio-uridine, 4-methoxy- 10 pseudouridine, and 4-methoxy-2-thio-pseudouridine. In certain embodiments, the mRNA comprises at least one nucleoside selected from the group consisting of 5-aza-cytidine, pseudoisocytidine, 3-methyl-cytidine, N4-acetylcytidine, 5-formylcytidine, N4-methylcytidine, 5-hydroxymethylcy- 15 tidine, 1-methyl-pseudoisocytidine, pyrrolo-cytidine, pyrrolo-pseudoisocytidine, 2-thio-cytidine, 2-thio-5-methyl-cytidine, 4-thio-pseudoisocytidine, 4-thio-1-methylpseudoisocytidine, 4-thio-1-methyl-1-deaza-1-methyl-1-deaza-pseudoisocytidine, 20 pseudoisocytidine. zebularine, 5-aza-zebularine, 5-methyl-zebularine, 5-aza-2thio-zebularine, 2-thio-zebularine, 2-methoxy-cytidine, 2-methoxy-5-methyl-cytidine, 4-methoxy-pseudoisocytidine, and 4-methoxy-1-methyl-pseudoisocytidine. In other embodiments, the mRNA comprises at least one nucleoside 25 selected from the group consisting of 2-aminopurine, 2,6diaminopurine, 7-deaza-adenine, 7-deaza-8-aza-adenine, 7-deaza-2-aminopurine, 7-deaza-8-aza-2-aminopurine, 7-deaza-2,6-diaminopurine, 7-deaza-8-aza-2,6-diaminopurine, 1-methyladenosine, N6-methyladenosine, N6-isopen- 30 N6-(cis-hydroxyisopentenyl)adenosine, tenyladenosine, 2-methylthio-N6-(cis-hydroxyisopentenyl) adenosine. N6-glycinylcarbamoyladenosine, N6-threonylcarbamoyladenosine, 2-methylthio-N6-threonyl carbamoyladenosine, N6,N6-dimethyladenosine, 7-methyladenine, 2-methylthio- 35 adenine, and 2-methoxy-adenine. In yet other embodiments, the mRNA comprises at least one nucleoside selected from the group consisting of inosine, 1-methyl-inosine, wyosine, wybutosine, 7-deaza-guanosine, 7-deaza-8-aza-guanosine, 6-thio-7-deaza-guanosine, 6-thio-7- 40 6-thio-guanosine, deaza-8-aza-guanosine, 7-methyl-guanosine, 6-thio-7methyl-guanosine, 7-methylinosine, 6-methoxy-guanosine, 1-methylguanosine, N2-methylguanosine, N2,N2-dimethylguanosine, 8-oxo-guanosine, 7-methyl-8-oxo-guanosine, 1-methyl-6-thio-guanosine, N2-methyl-6-thio-guanosine, 45 and N2,N2-dimethyl-6-thio-guanosine.

In some embodiments, the nucleic acids provided herein comprise a 5' untranslated region (UTR) and/or a 3'UTR, wherein each of the two different nucleoside modifications are independently present in the 5'UTR and/or 3'UTR. In 50 some embodiments, nucleic acids are provided herein, wherein at least one of the two different nucleoside modifications are present in the translatable region. In some embodiments, nucleic acids provided herein are capable of binding to at least one polypeptide that prevents or reduces 55 an innate immune response of a cell into which the nucleic acid is introduced.

Further provided herein are isolated nucleic acids (e.g., modified mRNAs described herein) comprising (i) a translatable region encoding an anti-microbial polypeptide (e.g., 60 an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, (ii) at least one nucleoside modification, and (iii) at least one intronic nucleotide sequence capable of being excised from the nucleic acid.

Further provided herein are isolated nucleic acids (e.g., 65 modified mRNAs described herein) comprising (i) a translatable region encoding an anti-microbial polypeptide (e.g.,

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an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, (ii) at least two different nucleoside modifications, and (iii) a degradation domain.

Further provided herein are isolated nucleic acids (e.g., modified mRNAs described herein) comprising (i) a translatable region encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, and (ii) at least two different nucleoside modifications, wherein the translatable region encodes a polypeptide variant having an altered activity relative to a reference polypeptide. In certain embodiments, isolated mRNAs are provided, wherein the altered activity comprises an increased activity or wherein the altered activity comprises a decreased activity.

Further provided herein are non-enzymatically synthesized nucleic acids (e.g., modified mRNAs described herein) comprising at least one nucleoside modification, and comprising a translatable region encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein. In certain embodiments, the non-enzymatically synthesized mRNA comprises at least two different nucleoside modifications.

Further provided herein are isolated nucleic acids (e.g., modified mRNAs described herein) comprising a noncoding region and at least one nucleoside modification that reduces an innate immune response of a cell into which the nucleic acid is introduced, wherein the nucleic acid sequesters one or more translational machinery components. In certain embodiments, the isolated nucleic acids comprising a noncoding region and at least one nucleoside modification described herein are provided in an amount effective to reduce protein expression in the cell. In certain embodiments, the translational machinery component is a ribosomal protein or a transfer RNA (tRNA). In certain embodiments, the nucleic acid comprises a small nucleolar RNA (sno-RNA), microRNA (miRNA), small interfering RNA (siRNA) or Piwi-interacting RNA (piRNA).

Further provided herein are isolated nucleic acids (e.g., modified mRNAs described herein) comprising (i) a first translatable region, (ii) at least one nucleoside modification, and (iii) an internal ribosome entry site (IRES). In certain embodiments, the IRES is obtained from a picornavirus, a pest virus, a polio virus, an encephalomyocarditis virus, a foot-and-mouth disease virus, a hepatitis C virus, a classical swine fever virus, a murine leukemia virus, a simian immune deficiency virus or a cricket paralysis virus. In certain embodiments, the isolated nucleic acid further comprises a second translatable region. In certain embodiments, the isolated nucleic acid further comprises a Kozak sequence. In some embodiments, the first translatable region encodes an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein. In some embodiments, the second translatable region encodes an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein. In some embodiments, the first and the second translatable regions encode anti-microbial polypeptides (e.g., an anti-bacterial polypeptides), e.g., anti-microbial polypeptides described herein.

Further, provided herein are compositions (e.g., pharmaceutical compositions) comprising the modified nucleic acids described herein. In certain embodiments, the composition further comprises a pharmaceutically acceptable carrier. In certain embodiments, the composition is formulated for systemic or local administration. In certain embodiments, the composition is formulated for intravenous administration. In certain embodiments, the composition is for-

mulated for oral administration. In certain embodiments, the composition is formulated for topical administration. In certain embodiments, the composition is formulated for administration via a dressing (e.g., wound dressing). In certain embodiments, the composition is formulated for administration via a bandage (e.g., adhesive bandage). In certain embodiments, the composition is formulated for administration by inhalation. In certain embodiments, the composition is formulated for rectal administration. In certain embodiments, the composition is formulated for vaginal administration. In certain embodiments, the composition comprises naked modified nucleic acids. In other embodiments, the modified nucleic acid is complexed or encapsulated. In another embodiment, the administration of the composition described herein may be administered at least

Provided herein are pharmaceutical compositions comprising: (i) an effective amount of a synthetic messenger ribonucleic acid (mRNA) encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an antimicrobial polypeptide described herein; and (ii) a pharma- 20 ceutically acceptable carrier, wherein i) the mRNA comprises pseudouridine, 5'methyl-cytidine, or a combination thereof, or ii) the mRNA does not comprise a substantial amount of a nucleotide or nucleotides selected from the group consisting of uridine, cytidine, and a combination of 25 uridine and cytidine, and wherein the composition is suitable for repeated administration (e.g., intravenous administration) to a mammalian subject in need thereof. In some embodiments, the anti-microbial polypeptide (e.g., antibacterial polypeptide) is under 10 kDa, e.g., under 8 kDa, 6 30 kDa, 4 kDa, 2 kDa, or 1 kDa. In some embodiments, the anti-microbial polypeptide (e.g., anti-bacterial polypeptide) comprises or consists of from about 6 to about 100 amino acids, e.g., from about 6 to about 75 amino acids, about 6 to about 50 amino acids, about 6 to about 25 amino acids, about 35 25 to about 100 amino acids, about 50 to about 100 amino acids, or about 75 to about 100 amino acids. In certain embodiments, the anti-microbial polypeptide (e.g., antibacterial polypeptide) comprises or consists of from about anti-microbial polypeptide (e.g., anti-bacterial polypeptide) is substantially cationic. In some embodiments, the antimicrobial polypeptide (e.g., anti-bacterial polypeptide) is substantially amphipathic. In certain embodiments, the antimicrobial polypeptide (e.g., anti-bacterial polypeptide) is 45 substantially cationic and amphipathic. In some embodiments, the anti-microbial polypeptide (e.g., anti-bacterial polypeptide) is cytostatic to a Gram-positive bacterium. In some embodiments, the anti-microbial polypeptide (e.g., anti-bacterial polypeptide) is cytotoxic to a Gram-positive 50 bacterium. In some embodiments, the anti-microbial polypeptide (e.g., anti-bacterial polypeptide) is cytostatic and cytotoxic to a Gram-positive bacterium. In some embodiments, the anti-microbial polypeptide (e.g., anti-bacterial polypeptide) is cytostatic to a Gram-negative bacterium. In 55 some embodiments, the anti-microbial polypeptide (e.g., anti-bacterial polypeptide) is cytotoxic to a Gram-negative bacterium. In some embodiments, the anti-microbial polypeptide (e.g., anti-bacterial polypeptide) is cytostatic and cytotoxic to a Gram-negative bacterium. In some embodi- 60 ments, the anti-microbial polypeptide is cytostatic to a virus, fungus, protozoan, parasite, prion, or a combination thereof. In some embodiments, the anti-microbial polypeptide is cytotoxic to a virus, fungus, protozoan, parasite, prion, or a combination thereof. In certain embodiments, the anti-mi- 65 crobial polypeptide is cytostatic and cytotoxic to a virus, fungus, protozoan, parasite, prion, or a combination thereof.

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In some embodiments, the anti-microbial polypeptide is cytotoxic to a tumor or cancer cell (e.g., human tumor or cancer cell). In some embodiments, the anti-microbial polypeptide is cytostatic to a tumor or cancer cell (e.g., human tumor or cancer cell). In certain embodiments, the antimicrobial polypeptide is cytotoxic and cytostatic to a tumor or cancer cell (e.g., human tumor or cancer cell). In some embodiments, the anti-microbial polypeptide (e.g., antibacterial polypeptide) is a secreted polypeptide. In certain embodiments, the anti-microbial polypeptide (e.g., antibacterial polypeptide) is selected from the group consisting of anti-microbial polypeptides (e.g., anti-bacterial polypeptides) and/or SEQ ID NOs: 1-2915. In certain embodiments, the anti-microbial polypeptide (e.g., anti-bacterial polypeptide) comprises or consists of hBD-2 (SEQ ID NO: 191 or 192), LL-37 (SEQ ID NO: 6), or RNase-7 (SEQ ID NO: 262). In some embodiments, the composition (e.g., pharmaceutical composition) provided herein further comprises a lipid-based transfection reagent. In some embodiments, the synthetic messenger ribonucleic acid (mRNA) encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, lacks at least one destabilizing element.

Further provided herein are pharmaceutical compositions comprising and/or consisting essentially of: (i) an effective amount of a synthetic messenger ribonucleic acid (mRNA) encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein; (ii) a cell penetration agent; and (iii) a pharmaceutically acceptable carrier, wherein i) the mRNA comprises pseudouridine, 5'methyl-cytidine or a combination thereof, or ii) the mRNA does not comprise a substantial amount of a nucleotide or nucleotides selected from the group consisting of uridine, cytidine, and a combination of uridine and cytidine, and wherein the composition is suitable for repeated administration (e.g., intravenous administration) to an animal (e.g., mammalian) subject in need thereof.

Provided herein are methods of treating a subject having 15 to about 45 amino acids. In some embodiments, the 40 and/or being suspected of having a microbial infection (e.g., a bacterial infection) and/or a disease, disorder, or condition, e.g., a disease, disorder, or condition associated with a microbial infection (e.g., a bacterial infection), the methods comprising administering to a subject in need of such treatment a composition described herein in an amount sufficient to treat the microbial infection and/or the disease. disorder, or condition. In specific embodiments, the disease, disorder, or condition is associated with one or more cellular and/or molecular changes affecting, for example, the level, activity, and/or localization of an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, precursors thereof, or a partially or fully processed form of these precursors. In certain embodiments, the method of treating a subject having or being suspected of having a microbial infection (e.g., a bacterial infection) and/or a disease, disorder, or condition, e.g., a disease, disorder, or condition associated with a microbial infection (e.g., a bacterial infection), comprises administering to the subject in need of such treatment a composition comprising a modified nucleic acid described herein in an amount sufficient to kill or reduce the growth of microorganisms (e.g., bacteria, fungi, viruses, protozoan, parasites, prions, or a combination thereof), to kill or reduce the growth of tumor/cancer cells, and/or to modulate one or more activities associated with, therefore to treat the microbial infection and/or the disease, disorder, or condition in the subject.

Further provided herein are methods of treating and/or preventing a microbial infection (e.g., a bacterial infection) of a target animal cell (e.g., mammalian cell), comprising the step of contacting the target animal cell (e.g., mammalian cell) with a composition comprising a synthetic messenger 5 ribonucleic acid (mRNA) encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide) in an amount effective to be cytostatic and/or cytotoxic to one or more microorganisms (e.g., bacteria) infecting the target animal cell (e.g., mammalian cell). In some embodiments, the 10 composition is effective to be cytostatic and/or cytotoxic to one or more microorganisms (e.g., bacteria) adjacent to the target animal cell (e.g., mammalian cell). In some embodiments, the target animal cell (e.g., mammalian cell) is present in an animal subject (e.g., a mammalian subject). In 15 certain embodiments, the subject is a human. In certain embodiments, the subject is a livestock animal. In some embodiments, the composition is administered to the subject by an intravenous route. In certain embodiments, the composition is administered to the subject orally. In certain 20 embodiments, the composition is administered to the subject topically. In certain embodiments, the composition is administered to the subject by inhalation. In certain embodiments, the composition is administered to the subject rectally. In certain embodiments, the composition is administered to the 25 subject vaginally. In certain embodiments, the method further comprises the step of administering an effective amount of an anti-microbial agent (e.g., an anti-bacterial agent), e.g., an anti-microbial agent described herein, to the subject at the same time or at a different time from the administering the 30 composition, e.g., before or after the administering the composition. In some embodiments, the anti-microbial agent is an anti-microbial polypeptide, e.g., a microbial polypeptide described herein. In some embodiments, the anti-microbial agent is a small molecule anti-microbial 35 agent, e.g., a small molecule anti-microbial agent described herein. In another embodiment, the administration of the composition described herein may be administered at least

Further provided herein are methods for treating and/or 40 preventing a microbial infection (e.g., a bacterial infection) and/or a disease, disorder, or condition associated with a microbial infection (e.g., a bacterial infection), and/or a symptom thereof, in a animal (e.g., a mammalian) subject, comprising contacting a cell of the subject with a nucleic 45 acid described herein, wherein the translatable region of the nucleic acid encodes an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), under conditions such that an effective amount of the anti-microbial polypeptide (e.g., an anti-bacterial polypeptide) is present in the cell, thereby 50 treating or preventing a microbial infection (e.g., bacterial infection) and/or a disease, disorder, or condition associated with the microbial infection (e.g., bacterial infection), and/or a symptom thereof, in the subject. In certain embodiments, the cell is an epithelial cell, an endothelial cell, or a 55 mesothelial cell. In certain embodiments, the nucleic acid comprises an RNA molecule formulated for administration by an intravenous route. In certain embodiments, the nucleic acid comprises an RNA molecule formulated for oral administration. In certain embodiments, the nucleic acid comprises 60 an RNA molecule formulated for topical administration. In certain embodiments, the nucleic acid comprises an RNA molecule formulated for administration by inhalation. In certain embodiments, the nucleic acid comprises an RNA molecule formulated for rectal administration. In certain 65 embodiments, the nucleic acid comprises an RNA molecule formulated for vaginal administration.

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Further provided herein are methods for inducing in vivo translation of a recombinant polypeptide (e.g., an antimicrobial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein) in an animal (e.g., a mammalian) subject in need thereof, comprising the step of administering to the subject an effective amount of a composition comprising a nucleic acid comprising: (i) a translatable region encoding the recombinant polypeptide; and (ii) at least one nucleoside modification, under conditions such that the nucleic acid is localized into a cell of the subject and the recombinant polypeptide is capable of being translated in the cell from the nucleic acid. In certain embodiments, the composition comprises mRNA. In certain embodiments, methods are provided, wherein the recombinant polypeptide comprises a functional activity substantially absent in the cell in which the recombinant polypeptide is translated. In certain embodiments, the recombinant polypeptide comprises a polypeptide substantially absent in the cell in the absence of the composition. In certain embodiments, the recombinant polypeptide comprises a polypeptide that antagonizes the activity of an endogenous protein present in, on the surface of, or secreted from the cell. In certain embodiments, the recombinant polypeptide comprises a polypeptide that antagonizes the activity of a biological moiety present in, on the surface of, or secreted from the cell. In certain embodiments, the biological moiety comprises a lipid, a lipoprotein, a nucleic acid, a carbohydrate, or a small molecule toxin. In certain embodiments, the recombinant polypeptide is capable of being secreted from the cell. In certain embodiments, the recombinant polypeptide is capable of being translocated to the plasma membrane of the cell. In certain embodiments, methods are provided, wherein the composition is formulated for administration intramuscularly, transarterially, intraperitoneally, intravenously, intranasally, subcutaneously, endoscopically, transdermally, or intrathecally. In certain embodiments, methods are provided, wherein the composition is formulated for extended release.

Further provided herein are methods for inducing translation of a recombinant polypeptide (e.g., an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein) in a cell population, comprising the step of contacting the cell population with an effective amount of a composition comprising a nucleic acid comprising: (i) a translatable region encoding the recombinant polypeptide; and (ii) at least one nucleoside modification, under conditions such that the nucleic acid is localized into one or more cells of the cell population and the recombinant polypeptide is translated in the cell from the nucleic acid. In certain embodiments, methods are provided, wherein the composition comprises mRNA. In certain embodiments, the composition comprises a cell penetrating compound. In certain embodiments, methods are provided, wherein the step of contacting the cell with the composition is repeated one or more times. In certain embodiments, the step of contacting the cell with the composition is repeated a sufficient number of times such that a predetermined efficiency of protein translation in the cell population.

Further provided herein are methods of reducing the innate immune response of a cell to an exogenous nucleic acid (e.g., a modified mRNA described herein), comprising the steps of: (a) contacting the cell with a first composition comprising a first dose of a first exogenous nucleic acid comprising a translatable region (e.g., encoding an antimicrobial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein) and at least one nucleoside modification; (b) determining the level

of the innate immune response of the cell to the first exogenous nucleic acid; (c) contacting the cell with a second composition comprising either: (i) a second dose of the first exogenous nucleic acid, wherein the second dose contains a lesser amount of the first exogenous nucleic acid as compared to the first dose; or (ii) a first dose of a second exogenous nucleic acid, thereby reducing the innate immune response of the cell. In certain embodiments, methods are provided, wherein the step of contacting the cell with the first composition and/or the second composition is repeated one or more times. In certain embodiments, a predetermined efficiency of protein translation in the cell is achieved.

Provided herein are methods of providing a composition (e.g., a composition described herein) to a target tissue of a subject (e.g., a mammalian subject) in need thereof, com- 15 prising the step of contacting the target tissue comprising one or more target cells with the composition under conditions such that the composition is substantially retained in the target tissue, and wherein the composition comprises: (a) an effective amount of a ribonucleic acid, wherein the 20 ribonucleic acid is engineered to avoid an innate immune response of a cell into which the ribonucleic acid enters, and wherein the ribonucleic acid comprises a nucleotide sequence encoding a polypeptide of interest (e.g., a antimicrobial polypeptide described herein), wherein the protein 25 of interest has an anti-microbial activity; (b) optionally, a cell penetration agent; and (c) a pharmaceutically acceptable carrier, under conditions such that the polypeptide of interest is produced in at least one target cell.

Further provided herein are isolated polypeptides (e.g., 30 anti-microbial polypeptides (e.g., anti-bacterial polypeptides), e.g., anti-microbial polypeptides described herein) produced by translation of the mRNAs described herein.

Further provided herein are isolated complexes comprising a conjugate of a protein and a nucleic acid (e.g., a nucleic acid described herein), comprising (i) an mRNA comprising a translatable region encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, and at least two different nucleoside modifications; and (ii) one or more polypeptides 40 bound to the mRNA in an amount effective to prevent or reduce an innate immune response of a cell into which the complex is introduced.

Further provided herein are libraries comprising a plurality of polynucleotides, wherein the polynucleotides indi- 45 vidually comprise: (i) a nucleic acid sequence encoding a polypeptide (e.g., an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein); and (ii) at least one nucleoside modification. In certain embodiments, libraries are provided, 50 wherein the polypeptide comprises an antibody or functional portion thereof. In certain embodiments, libraries are provided, wherein the polynucleotides comprise mRNA. In certain embodiments, libraries are provided, wherein the at least one nucleoside modification is selected from the group 55 consisting of pyridin-4-one ribonucleoside, 5-aza-uridine, 2-thio-5-aza-uridine, 2-thiouridine, 4-thio-pseudouridine, 2-thio-pseudouridine, 5-hydroxyuridine, 3-methyluridine, 5-carboxymethyl-uridine, 1-carboxymethyl-pseudouridine, 5-propynyl-uridine, 1-propynyl-pseudouridine, 5-taurinom- 60 ethyluridine, 1-taurinomethyl-pseudouridine, 5-taurinomethyl-2-thio-uridine, 1-taurinomethyl-4-thio-uridine, 5-methyl-uridine, 1-methyl-pseudouridine, 4-thio-1-methylpseudouridine, 2-thio-1-methyl-pseudouridine, 1-methyl-1deaza-pseudouridine, 2-thio-1-methyl-1-deaza-pseudouri- 65 dihydrouridine, dihydropseudouridine, dihydrouridine, 2-thio-dihydropseudouridine,

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2-methoxyuridine, 2-methoxy-4-thio-uridine, 4-methoxypseudouridine, 4-methoxy-2-thio-pseudouridine, 5-aza-cytidine, pseudoisocytidine, 3-methyl-cytidine, N4-acetylcyti-5-formylcytidine, N4-methylcytidine, dine. 5-hydroxymethylcytidine, 1-methyl-pseudoisocytidine, pyrrolo-cytidine, pyrrolo-pseudoisocytidine, 2-thio-cytidine, 2-thio-5-methyl-cytidine, 4-thio-pseudoisocytidine, 4-thio-1-methyl-pseudoisocytidine, 4-thio-1-methyl-1-deaza-1-methyl-1-deaza-pseudoisocytidine, pseudoisocytidine, zebularine, 5-aza-zebularine, 5-methyl-zebularine, 5-aza-2thio-zebularine, 2-thio-zebularine, 2-methoxy-cytidine, 2-methoxy-5-methyl-cytidine, 4-methoxy-pseudoisocytidine, 4-methoxy-1-methyl-pseudoisocytidine, 2-aminopurine, 2,6-diaminopurine, 7-deaza-adenine, 7-deaza-8-azaadenine, 7-deaza-2-aminopurine, 7-deaza-8-aza-2aminopurine, 7-deaza-2,6-diaminopurine, 7-deaza-8-aza-2, 6-diaminopurine, 1-methyladenosine, N6-methyladenosine, N6-isopentenyladenosine, N6-(cis-hydroxyisopentenyl)adenosine, 2-methylthio-N6-(cis-hydroxyisopentenyl) adenosine, N6-glycinylcarbamoyladenosine, N6-threonylcarbam-2-methylthio-N6-threonyl carbamoyladenosine, N6,N6-dimethyladenosine, 7-methyladenine, 2-methylthio-adenine, 2-methoxy-adenine, inosine, 1-methyl-inosine, wyosine, wybutosine, 7-deaza-guanosine, 7-deaza-8-aza-guanosine, 6-thio-guanosine, 6-thio-7-deazaguanosine, 6-thio-7-deaza-8-aza-guanosine, guanosine, 6-thio-7-methyl-guanosine, 7-methylinosine, 6-methoxy-guanosine, 1-methylguanosine, N2-methylguanosine, N2,N2-dimethylguanosine, 8-oxo-guanosine, 7-methyl-8-oxo-guanosine, 1-methyl-6-thio-guanosine, N2-methyl-6-thio-guanosine, and N2,N2-dimethyl-6-thioguanosine.

Further provided herein are methods for enhancing protein (e.g., an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein) product yield in a cell culture process, comprising the steps of: (a) providing a cell culture comprising a plurality of host cells; (b) contacting the cell culture with a composition comprising a nucleic acid comprising a translatable region encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein), and at least one nucleoside modification, wherein the nucleic acid exhibits increased protein production efficiency in a cell culture into which the nucleic acid is introduced, relative to a corresponding unmodified nucleic acid. In certain embodiments, methods are provided, wherein the increased protein production efficiency comprises increased cell transfection. In certain embodiments, the increased protein production efficiency comprises increased protein translation from the nucleic acid. In certain embodiments, the increased protein production efficiency comprises decreased nucleic acid degradation. In certain embodiments, the increased protein production efficiency comprises reduced innate immune response of the host cell. In certain embodiments, methods are provided, wherein the cell culture comprises a fed-batch mammalian cell culture process.

Further provided herein are methods for optimizing expression of an engineered protein (e.g., an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein) in a target cell, comprising the steps of: (a) providing a plurality of target cell types; (b) independently contacting with each of the plurality of target cell types an isolated nucleic acid comprising a translatable region encoding an engineered polypeptide and at least one nucleoside modification; and (c) detecting the presence and/or level of the engineered poly-

peptide in the plurality of target cell types, thereby optimizing expression of an engineered polypeptide in a target cell. In certain embodiments, the engineered polypeptide comprises a post-translational modification. In certain embodiments, the engineered polypeptide comprises a tertiary structure. In certain embodiments, methods are provided, wherein the target cell comprises a mammalian cell line.

Further provided herein are methods of antagonizing a biological pathway in a cell, e.g., a biological pathway associated with a microbial infection (e.g., a bacterial infection), comprising the step of contacting the cell with an effective amount of a composition comprising a nucleic acid comprising: (i) a translatable region encoding a recombinant polypeptide (e.g., an anti-microbial polypeptide (e.g., an 15 anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein); and (ii) at least one nucleoside modification, under conditions such that the nucleic acid is localized into the cell and the recombinant polypeptide is capable of being translated in the cell from the nucleic acid, 20 wherein the recombinant polypeptide inhibits the activity of a polypeptide functional in the biological pathway. In certain embodiments, methods are provided, wherein the biological pathway is defective in a cell having a microbial infection (e.g., a bacterial infection) and/or in a disease, disorder or 25 condition (e.g., a disease, disorder, or condition described herein) associated with a microbial infection (e.g., a bacterial infection).

Further provided herein are methods of agonizing a biological pathway in a cell, e.g. a biological pathway 30 associated with a microbial infection (e.g., a bacterial infection), comprising the step of contacting the cell with an effective amount of a composition comprising a nucleic acid comprising: (i) a translatable region encoding a recombinant polypeptide (e.g., an anti-microbial polypeptide (e.g., an 35 anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein); and (ii) at least one nucleoside modification, under conditions such that the nucleic acid is localized into the cell and the recombinant polypeptide is capable of being translated in the cell from the nucleic acid, 40 wherein the recombinant polypeptide induces the activity of a polypeptide functional in the biological pathway. In certain embodiments, the agonized biological pathway modulates an anti-microbial (e.g., anti-bacterial) activity. In certain embodiments, the biological pathway is reversibly agonized. 45

Further provided herein are methods for enhancing nucleic acid delivery into a cell population, comprising the steps of: (a) providing a cell culture comprising a plurality of host cells; (b) contacting the cell population with a composition comprising an enhanced nucleic acid compris- 50 ing a translatable region encoding a polypeptide (e.g., an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein) and at least one nucleoside modification, wherein the enhanced nucleic acid exhibits enhanced retention in the cell 55 population, relative to a corresponding unmodified nucleic acid. In certain embodiments, methods are provided, wherein the retention of the enhanced nucleic acid is at least about 50% greater than the retention of the unmodified nucleic acid. In some embodiments, the retention of the 60 enhanced nucleic acid is at least about 100% greater than the retention of the unmodified nucleic acid. In other embodiments, the retention of the enhanced nucleic acid is at least about 200% greater than the retention of the unmodified nucleic acid. In certain embodiments, methods are provided, 65 wherein the step of contacting the cell with the composition is repeated one or more times.

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Further provided herein are methods of nucleic acid co-delivery into a cell population, comprising the steps of:
(a) providing a cell culture comprising a plurality of host cells; (b) contacting the cell population with a composition comprising: (i) a first enhanced nucleic acid comprising a translatable region encoding a polypeptide (e.g., an antimicrobial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein) and at least one nucleoside modification; and (ii) a first unmodified nucleic acid, wherein the composition does not substantially induce an innate immune response of the cell population.

Further provided herein are methods of nucleic acid delivery into a cell population, comprising the steps of: (a) providing a cell culture comprising a plurality of host cells; (b) contacting the cell population with a first composition comprising: (i) a first enhanced nucleic acid comprising a translatable region encoding a recombinant polypeptide (e.g., an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein) and at least one nucleoside modification; and (ii) a first unmodified nucleic acid, wherein the composition does not substantially induce an innate immune response of the cell population; and (c) contacting the cell population with a second composition comprising a first unmodified nucleic acid.

Further provided herein are kits comprising a composition (e.g., a pharmaceutical composition) comprising a modified mRNA encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, in one or more containers, and instructions for use thereof.

Further provided here are kits for polypeptide production in a subject (e.g., a mammalian subject) suffering from or at risk of developing a microbial infection, comprising a first isolated nucleic acid comprising a translatable region and a nucleic acid modification, wherein the nucleic acid is capable of evading an innate immune response of a cell of the subject into which the first isolated nucleic acid is introduced, wherein the translatable region encodes a therapeutic polypeptide, e.g., a therapeutic polypeptide comprising an anti-microbial activity (e.g., a anti-microbial polypeptide described herein), and packaging and instructions therefore. In some embodiments, the instructions comprise instructions for the repeated administration of the first isolated nucleic acid to a cell or a population of cells. In some embodiments, the therapeutic polypeptide is useful in the treatment of an infection in the mammalian subject by a microbial pathogen. In some embodiments, the kit further comprises a second isolated nucleic acid comprising a translatable region. In some embodiments, the translatable region in the second isolated nucleic acid encodes an antimicrobial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein. In some embodiments, the translatable region of the second isolated nucleic acid encodes the same anti-microbial polypeptide as the first isolated nucleic acid. In some embodiments, the translatable region of the second isolated nucleic acid encodes a different anti-microbial polypeptide than the first isolated nucleic acid. In some embodiments, the second nucleic acid comprises a nucleic acid modification. In some embodiments, the second nucleic acid does not comprise a nucleic acid modification.

Further provided herein are dressings (e.g., wound dressings) or bandages (e.g., adhesive bandages) comprising a pharmaceutical composition comprising a modified mRNA

encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein.

DETAILED DESCRIPTION OF THE INVENTION

In general, exogenous nucleic acids, particularly viral nucleic acids, introduced into cells induce an innate immune response, resulting in interferon (IFN) production and cell 10 death. However, it is of great interest for therapeutics, diagnostics, reagents and for biological assays to deliver a nucleic acid, e.g., a ribonucleic acid (RNA) inside a cell, either in vivo or ex vivo, such as to cause intracellular translation of the nucleic acid and production of the encoded 15 protein. Of particular importance is the delivery and function of a non-integrative nucleic acid, as nucleic acids characterized by integration into a target cell are generally imprecise in their expression levels, deleteriously transferable to progeny and neighbor cells, and suffer from the 20 substantial risk of mutation. Provided herein in part are nucleic acids encoding useful polypeptides capable of killing or reducing the growth of microorganisms (e.g., bacteria), killing or reducing the growth of tumor or cancer cells, and/or modulating a cell's function and/or activity, and 25 methods of making and using these nucleic acids and polypeptides. As described herein, these nucleic acids are capable of reducing the innate immune activity of a population of cells into which they are introduced, thus increasing the efficiency of protein production in that cell population. 30 Further, one or more additional advantageous activities and/or properties of the nucleic acids and proteins of the invention are described.

Provided herein are modified nucleic acids encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypep- 35 tide), e.g., an anti-microbial polypeptide described herein, precursors thereof, or partially or fully processed forms of these precursors. In certain embodiments, the modified nucleic acid comprises mRNA. In particular embodiments, the modified mRNA (mmRNA) is derived from cDNA. In 40 certain embodiments, the mmRNA comprises at least two nucleoside modifications. In certain embodiments, these nucleoside modifications comprise 5-methylcytosine and pseudouridine. In some embodiments, around 25%, around 50%, around 75%, or up to and including 100% of cytosine 45 and uridine nucleotides of the modified nucleic acid are modified nucleotides. In certain embodiments, the mmRNA comprises a 5' cap structure and a 3' poly-A tail. In specific embodiments, the 5' cap structure is a Cap 1 structure. In specific embodiments, the poly-A tail comprises at least 10, 50 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, or 200 nucleotides.

Further, provided herein are compositions comprising the modified nucleic acids described herein. In certain embodiments, the composition further comprises a pharmaceutically acceptable carrier. In certain embodiments, the carrier is formulated for systemic or local administration. In certain embodiments, the composition is formulated for intravenous administration. In certain embodiments, the composition is formulated for oral administration. In certain embodiments, the composition is formulated for administration in certain embodiments, the composition is formulated for administration via a dressing (e.g., wound dressing). In certain embodiments, the composition is formulated for administration via a bandage (e.g., adhesive bandage). In 65 certain embodiments, the composition is formulated for administration by inhalation. In certain embodiments, the

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composition is formulated for rectal administration. In certain embodiments, the composition is formulated for vaginal administration. In certain embodiments, the composition comprises naked modified nucleic acids. In other embodiments, the modified nucleic acid is complexed or encapsulated. For example, the modified nucleic acid may be complexed in liposomal form or may be encapsulated in a nanoparticle. In certain embodiments, the modified nucleic acid, the complex, or the nanoparticle further comprises one or more targeting moieties. These moieties can be used to target delivery in vivo to certain organs, tissues, or cells.

Provided herein are methods of treating a subject having or being suspected of having a microbial infection (e.g., a bacterial infection) and/or a disease, disorder, or condition associated with a microbial infection (e.g., a bacterial infection), the methods comprising administering to a subject in need of such treatment a composition described herein in an amount sufficient to treat the microbial infection (e.g., bacterial infection) and/or the disease, disorder, or condition associated with the microbial infection (e.g., bacterial infection). In specific embodiments, the disease, disorder, or condition is associated with one or more cellular and/or molecular changes affecting, for example, the level, activity, and/or localization of an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, precursors thereof, or a partially or fully processed form of these precursors. Cellular and/or molecular changes may affect transcription, translation, posttranslational modification, processing, folding, intraand/or extracellular trafficking, intra- and/or extracellular stability/turnover, and/or signaling of one or more molecules associated with an anti-microbial (e.g., anti-bacterial) activity. In certain embodiments, activities associated with an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, are compromised, e.g., 90%, 80%, 70%, 60%, 50%, 40%, 30%, 20%, 10%, 5% or less of wild-type activity. In certain embodiments, the method of treating a subject having or being suspected of having a microbial infection (e.g., a bacterial infection) and/or a disease, disorder, or condition associated with a microbial infection (e.g., a bacterial infection) comprises administering to the subject in need of such treatment a composition comprising a modified nucleic acid described herein in an amount sufficient to kill, reduce, or inhibit the growth of microorganisms (e.g., bacteria) and/or to treat the disease, disorder, or condition.

A major drawback of many current treatments for diseases described herein is the necessity to produce anti-microbial agents (e.g., anti-bacterial agents) as polypeptides. Polypeptides are ordinarily expressed in and isolated from mammalian or bacterial cultures. Bacterial cultures and many cancer-derived cell culture systems do not faithfully recapitulate post-translational modifications, e.g., glycosylation and amidation, and protein precursors may not be fully processed. In some instances, the lack of posttranslational modification and processing influences the activity of the final protein product, its localization and/or its target specificity. In other instances, precursors and final cleavage products can have different physiological effects. For production of recombinant proteins, the polypeptide product that is effective for a particular treatment must usually be predetermined because the proteins if administered do not undergo any additional processing. Any modification that is vital for activity must also be present on the recombinant protein because they will not be added by the host when the recombinant proteins are administered. Recombinant protein production and purification is expensive and labor intensive. Protein expression

host systems may harbor pathogens (e.g. viruses) that may contaminate the purified product. Proteins and particularly protein modifications are inherently unstable and require specific storage conditions and generally have a short shelf life. To be efficacious, recombinant proteins must be further 5 modified, particularly by pegylation to avoid rapid degradation in vivo. Still, site-specific pegylation remains difficult because it can lead to loss of activity, loss of target specificity and/or protein aggregation. Veronese et al. Bioconjugate Chem. 18:1824-1830 (2007).

The modified mRNA molecules described herein do not share these problems. In comparison to recombinant proteins, they exhibit increased stability for shipping, handling and storage, are easy to mass produce, and when translated from the modified mRNA, the polypeptide can undergo an 15 array of cell- and/or tissue-specific posttranslational processing, folding and modification.

Anti-Microbial Polypeptide

Anti-microbial polypeptides (AMPs) are small peptides of variable length, sequence and structure with broad spec- 20 trum activity against a wide range of microorganisms including bacteria, viruses, fungi, protozoa, parasites, prions, and tumor/cancer cells. See, e.g., Zaiou, J Mol Med, 2007; 85:317. It has been shown that AMPs have broadspectrum of rapid onset of killing activities, with potentially 25 low levels of induced resistance and concomitant broad anti-inflammatory effects. In some embodiments, the antimicrobial polypeptide (e.g., an anti-bacterial polypeptide) is under 10 kDa, e.g., under 8 kDa, 6 kDa, 4 kDa, 2 kDa, or 1 kDa. In some embodiments, the anti-microbial polypeptide 30 (e.g., an anti-bacterial polypeptide) consists of from about 6 to about 100 amino acids, e.g., from about 6 to about 75 amino acids, about 6 to about 50 amino acids, about 6 to about 25 amino acids, about 25 to about 100 amino acids, about 50 to about 100 amino acids, or about 75 to about 100 35 amino acids. In certain embodiments, the anti-microbial polypeptide (e.g., an anti-bacterial polypeptide) consists of from about 15 to about 45 amino acids. In some embodiments, the anti-microbial polypeptide (e.g., an anti-bacterial polypeptide) is substantially cationic. In some embodiments, 40 not limited to hBD-2, LL-37, and RNase-7. the anti-microbial polypeptide (e.g., an anti-bacterial polypeptide) is substantially amphipathic. In certain embodiments, the anti-microbial polypeptide (e.g., an anti-bacterial polypeptide) is substantially cationic and amphipathic. In some embodiments, the anti-microbial polypeptide (e.g., an 45 anti-bacterial polypeptide) is cytostatic to a Gram-positive bacterium. In some embodiments, the anti-microbial polypeptide (e.g., an anti-bacterial polypeptide) is cytotoxic to a Gram-positive bacterium. In some embodiments, the antimicrobial polypeptide (e.g., an anti-bacterial polypeptide) is 50 cytostatic and cytotoxic to a Gram-positive bacterium. In some embodiments, the anti-microbial polypeptide (e.g., an anti-bacterial polypeptide) is cytostatic to a Gram-negative bacterium. In some embodiments, the anti-microbial polypeptide (e.g., an anti-bacterial polypeptide) is cytotoxic to a 55 Gram-negative bacterium. In some embodiments, the antimicrobial polypeptide (e.g., an anti-bacterial polypeptide) is cytostatic and cytotoxic to a Gram-positive bacterium. In some embodiments, the anti-microbial polypeptide is cytostatic to a virus, fungus, protozoan, parasite, prion, or a 60 combination thereof. In some embodiments, the anti-microbial polypeptide is cytotoxic to a virus, fungus, protozoan, parasite, prion, or a combination thereof. In certain embodiments, the anti-microbial polypeptide is cytostatic and cytotoxic to a virus, fungus, protozoan, parasite, prion, or a 65 combination thereof. In some embodiments, the anti-microbial polypeptide is cytotoxic to a tumor or cancer cell (e.g.,

a human tumor or cancer cell). In some embodiments, the anti-microbial polypeptide is cytostatic to a tumor or cancer cell (e.g., a human tumor or cancer cell). In certain embodiments, the anti-microbial polypeptide is cytotoxic and cytostatic to a tumor or cancer cell (e.g., a human tumor or cancer cell). In some embodiments, the anti-microbial polypeptide (e.g., an anti-bacterial polypeptide) is a secreted polypeptide.

AMPs have been isolated and described from a wide 10 range of animals: microorganisms, invertebrates, plants, amphibians, birds, fish, and mammals (Wang et al., Nucleic Acids Res. 2009; 37 (Database issue): D933-7). For example, anti-microbial polypeptides are described in Antimicrobial Peptide Database (Wang et al., Nucleic Acids Res. 2009; 37 (Database issue):D933-7), CAMP: Collection of Anti-Microbial Peptides (Thomas et al., Nucleic Acids Res. 2010; 38 (Database issue):D774-80), U.S. Pat. Nos. 5,221,732, 5,447, 914, 5,519,115, 5,607,914, 5,714,577, 5,734,015, 5,798,336, 5,821,224, 5,849,490, 5,856,127, 5,905,187, 5,994,308, 5,998,374, 6,107,460, 6,191,254, 6,211,148, 6,300,489, 6,329,504, 6,399,370, 6,476,189, 6,478,825, 6,492,328, 6,514,701, 6,573,361, 6,573,361, 6,576,755, 6,605,698, 6,624,140, 6,638,531, 6,642,203, 6,653,280, 6,696,238, 6,727,066, 6,730,659, 6,743,598, 6,743,769, 6,747,007, 6,790,833, 6,794,490, 6,818,407, 6,835,536, 6,835,713, 6,838,435, 6,872,705, 6,875,907, 6,884,776, 6,887,847, 6,906,035, 6,911,524, 6,936,432, 7,001,924, 7,071,293, 7,078,380, 7,091,185, 7,094,759, 7,166,769, 7,244,710, 7,314,858, and U.S. Pat. No. 7,582,301, the contents of which are incorporated by reference in their entirety.

In certain embodiments, the anti-microbial polypeptide (e.g., anti-bacterial polypeptide) is selected from the group consisting of anti-microbial polypeptides (e.g., anti-bacterial polypeptides) provided in Lengthy Table 1. Shown in Lengthy Table 1, in addition to the name of the antimicrobial polypeptide (e.g., anti-bacterial polypeptide) is the definition of the polypeptide and the sequence and SEQ ID NO of the polypeptide.

Exemplary anti-microbial polypeptides also include, but

The human defensin hBD-2 is expressed throughout human epithelia. The sequence of the precursor peptide consists of 41 residues present in the mature peptide as well as a leader sequence of secreted peptide. Disruption of hBD-2 expression, as in cystic fibrosis, might be associated with recurrent infections of skin and other epithelia.

The anti-microbial peptide, LL-37 is processed from the cathelicidin precursor hCAP18. The inhibition of LL-37 expression by Shigella likely causes about 160 million people develop intestinal infections yearly, resulting in over 1 million deaths. It is a multifunctional effector molecule capable of directly killing pathogens, modulating the immune response, stimulating proliferation, angiogenesis, and cellular migration, inhibiting apoptosis, and is associated with inflammation. It may play a part in epithelial cell proliferation as a part in wound closure and that its reduction in chronic wounds impairs re-epithelialization and may contribute to their failure to heal.

RNAse-7 is a potent AMP that was identified in the skin, human kidney and urinary tract. The systemic delivery of this mRNAs will likely allow expression of natural for the body antibiotic polypeptides even in tissues which are not supposed to be under microbial attack at normal physiological stage but have that danger under disease conditions.

In some embodiments, the anti-microbial polypeptide comprises or consists of a defensin. Exemplary defensins include, but not limited to, \alpha-defensins (e.g., neutrophil

defensin 1, defensin alpha 1, neutrophil defensin 3, neutrophil defensin 4, defensin 5, defensin 6), β -defensins (e.g., beta-defensin 1, beta-defensin 2, beta-defensin 103, beta-defensin 107, beta-defensin 110, beta-defensin 136), and θ -defensins. In other embodiments, the anti-microbial polypeptide comprises or consists of a cathelicidin (e.g., hCAP 18)

The anti-microbial polypeptides described herein may block cell fusion and/or viral entry by one or more enveloped viruses (e.g., HIV, HCV). For example, the antimicrobial polypeptide can comprise or consist of a synthetic peptide corresponding to a region, e.g., a consecutive sequence of at least about 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, or 60 amino acids of the transmembrane subunit of a viral envelope protein, e.g., HIV-1 gp 120 or gp41. The 15 amino acid and nucleotide sequences of HIV-1 gp 120 or gp41 are described in, e.g., Kuiken et al., (2008). "HIV Sequence Compendium", Los Alamos National Laboratory. In some embodiments, the anti-microbial polypeptide has at least about 75%, 80%, 85%, 90%, 95%, 100% sequence 20 homology to the corresponding viral protein sequence. In certain embodiments, the anti-microbial polypeptide comprises or consists of enfuvirtide (FUZEON®): Ac-Tyr-Thr-Ser-Leu-Ile-His-Ser-Leu-Ile-Glu-Glu-Ser-Gln-Asn-Gln-Gln-Glu-Lys-Asn-Glu-Glu-Leu-Leu-Glu-Leu-Asp-Lys-Trp-Ala-Ser-Leu-Trp-Asn- Trp-Phe-NH₂ (SEQ ID NO: 178).

The anti-microbial polypeptides described herein may block viral particle assembly and formation of one or more infective enveloped viruses (e.g., HIV, HCV). For example, 30 the anti-microbial polypeptide can comprise or consist of a synthetic peptide corresponding to a region, e.g., a consecutive sequence of at least about 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, or 60 amino acids of the capsid subunit of a viral capsid protein, e.g., the HIV capsid protein. The amino acid 35 and nucleotide sequences of the HIV-1 capsid protein are described in, e.g., Kuiken et al., (2008). "HIV Sequence Compendium", Los Alamos National Laboratory. In some embodiments, the anti-microbial polypeptide has at least about 75%, 80%, 85%, 90%, 95%, 100% sequence homol- 40 ogy to the corresponding viral protein sequence. In other embodiments, the anti-microbial polypeptide comprises or consists of a synthetic peptide corresponding to a region, e.g., a consecutive sequence of at least about 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, or 60 amino acids of the binding 45 domain of a capsid binding protein. In some embodiments, the anti-microbial polypeptide has at least about 75%, 80%, 85%, 90%, 95%, 100% sequence homology to the corresponding sequence of the capsid binding protein.

The anti-microbial polypeptides described herein may 50 block protease dimerization and inhibit cleavage of viral proproteins (e.g., HIV Gag-pol processing) into functional proteins thereby preventing release of one or more enveloped viruses (e.g., HIV, HCV). For example, the antimicrobial polypeptide can comprise or consist of a synthetic 55 peptide corresponding to a region, e.g., a consecutive sequence of at least about 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, or 60 amino acids of a viral protease, e.g., the HIV-1 protease. The amino acid and nucleotide sequences of the HIV-1 protease are described in, e.g., Kuiken et al., (2008). 60 "HIV Sequence Compendium", Los Alamos National Laboratory. In some embodiments, the anti-microbial polypeptide has at least about 75%, 80%, 85%, 90%, 95%, 100% sequence homology to the corresponding viral protein sequence. In other embodiments, the anti-microbial polypeptide can comprise or consist of a synthetic peptide corresponding to a region, e.g., a consecutive sequence of at

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least about 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, or 60 amino acids of the binding domain of a protease binding protein. In some embodiments, the anti-microbial polypeptide has at least about 75%, 80%, 85%, 90%, 95%, 100% sequence homology to the corresponding sequence of the protease binding protein.

The anti-microbial polypeptides described herein can include a polypeptide corresponding to the inhibitory region of the endogenous human protein TRIM5- α or cyclophilin A (peptidylprolyl isomerase A). The sequences of human TRIM5- α and cyclophilin A are described, e.g., in Stremlau et al., *Nature*. 2004; 427(6977):848-53 and Takahashi et al., *Nature* 1989; 337 (6206), 473-475, respectively.

The anti-microbial polypeptides described herein can include an in vitro-evolved polypeptide directed against a viral pathogen, e.g., a polypeptide identified or selected by the method described in Example 7.

Modified Nucleic Acids.

This invention provides nucleic acids, including RNAs such as mRNAs that contain one or more modified nucleosides (termed "modified nucleic acids"), which have useful properties including the lack of a substantial induction of the innate immune response of a cell into which the mRNA is introduced. Because these modified nucleic acids enhance the efficiency of protein production, intracellular retention of nucleic acids, and viability of contacted cells, as well as possess reduced immunogenicity, these nucleic acids having these properties are termed "enhanced nucleic acids" herein.

The term "nucleic acid," in its broadest sense, includes any compound and/or substance that is or can be incorporated into an oligonucleotide chain. Exemplary nucleic acids for use in accordance with the present invention include, but are not limited to, one or more of DNA, RNA, hybrids thereof, RNAi-inducing agents, RNAi agents, siRNAs, shRNAs, miRNAs, antisense RNAs, ribozymes, catalytic DNA, RNAs that induce triple helix formation, aptamers, vectors, etc., described in detail herein.

Provided are modified nucleic acids containing a translatable region encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, and one, two, or more than two different nucleoside modifications. In some embodiments, the modified nucleic acid exhibits reduced degradation in a cell into which the nucleic acid is introduced, relative to a corresponding unmodified nucleic acid. For example, the degradation rate of the nucleic acid is reduced by at least 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, or 90%, compared to the degradation rate of the corresponding unmodified nucleic acid. Exemplary nucleic acids include ribonucleic acids (RNAs), deoxyribonucleic acids (DNAs), threose nucleic acids (TNAs), glycol nucleic acids (GNAs), peptide nucleic acids (PNAs), locked nucleic acids (LNAs) or a hybrid thereof. In preferred embodiments, the modified nucleic acid includes messenger RNAs (mRNAs). As described herein, the nucleic acids of the invention do not substantially induce an innate immune response of a cell into which the mRNA is introduced.

In some embodiments, modified nucleosides include pyridin-4-one ribonucleoside, 5-aza-uridine, 2-thio-5-aza-uridine, 2-thiouridine, 4-thio-pseudouridine, 2-thio-pseudouridine, 5-hydroxyuridine, 3-methyluridine, 5-carboxymethyluridine, 1-carboxymethyl-pseudouridine, 5-propynyluridine, 1-propynyl-pseudouridine, 5-taurinomethyluridine, 1-taurinomethyl-pseudouridine, 5-taurinomethyl-2-thio-uridine, 1-taurinomethyl-4-thio-uridine, 5-methyl-uridine, 1-methyl-pseudouridine, 4-thio-1-methyl-pseudouridine, 2-thio-1-methyl-pseudouridine, 1-methyl-1-deaza-pseudouridine, 1-methyl

ridine, 2-thio-1-methyl-1-deaza-pseudouridine, dihydrouridine, dihydropseudouridine, 2-thio-dihydrouridine, 2-thio-dihydropseudouridine, 2-methoxyuridine, 2-methoxy-4-thio-uridine, 4-methoxy-pseudouridine, and 4-methoxy-2-thio-pseudouridine.

In some embodiments, modified nucleosides include 5-aza-cytidine, pseudoisocytidine, 3-methyl-cytidine, N4-acetylcytidine, 5-formylcytidine, N4-methylcytidine, 5-hydroxymethylcytidine, 1-methyl-pseudoisocytidine, pyrrolo-cytidine, pyrrolo-pseudoisocytidine, 2-thio-cytidine, 10 2-thio-5-methyl-cytidine, 4-thio-pseudoisocytidine, 4-thio-1-methyl-pseudoisocytidine, 4-thio-1-methyl-1-deaza-1-methyl-1-deaza-pseudoisocytidine, pseudoisocytidine, zebularine, 5-aza-zebularine, 5-methyl-zebularine, 5-aza-2thio-zebularine, 2-thio-zebularine, 2-methoxy-cytidine, 15 2-methoxy-5-methyl-cytidine, 4-methoxy-pseudoisocytidine, and 4-methoxy-1-methyl-pseudoisocytidine.

In other embodiments, modified nucleosides include 2,6-diaminopurine, 7-deaza-adenine, 2-aminopurine, 7-deaza-8-aza-adenine, 7-deaza-2-aminopurine, 7-deaza-8- 20 aza-2-aminopurine, 7-deaza-2,6-diaminopurine, 7-deaza-8aza-2,6-diaminopurine, 1-methyladenosine, N6-methylad-N6-isopentenyladenosine, N6-(cisenosine. hydroxyisopentenyl)adenosine, 2-methylthio-N6-(cishydroxyisopentenyl) adenosine, 25 N6-glycinylcarbamoyladenosine, N6-threonylcarbamoyladenosine, 2-methylthio-N6-threonyl carbamoyladenosine, N6,N6-dimethyladenosine, 7-methyladenine, 2-methylthioadenine, and 2-methoxy-adenine.

In certain embodiments it is desirable to intracellularly 30 degrade a modified nucleic acid introduced into the cell, for example if precise timing of protein production is desired. Thus, the invention provides a modified nucleic acid containing a degradation domain, which is capable of being acted on in a directed manner within a cell.

In other embodiments, modified nucleosides include inosine, 1-methyl-inosine, wyosine, wybutosine, 7-deazaguanosine, 7-deaza-guanosine, 6-thio-7-deaza-guanosine, 6-thio-7-deaza-guanosine, 6-thio-7-deaza-guanosine, 7-methyl-guanosine, 7-methyl-guanosine, 7-methyl-guanosine, 1-methylguanosine, N2-methylguanosine, N2-methylguanosine, N2-methyl-8-oxo-guanosine, 1-methyl-6-thioguanosine, N2-methyl-6-thioguanosine, and N2,N2-dimethyl-6-thioguanosine.

Other components of nucleic acid are optional, and are beneficial in some embodiments. For example, a 5' untranslated region (UTR) and/or a 3'UTR are provided, wherein either or both may independently contain one or more different nucleoside modifications. In such embodiments, 50 nucleoside modifications may also be present in the translatable region. Also provided are nucleic acids containing a Kozak sequence.

Additionally, nucleic acids encoding anti-microbial polypeptides (e.g., anti-bacterial polypeptides), e.g., anti-microbial polypeptides described herein, and containing one or more intronic nucleotide sequences capable of being excised from the nucleic acid are provided herein.

Further, nucleic acids encoding anti-microbial polypeptides (e.g., anti-bacterial polypeptides), e.g., anti-microbial 60 polypeptides described herein, and containing an internal ribosome entry site (IRES) are provided herein. An IRES may act as the sole ribosome binding site, or may serve as one of multiple ribosome binding sites of an mRNA. An mRNA containing more than one functional ribosome binding site may encode several peptides or polypeptides that are translated independently by the ribosomes ("multicistronic

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mRNA"). When nucleic acids are provided with an IRES, further optionally provided is a second translatable region. Examples of IRES sequences that can be used according to the invention include without limitation, those from picornaviruses (e.g., FMDV), pest viruses (CFFV), polio viruses (PV), encephalomyocarditis viruses (ECMV), foot-and-mouth disease viruses (FMDV), hepatitis C viruses (HCV), classical swine fever viruses (CSFV), murine leukemia virus (MLV), simian immune deficiency viruses (SIV) or cricket paralysis viruses (CrPV).

Prevention or Reduction of Innate Cellular Immune Response Activation Using Modified Nucleic Acids.

The term "innate immune response" includes a cellular response to exogenous single stranded nucleic acids, generally of viral or bacterial origin, which involves the induction of cytokine expression and release, particularly the interferons, and cell death. Protein synthesis is also reduced during the innate cellular immune response. While it is advantageous to eliminate the innate immune response in a cell, the invention provides modified mRNAs that substantially reduce the immune response, including interferon signaling, without entirely eliminating such a response. In some embodiments, the immune response is reduced by 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95%, 99%, 99.9%, or greater than 99.9% as compared to the immune response induced by a corresponding unmodified nucleic acid. Such a reduction can be measured by expression or activity level of Type 1 interferons or the expression of interferon-regulated genes such as the toll-like receptors (e.g., TLR7 and TLR8). Reduction of innate immune response can also be measured by decreased cell death following one or more administrations of modified RNAs to a cell population; e.g., cell death is 10%, 25%, 50%, 75%, 85%, 90%, 95%, or over 95% less than the cell death 35 frequency observed with a corresponding unmodified nucleic acid. Moreover, cell death may affect fewer than 50%, 40%, 30%, 20%, 10%, 5%, 1%, 0.1%, 0.01% or fewer than 0.01% of cells contacted with the modified nucleic

The invention provides for the repeated introduction (e.g., transfection) of modified nucleic acids into a target cell population, e.g., in vitro, ex vivo, or in vivo. The step of contacting the cell population may be repeated one or more times (such as two, three, four, five or more than five times). In some embodiments, the step of contacting the cell population with the modified nucleic acids is repeated a number of times sufficient such that a predetermined efficiency of protein translation in the cell population is achieved. Given the reduced cytotoxicity of the target cell population provided by the nucleic acid modifications, such repeated transfections are achievable in a diverse array of cell types. Polypeptide Variants.

Provided are nucleic acids that encode variant polypeptides, which have a certain identity with a reference polypeptide (e.g., an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein) sequence. The term "identity" as known in the art, refers to a relationship between the sequences of two or more peptides, as determined by comparing the sequences. In the art, "identity" also means the degree of sequence relatedness between peptides, as determined by the number of matches between strings of two or more amino acid residues. "Identity" measures the percent of identical matches between the smaller of two or more sequences with gap alignments (if any) addressed by a particular mathematical model or computer program (i.e., "algorithms"). Identity of related peptides can be readily calculated by known

methods. Such methods include, but are not limited to, those described in Computational Molecular Biology, Lesk, A. M., ed., Oxford University Press, New York, 1988; Biocomputing: Informatics and Genome Projects, Smith, D. W., ed., Academic Press, New York, 1993; Computer Analysis of 5 Sequence Data, Part 1, Griffin, A. M., and Griffin, H. G., eds., Humana Press, New Jersey, 1994; Sequence Analysis in Molecular Biology, von Heinje, G., Academic Press, 1987; Sequence Analysis Primer, Gribskov, M. and Devereux, J., eds., M. Stockton Press, New York, 1991; and 10 Carillo et al., SIAM J. Applied Math. 48, 1073 (1988).

In some embodiments, the polypeptide variant has the same or a similar activity as the reference polypeptide. Alternatively, the variant has an altered activity (e.g., increased or decreased) relative to a reference polypeptide. 15 Generally, variants of a particular polynucleotide or polypeptide of the invention will have at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more sequence identity to that particular reference polynucleotide 20 or polypeptide as determined by sequence alignment programs and parameters described herein and known to those skilled in the art.

As recognized by those skilled in the art, protein fragments, functional protein domains, and homologous proteins 25 or more noncoding regions. Such modified nucleic acids are are also considered to be within the scope of this invention. For example, provided herein is any protein fragment of a reference protein (meaning a polypeptide sequence at least one amino acid residue shorter than a reference polypeptide sequence but otherwise identical) 5, 10, 15, 20, 25, 30, 35, 30 40, 45, 50, 55, 60, 70, 80, 90, 100, or greater than 100 amino acids in length In another example, any protein that includes a stretch of about 20, about 30, about 40, about 50, or about 100 amino acids which are about 40%, about 50%, about 60%, about 70%, about 80%, about 90%, about 95%, about 35 98%, or about 100% identical to any of the sequences described herein can be utilized in accordance with the invention. In certain embodiments, a protein sequence to be utilized in accordance with the invention includes 2, 3, 4, 5, 6, 7, 8, 9, 10, or more mutations as shown in any of the 40 sequences provided or referenced herein. Polynucleotide Libraries.

Also provided are polynucleotide libraries containing nucleoside modifications, wherein the polynucleotides individually contain a first nucleic acid sequence encoding a 45 polypeptide, such as an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein. Preferably, the polynucleotides are mRNA in a form suitable for direct introduction into a target cell host, which in turn synthesizes the encoded polypeptide. 50

In certain embodiments, multiple variants of a protein, each with different amino acid modification(s), are produced and tested to determine the best variant in terms of pharmacokinetics, stability, biocompatibility, and/or biological activity, or a biophysical property such as expression level. 55 Such a library may contain $10, 10^2, 10^3, 10^4, 10^5, 10^6, 10^7$, 10⁸, 10⁹, or over 10⁹ possible variants (including substitutions, deletions of one or more residues, and insertion of one or more residues).

Polypeptide-Nucleic Acid Complexes.

Proper protein translation involves the physical aggregation of a number of polypeptides and nucleic acids associated with the mRNA. Provided by the invention are complexes containing conjugates of protein and nucleic acids, containing a translatable mRNA encoding an anti-microbial 65 polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein), and having

one or more nucleoside modifications (e.g., at least two different nucleoside modifications) and one or more polypeptides bound to the mRNA. Generally, the proteins are provided in an amount effective to prevent or reduce an innate immune response of a cell into which the complex is

Targeting Moieties.

In embodiments of the invention, modified nucleic acids are provided to express a protein-binding partner or a receptor on the surface of the cell, which functions to target the cell to a specific tissue space or to interact with a specific moiety, either in vivo or in vitro. Suitable protein-binding partners include antibodies and functional fragments thereof, scaffold proteins, or peptides. Additionally, modified nucleic acids can be employed to direct the synthesis and extracellular localization of lipids, carbohydrates, or other biological moieties.

Untranslatable Modified Nucleic Acids; Vaccines.

As described herein, provided are mRNAs having sequences that are substantially not translatable. Such mRNA is effective as a vaccine when administered to a mammalian subject.

Also provided are modified nucleic acids that contain one generally not translated, but are capable of binding to and sequestering one or more translational machinery component such as a ribosomal protein or a transfer RNA (tRNA), thereby effectively reducing protein expression in the cell. The modified nucleic acid may contain a small nucleolar RNA (sno-RNA), micro RNA (miRNA), small interfering RNA (siRNA), or Piwi-interacting RNA (piRNA).

Additionally, certain modified nucleosides, or combinations thereof, when introduced into modified nucleic acids activate the innate immune response. Such activating modified nucleic acids, e.g., modified RNAs, are useful as adjuvants when combined with polypeptides (e.g., anti-microbial polypeptides) or other vaccines. In certain embodiments, the activated modified mRNAs contain a translatable region which encodes for a polypeptide (e.g., an anti-microbial polypeptide (e.g., an anti-microbial polypeptide described herein)) sequence useful as a vaccine, thus providing the ability to be a self-adjuvant.

Modified Nucleic Acid Synthesis.

Nucleic acids for use in accordance with the invention may be prepared according to any available technique including, but not limited to chemical synthesis, enzymatic synthesis, which is generally termed in vitro transcription, enzymatic or chemical cleavage of a longer precursor, etc. Methods of synthesizing RNAs are known in the art (see, e.g., Gait, M. J. (ed.) Oligonucleotide synthesis: a practical approach, Oxford (Oxfordshire), Washington, D.C.: IRL Press, 1984; and Herdewijn, P. (ed.) Oligonucleotide synthesis: methods and applications, Methods in Molecular Biology, v. 288 (Clifton, N.J.) Totowa, N.J.: Humana Press, 2005; both of which are incorporated herein by reference).

Modified nucleic acids need not be uniformly modified along the entire length of the molecule. Different nucleotide modifications and/or backbone structures may exist at various positions in the nucleic acid. One of ordinary skill in the art will appreciate that the nucleotide analogs or other modification(s) may be located at any position(s) of a nucleic acid such that the function of the nucleic acid is not substantially decreased. A modification may also be a 5' or 3' terminal modification. The nucleic acids may contain at a minimum one and at maximum 100% modified nucleotides,

or any intervening percentage, such as at least 50% modified nucleotides, at least 80% modified nucleotides, or at least 90% modified nucleotides.

Generally, the length of a modified mRNA of the present invention is greater than 30 nucleotides in length. In another 5 embodiment, the RNA molecule is greater than 35, 40, 45, 50, 60, 75, 100, 125, 150, 175, 200, 250, 300, 350, 400, 450, 500, 600, 700, 800, 900, 1000, 1100, 1200, 1300, 1400, 1500, 1800, 2000, 3000, 4000, 5000 nucleotides, or greater than 5000 nucleotides.

Uses of Modified Nucleic Acids.

Therapeutic Agents.

The modified nucleic acids described herein can be used as therapeutic agents to treat or prevent microbial infections and/or diseases, disorders, or conditions associated with 15 microbial infections. Provided herein are compositions (e.g., pharmaceutical compositions), formulations, methods, kits, dressings (e.g., wound dressings), bandages (e.g., adhesive bandages), and reagents for treatment or prevention of diseases, disorders, or conditions, e.g., diseases, disorders, 20 or conditions associated with microbial infections (e.g., bacterial infections), in humans and other animals (e.g., mammals). The active therapeutic agents of the invention include modified nucleic acids, cells containing modified nucleic acids or polypeptides translated from the modified 25 nucleic acids, polypeptides translated from modified nucleic acids, and cells contacted with cells containing modified nucleic acids or polypeptides translated from the modified nucleic acids.

Provided are methods of inducing translation of a recombinant polypeptide (e.g., an anti-microbial polypeptide described herein) in a cell population using the modified nucleic acids described herein. Such translation can be in vivo, ex vivo, in culture, or in vitro. The cell population is contacted with an effective amount of a composition con- 35 taining a nucleic acid that has at least one nucleoside modification, and a translatable region encoding the recombinant polypeptide. The population is contacted under conditions such that the nucleic acid is localized into one or more cells of the cell population and the recombinant 40 polypeptide is translated in the cell from the nucleic acid.

An effective amount of the composition is provided based, at least in part, on the target tissue, target cell type, means of administration, physical characteristics of the nucleic acid (e.g., size, and extent of modified nucleosides), and other 45 determinants. In general, an effective amount of the composition provides efficient protein production in the cell, preferably more efficient than a composition containing a corresponding unmodified nucleic acid. Increased efficiency may be demonstrated by increased cell transfection (i.e., the 50 percentage of cells transfected with the nucleic acid), increased protein translation from the nucleic acid, decreased nucleic acid degradation (as demonstrated, e.g., by increased duration of protein translation from a modified nucleic acid), or reduced innate immune response of the host 55 normal level, or from a normal level to a super-normal level.

Aspects of the disclosures are directed to methods of inducing in vivo translation of a recombinant polypeptide (e.g., an anti-microbial polypeptide described herein) in a human or animal (e.g., mammalian) subject in need thereof. 60 Therein, an effective amount of a composition containing a nucleic acid that has at least one nucleoside modification and a translatable region encoding the recombinant polypeptide (e.g., an anti-microbial polypeptide described herein) is administered to the subject using the delivery methods 65 described herein. The nucleic acid is provided in an amount and under other conditions such that the nucleic acid is

localized into a cell of the subject and the recombinant polypeptide is translated in the cell from the nucleic acid. The cell in which the nucleic acid is localized, or the tissue in which the cell is present, may be targeted with one or more than one rounds of nucleic acid administration.

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Other aspects of the disclosures relate to transplantation of cells containing modified nucleic acids to a human or animal (e.g., mammalian) subject. Administration of cells to human or animal (e.g., mammalian) subjects is known to those of ordinary skill in the art, such as local implantation (e.g., topical or subcutaneous administration), organ delivery or systemic injection (e.g., intravenous injection or inhalation), as is the formulation of cells in pharmaceutically acceptable carrier. Compositions containing modified nucleic acids are formulated for administration intramuscularly, transarterially, intraocularly, vaginally, rectally, intraperitoneally, intravenously, intranasally, subcutaneously, endoscopically, transdermally, or intrathecally. In some embodiments, the composition is formulated for extended release.

The subject to whom the therapeutic agent is administered suffers from or is at risk of developing a disease, disorder, or deleterious condition. Provided are methods of identifying, diagnosing, and classifying subjects on these bases, which may include clinical diagnosis, biomarker levels, genomewide association studies (GWAS), and other methods known in the art.

In certain embodiments, nucleic acids encoding an antimicrobial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, are administered to subjects in need of anti-microbial polypeptide (e.g., an anti-bacterial polypeptide) administration.

In certain embodiments, the administered modified nucleic acid directs production of one or more recombinant polypeptides that provide a functional activity which is substantially absent in the cell in which the recombinant polypeptide is translated. For example, the missing functional activity may be enzymatic, structural, or gene regulatory in nature. In related embodiments, the administered modified nucleic acid directs production of one or more recombinant polypeptides that increases (e.g., synergistically) a functional activity which is present but substantially deficient in the cell in which the recombinant polypeptide is translated.

In other embodiments, the administered modified nucleic acid directs production of one or more recombinant polypeptides that replace a polypeptide (or multiple polypeptides) that is substantially absent in the cell in which the recombinant polypeptide is translated. Such absence may be due to genetic mutation of the encoding gene or regulatory pathway thereof. In some embodiments, the recombinant polypeptide increases the level of an endogenous protein in the cell to a desirable level; such an increase may bring the level of the endogenous protein from a subnormal level to a

Alternatively, the recombinant polypeptide functions to antagonize the activity of an endogenous protein present in, on the surface of, or secreted from the cell. Usually, the activity of the endogenous protein is deleterious to the subject, for example, due to mutation of the endogenous protein resulting in altered activity or localization. Additionally, the recombinant polypeptide antagonizes, directly or indirectly, the activity of a biological moiety present in, on the surface of, or secreted from the cell. Examples of antagonized biological moieties include lipids (e.g., cholesterol), a lipoprotein (e.g., low density lipoprotein), a nucleic acid, a carbohydrate, a protein toxin such as shiga and

tetanus toxins, or a small molecule toxin such as botulinum, cholera, and diphtheria toxins. Additionally, the antagonized biological molecule may be an endogenous protein that exhibits an undesirable activity, such as a cytotoxic or cytostatic activity.

The recombinant proteins described herein are engineered for localization within the cell, potentially within a specific compartment such as the nucleus, or are engineered for secretion from the cell or translocation to the plasma membrane of the cell.

As described herein, a useful feature of the modified nucleic acids of the invention is the capacity to reduce the innate immune response of a cell to an exogenous nucleic acid. Provided are methods for performing the titration, reduction or elimination of the immune response in a cell or a population of cells. In some embodiments, the cell is contacted with a first composition that contains a first dose of a first exogenous nucleic acid including a translatable region and at least one nucleoside modification, and the level 20 of the innate immune response of the cell to the first exogenous nucleic acid is determined. Subsequently, the cell is contacted with a second composition, which includes a second dose of the first exogenous nucleic acid, the second dose containing a lesser amount of the first exogenous 25 nucleic acid as compared to the first dose. Alternatively, the cell is contacted with a first dose of a second exogenous nucleic acid. The second exogenous nucleic acid may contain one or more modified nucleosides, which may be the same or different from the first exogenous nucleic acid or, alternatively, the second exogenous nucleic acid may not contain modified nucleosides. The steps of contacting the cell with the first composition and/or the second composition may be repeated one or more times. Additionally, efficiency of protein production (e.g., protein translation) in the cell is optionally determined, and the cell may be re-transfected with the first and/or second composition repeatedly until a target protein production efficiency is

Topical Delivery Applied to the Skin.

The skin is a desirable target site for nucleic acid delivery. It is readily accessible, and gene expression may be restricted not only to the skin, potentially avoiding nonspecific toxicity, but also to specific layers and cell types within 45 the skin. The site of cutaneous expression of the delivered nucleic acid will depend on the route of nucleic acid delivery. Three routes are commonly considered to deliver nucleic acids to the skin: (i) topical application (e.g. for local/regional treatment); (ii) intradermal injection (e.g. for 50 local/regional treatment); and (iii) systemic delivery (e.g., for treatment of dermatologic diseases that affect both cutaneous and extracutaneous regions). Nucleic acids can be delivered to the skin by several different approaches. Most have been shown to work for DNA, such as, topical appli- 55 cation of non-cationic liposome—DNA complex, cationic liposome—DNA complex, particle-mediated (gene gun), puncture-mediated gene transfections, and viral delivery approaches. After gene delivery, gene products have been detected in a number of skin cell types, including but not 60 limited to, basal keratinocytes, sebaceous gland cells, dermal fibroblasts and dermal macrophages.

In certain embodiments, dressing compositions comprising a modified nucleic acid encoding for an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an 65 anti-microbial polypeptide described herein, precursor or a partially or fully processed form are provided herein.

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In certain embodiments, the composition described herein is formulated for administration via a bandage (e.g., adhesive bandage).

The modified nucleic acids encoding for an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, precursor or a partially or fully processed form described herein may be intermixed with the dressing compositions or may be applied separately, e.g. by soaking or spraying. Targeting Moieties.

In embodiments of the disclosure, modified nucleic acids are provided to express a protein-binding partner or a receptor on the surface of the cell, which functions to target the cell to a specific tissue space or to interact with a specific moiety, either in vivo or in vitro. Suitable protein-binding partners include antibodies and functional fragments thereof, scaffold proteins, or peptides. Additionally, modified nucleic acids can be employed to direct the synthesis and extracellular localization of lipids, carbohydrates, or other biological moieties.

Methods of Treating Diseases and Conditions.

Provided are methods for treating or preventing a microbial infection (e.g., a bacterial infection) and/or a disease, disorder, or condition associated with a microbial infection (e.g., a bacterial infection), and/or a symptom thereof, by providing an anti-microbial (e.g., anti-bacterial) activity. Because of the rapid initiation of protein production following introduction of modified mRNAs, as compared to viral DNA vectors, the compounds of the present invention are particularly advantageous in treating acute or chronic diseases such as microbial infections and sepsis. Moreover, the lack of transcriptional regulation of the modified mRNAs of the invention is advantageous in that accurate titration of protein production is achievable. In some embodiments, modified mRNAs and their encoded polypeptides in accordance with the present invention may be used for therapeutic purposes.

In some embodiments, modified mRNAs and their encoded polypeptides in accordance with the present dis40 closure may be used for treatment of microbial infections and/or any of a variety of diseases, disorders, and/or conditions associated with microbial infections. Microbial infections can include, but not limited to, bacterial infections, viral infections, fungal infections, and protozoan 45 infections.

In one embodiment, modified mRNAs and their encoded polypeptides in accordance with the present disclosure may be useful in the treatment of inflammatory disorders coincident with or resulting from infection.

Exemplary diseases, disorders, or conditions associated with bacterial infections include, but not limited to one or more of the following: abscesses, actinomycosis, acute prostatitis, aeromonas hydrophila, annual ryegrass toxicity, anthrax, bacillary peliosis, bacteremia, bacterial gastroenteritis, bacterial meningitis, bacterial pneumonia, bacterial vaginosis, bacterium-related cutaneous conditions, bartonellosis, BCG-oma, botryomycosis, botulism, Brazilian purpuric fever, Brodie abscess, brucellosis, Buruli ulcer, campylobacteriosis, caries, Carrion's disease, cat scratch disease, cellulitis, chlamydia infection, cholera, chronic bacterial prostatitis, chronic recurrent multifocal osteomyelitis, clostridial necrotizing enteritis, combined periodontic-endodontic lesions, contagious bovine pleuropneumonia, diphtheria, diphtheritic stomatitis, ehrlichiosis, erysipelas, piglottitis, erysipelas, Fitz-Hugh-Curtis syndrome, fleaborne spotted fever, foot rot (infectious pododermatitis), Garre's sclerosing osteomyelitis, Gonorrhea, Granuloma

inguinale, human granulocytic anaplasmosis, human monocytotropic ehrlichiosis, hundred days' cough, impetigo, late congenital syphilitic oculopathy, legionellosis, Lemierre's syndrome, leprosy (Hansen's Disease), leptospirosis, listeriosis, Lyme disease, lymphadenitis, melioidosis, menin- 5 gococcal disease, meningococcal septicaemia, methicillinresistant Staphylococcus aureus (MRSA) infection, mycobacterium avium-intracellulare (MAI), mycoplasma pneumonia, necrotizing fasciitis, nocardiosis, noma (cancrum oris or gangrenous stomatitis), omphalitis, orbital 10 cellulitis, osteomyelitis, overwhelming post-splenectomy infection (OPSI), ovine brucellosis, pasteurellosis, periorbital cellulitis, pertussis (whooping cough), plague, pneumococcal pneumonia, Pott disease, proctitis, pseudomonas infection, psittacosis, pyaemia, pyomyositis, Q fever, relapsing fever (typhinia), rheumatic fever, Rocky Mountain spotted fever (RMSF), rickettsiosis, salmonellosis, scarlet fever, sepsis, serratia infection, shigellosis, southern tick-associated rash illness, staphylococcal scalded skin syndrome, streptococcal pharyngitis, swimming pool granuloma, swine 20 brucellosis, syphilis, syphilitic aortitis, tetanus, toxic shock syndrome (TSS), trachoma, trench fever, tropical ulcer, tuberculosis, tularemia, typhoid fever, typhus, urogenital tuberculosis, urinary tract infections, vancomycin-resistant Staphylococcus aureus infection, Waterhouse-Friderichsen 25 syndrome, pseudotuberculosis (Yersinia) disease, and yersiniosis. Other diseases, disorders, and/or conditions associated with bacterial infections can include, for example, Alzheimer's disease, anorexia nervosa, asthma, atherosclerosis, attention deficit hyperactivity disorder, autism, auto- 30 immune diseases, bipolar disorder, cancer (e.g., colorectal cancer, gallbladder cancer, lung cancer, pancreatic cancer, and stomach cancer), chronic fatigue syndrome, chronic obstructive pulmonary disease, Crohn's disease, coronary heart disease, dementia, depression, Guillain-Barré syn- 35 drome, metabolic syndrome, multiple sclerosis, myocardial infarction, obesity, obsessive-compulsive disorder, panic disorder, psoriasis, rheumatoid arthritis, sarcoidosis, schizophrenia, stroke, thromboangiitis obliterans (Buerger's disease), and Tourette syndrome.

The bacterium described herein can be a Gram-positive bacterium or a Gram-negative bacterium. Exemplary bacterial pathogens include, but not limited to, Acinetobacter baumannii, Bacillus anthracis, Bacillus subtilis, Bordetella pertussis, Borrelia burgdorferi, Brucella abortus, Brucella 45 canis, Brucella melitensis, Brucella suis, Campylobacter ieiuni, Chlamydia pneumoniae, Chlamydia trachomatis, psittaci, Chlamydophila Clostridium botulinum. Clostridium difficile, Clostridium perfringens, Clostridium tetani, coagulase Negative Staphylococcus, Corynebacte- 50 rium diphtheria, Enterococcus faecalis, Enterococcus faecium, Escherichia coli, enterotoxigenic Escherichia coli (ETEC), enteropathogenic E. coli, E. coli O157:H7, Enterobacter sp., Francisella tularensis, Haemophilus influenzae, Helicobacter pylori, Klebsiella pneumoniae, Legionella 55 pneumophila, Leptospira interrogans, Listeria monocytogenes, Moraxella catarralis, Mycobacterium leprae, Mycobacterium tuberculosis, Mycoplasma pneumoniae, Neisseria gonorrhoeae, Neisseria meningitides, Preteus mirabilis, Proteus sps., Pseudomonas aeruginosa, Rickettsia rickettsii, 60 Salmonella typhi, Salmonella typhimurium, Serratia marcesens, Shigella flexneri, Shigella sonnei, Staphylococcus aureus, Staphylococcus epidermidis, Staphylococcus saprophyticus, Streptococcus agalactiae, Streptococcus mutans, Streptococcus pneumoniae, Streptococcus pyogenes, 65 Treponema pallidum, Vibrio cholerae, and Yersinia pestis. Bacterial pathogens may also include bacteria that cause

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resistant bacterial infections, for example, clindamycinresistant Clostridium difficile, fluoroquinolon-resistant Clostridium difficile, methicillin-resistant Staphylococcus aureus (MRSA), multidrug-resistant Enterococcus faecalis, multidrug-resistant Enterococcus faecium, multidrug-resistance Pseudomonas aeruginosa, multidrug-resistant Acinetobacter baumannii, and vancomycin-resistant Staphylococcus aureus (VRSA).

Exemplary diseases, disorders, or conditions associated with viral infections include, but not limited to, acute febrile pharyngitis, pharyngoconjunctival fever, epidemic keratoconjunctivitis, infantile gastroenteritis, Coxsackie infections, infectious mononucleosis, Burkitt lymphoma, acute hepatitis, chronic hepatitis, hepatic cirrhosis, hepatocellular carcinoma, primary HSV-1 infection (e.g., gingivostomatitis in children, tonsillitis and pharyngitis in adults, keratoconjunctivitis), latent HSV-1 infection (e.g., herpes labialis and cold sores), primary HSV-2 infection, latent HSV-2 infection, aseptic meningitis, infectious mononucleosis, Cytomegalic inclusion disease, Kaposi sarcoma, multicentric Castleman disease, primary effusion lymphoma, AIDS, influenza, Reye syndrome, measles, postinfectious encephalomyelitis, Mumps, hyperplastic epithelial lesions (e.g., common, flat, plantar and anogenital warts, laryngeal papillomas, epidermodysplasia verruciformis), cervical carcinoma, squamous cell carcinomas, croup, pneumonia, bronchiolitis, common cold, Poliomyelitis, Rabies, bronchiolitis, pneumonia, influenza-like syndrome, severe bronchiolitis with pneumonia, German measles, congenital rubella, Varicella, and herpes zoster.

Exemplary viral pathogens include, but not limited to, adenovirus, coxsackievirus, dengue virus, encephalitis virus, Epstein-Barr virus, hepatitis A virus, hepatitis B virus, hepatitis C virus, herpes simplex virus type 1, herpes simplex virus type 2, cytomegalovirus, human herpesvirus type 8, human immunodeficiency virus, influenza virus, measles virus, mumps virus, human papillomavirus, parainfluenza virus, poliovirus, rabies virus, respiratory syncytial virus, rubella virus, varicella-zoster virus, West Nile virus, and yellow fever virus. Viral pathogens may also include viruses that cause resistant viral infections.

Exemplary diseases, disorders, or conditions associated with fungal infections include, but not limited to, aspergilloses, blastomycosis, candidasis, coccidioidomycosis, cryptococcosis, histoplasmosis, mycetomas, paracoccidioidomycosis, and tinea pedis. Furthermore, persons with immunodeficiencies are particularly susceptible to disease by fungal genera such as *Aspergillus, Candida, Cryptococcus, Histoplasma*, and *Pneumocystis*. Other fungi can attack eyes, nails, hair, and especially skin, the so-called dermatophytic fungi and keratinophilic fungi, and cause a variety of conditions, of which ringworms such as athlete's foot are common. Fungal spores are also a major cause of allergies, and a wide range of fungi from different taxonomic groups can evoke allergic reactions in some people.

Exemplary fungal pathogens include, but not limited to, Ascomycota (e.g., Fusarium oxysporum, Pneumocystis jirovecii, Aspergillus spp., Coccidioides immitis/posadasii, Candida albicans), Basidiomycota (e.g., Filobasidiella neoformans, Trichosporon), Microsporidia (e.g., Encephalitozoon cuniculi, Enterocytozoon bieneusi), and Mucoromycotina (e.g., Mucor circinelloides, Rhizopus oryzae, Lichtheimia corymbifera).

Exemplary diseases, disorders, or conditions associated with protozoal infections include, but not limited to, amoebiasis, giardiasis, trichomoniasis, African Sleeping Sickness,

American Sleeping Sickness, leishmaniasis (Kala-Azar), balantidiasis, toxoplasmosis, malaria, *acanthamoeba keratitis*, and babesiosis.

Exemplary protozoal pathogens include, but not limited to, Entamoeba histolytica, Giardia lambila, Trichomonas vaginalis, Trypanosoma brucei, T. cruzi, Leishmania donovani, Balantidium coli, Toxoplasma gondii, Plasmodium spp., and Babesia microti.

Exemplary diseases, disorders, or conditions associated with parasitic infections include, but not limited to, *acan-thamoeba keratitis*, amoebiasis, ascariasis, babesiosis, balantidiasis, baylisascariasis, chagas disease, clonorchiasis, *cochliomyia*, cryptosporidiosis, diphyllobothriasis, dracunculiasis, echinococcosis, elephantiasis, enterobiasis, fascioliasis, fasciolopsiasis, filariasis, giardiasis, gnathostomiasis, hymenolepiasis, isosporiasis, katayama fever, leishmaniasis, lyme disease, malaria, metagonimiasis, myiasis, onchocerciasis, pediculosis, scabies, schistosomiasis, sleeping sickness, strongyloidiasis, taeniasis, toxocariasis, toxoplasmosis, trichinosis, and trichuriasis.

Exemplary parasitic pathogens include, but not limited to, Acanthamoeba, Anisakis, Ascaris lumbricoides, botfly, Balantidium coli, bedbug, Cestoda, chiggers, Cochliomyia hominivorax, Entamoeba histolytica, Fasciola hepatica, 25 Giardia lamblia, hookworm, Leishmania, Linguatula serrata, liver fluke, Loa loa, Paragonimus, pinworm, Plasmodium falciparum, Schistosoma, Strongyloides stercoralis, mite, tapeworm, Toxoplasma gondii, Trypanosoma, whipworm, Wuchereria bancrofti.

Exemplary diseases, disorders, or conditions associated with prion infections include, but not limited to Creutzfeldt-Jakob disease (CJD), iatrogenic Creutzfeldt-Jakob disease (iCJD), variant Creutzfeldt-Jakob disease (vCJD), familial Creutzfeldt-Jakob disease (fCJD), sporadic Creutzfeldt-Jakob disease (sCJD), Gerstmann-Sträussler-Scheinker syndrome (GSS), fatal familial insomnia (FFI), Kuru, Scrapie, bovine spongiform encephalopathy (BSE), mad cow disease, transmissible mink encephalopathy (TME), chronic 40 wasting disease (CWD), feline spongiform encephalopathy (FSE), exotic ungulate encephalopathy (EUE), and spongiform encephalopathy.

Provided herein, are methods to prevent infection and/or sepsis in a subject at risk of developing infection and/or 45 sepsis, the method comprising administering to a subject in need of such prevention a composition comprising a modified nucleic acid precursor encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, or a partially or 50 fully processed form thereof in an amount sufficient to prevent infection and/or sepsis. In certain embodiments, the subject at risk of developing infection and/or sepsis is a cancer patient. In certain embodiments, the cancer patient has undergone a conditioning regimen. In some embodiments, the conditioning regiment comprises chemotherapy, radiation therapy, or both.

Further provided herein, are methods to treat infection and/or sepsis in a subject, the method comprising administering to a subject in need of such treatment a composition 60 comprising a modified nucleic acid precursor encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein, or a partially or fully processed form thereof in an amount sufficient to treat an infection and/or sepsis. In certain 65 embodiments, the subject in need of treatment is a cancer patient. In certain embodiments, the cancer patient has

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undergone a conditioning regimen. In some embodiments, the conditioning regiment comprises chemotherapy, radiation therapy, or both.

In one embodiment, the modified mRNAs of the present invention may be administered in conjunction with one or more antibiotics. These include, but are not limited to Aknilox, Ambisome, Amoxycillin, Ampicillin, Augmentin, Avelox, Azithromycin, Bactroban, Betadine, Betnovate, Blephamide, Cefaclor, Cefadroxil, Cefdinir, Cefepime, Cefix, Cefixime, Cefoxitin, Cefpodoxime, Cefprozil, Cefuroxime, Cefzil, Cephalexin, Cephazolin, Ceptaz, Chloramphenicol, Chlorhexidine, Chloromycetin, Chlorsig, Cipro-Clarithromycin, Clindagel, Clindamycin, Clindatech, Cloxacillin, Colistin, Co-trimoxazole, Demeclocycline, Diclocil, Dicloxacillin, Doxycycline, Duricef, Erythromycin, Flamazine, Floxin, Framycetin, Fucidin, Furadantin, Fusidic, Gatifloxacin, Gemifloxacin, Gemifloxacin, Ilosone, Iodine, Levaquin, Levofloxacin, Lomefloxacin, Maxaquin, Mefoxin, Meronem, Minocycline, Moxifloxacin, Myambutol, Mycostatin, Neosporin, Netromycin, Nitrofurantoin, Norfloxacin, Norilet, Ofloxacin, Omnicef, Ospamox, Oxytetracycline, Paraxin, Penicillin, Pneumovax, Polyfax, Povidone, Rifadin, Rifampin, Rifaximin, Rifinah, Rimactane, Rocephin, Roxithromycin, Seromycin, Soframycin, Sparfloxacin, Staphlex, Targocid, Tetracycline, Tetradox, Tetralysal, tobramycin, Tobramycin, Trecator, Tygacil, Vancocin, Velosef, Vibramycin, Xifaxan, Zagam, Zitrotek, Zoderm, Zymar, and Zyvox.

In certain embodiments, the subject exhibits acute or chronic microbial infections (e.g., bacterial infections). In certain embodiments, the subject has received or is receiving a therapy. In certain embodiments, the therapy is radiotherapy, chemotherapy, steroids, ultraviolet radiation, or a combination thereof. In certain embodiments, the patient suffers from a microvascular disorder. In some embodiments, the microvascular disorder is diabetes. In certain embodiments, the patient has a wound. In some embodiments, the wound is an ulcer. In a specific embodiment, the wound is a diabetic foot ulcer. In certain embodiments, the subject has one or more burn wounds. In certain embodiments, the administration is local or systemic. In certain embodiments, the administration is subcutaneous. In certain embodiments, the administration is intravenous. In certain embodiments, the administration is oral. In certain embodiments, the administration is topical. In certain embodiments, the administration is by inhalation. In certain embodiments, the administration is rectal. In certain embodiments, the administration is vaginal.

Combination Therapy

Provided are methods for treating or preventing a microbial infection (e.g., a bacterial infection) and/or a disease, disorder, or condition associated with a microbial infection (e.g., a bacterial infection), or a symptom thereof, in a subject, by administering a modified nucleic acid encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein in combination with an anti-microbial agent (e.g., an anti-bacterial agent), e.g., an anti-microbial polypeptide or a small molecule anti-microbial compound described herein. The anti-microbial agents include, but not limited to, anti-bacterial agents, anti-viral agents, anti-fungal agents, anti-protozoal agents, anti-parasitic agents, and anti-prion agents.

The agents can be administered simultaneously, for example in a combined unit dose (e.g., providing simultaneous delivery of both agents). Alternatively, the agents can be administered at a specified time interval, for example, an interval of minutes, hours, days or weeks. Generally, the

agents are concurrently bioavailable, e.g., detectable, in the subject. In some embodiments, the agents are administered essentially simultaneously, for example two unit dosages administered at the same time, or a combined unit dosage of the two agents. In other embodiments, the agents are delivered in separate unit dosages. The agents can be administered in any order, or as one or more preparations that includes two or more agents. In a preferred embodiment, at least one administration of one of the agents, e.g., the first agent, is made within minutes, one, two, three, or four hours, or even within one or two days of the other agent, e.g., the second agent. In some embodiments, combinations can achieve synergistic results, e.g., greater than additive results, e.g., at least 25, 50, 75, 100, 200, 300, 400, or 500% greater than additive results.

Exemplary anti-bacterial agents include, but not limited to, aminoglycosides (e.g., amikacin (AMIKIN®), gentamicin (GARAMYCIN®), kanamycin (KANTREX®), neomycin (MYCIFRADIN®), netilmicin (NETROMYCIN®), tobramycin (NEBCIN®), Paromomycin (HUMATIN®), 20 ansamycins (e.g., geldanamycin, herbimycin), carbacephem (e.g., loracarbef (LORABID®), Carbapenems (e.g., ertapenem (INVANZ®), doripenem (DORIBAX®), imipenem/cilastatin (PRIMAXIN®), meropenem (MER-REM®), cephalosporins (first generation) (e.g., cefadroxil 25 (DURICEF®), cefazolin (ANCEF®), cefalotin or cefalothin (KEFLIN®), cefalexin (KEFLEX®), cephalosporins (second generation) (e.g., cefaclor (CECLOR®), cefamandole (MANDOL®), cefoxitin (MEFOXIN®), cefprozil (CEF-ZIL®), cefuroxime (CEFTIN®, ZINNAT®), cepha- 30 losporins (third generation) (e.g., cefixime (SUPRAX®), cefdinir (OMNICEF®, CEFDIEL®), cefditoren (SPEC-TRACEF®), cefoperazone (CEFOBID®), cefotaxime (CLAFORAN®), cefpodoxime (VANTIN®), ceftazidime (FORTAZ®), ceftibuten (CEDAX®), ceftizoxime 35 (CEFIZOX®), ceftriaxone (ROCEPHIN®), cephalosporins (fourth generation) (e.g., cefepime (MAXIPIME®), cephalosporins (fifth generation) (e.g., ceftobiprole (ZEFT-ERA®), glycopeptides (e.g., teicoplanin (TARGOCID®), vancomycin (VANCOCIN®), telavancin (VIBATIV®), lin- 40 cosamides (e.g., clindamycin (CLEOCIN®), lincomycin (LINCOCIN®), lipopeptide (e.g., daptomycin (CUBI-CIN®), macrolides (e.g., azithromycin (ZITHROMAX®, SUMAMED®, ZITROCIN®), clarithromycin (BIAXIN®), dirithromycin (DYNABAC®), erythromycin (ERYTHO- 45 CIN®, ERYTHROPED®), roxithromycin, troleandomycin (TAO®), telithromycin (KETEK®), spectinomycin (TRO-BICIN®), monobactams (e.g., aztreonam (AZACTAM®), nitrofurans (e.g., furazolidone (FUROXONE®), nitrofurantoin (MACRODANTIN®, MACROBID®), penicillins 50 (e.g., amoxicillin (NOVAMOX®, AMOXIL®), ampicillin (PRINCIPEN®), azlocillin, carbenicillin (GEOCILLIN®), cloxacillin (TEGOPEN®), dicloxacillin (DYNAPEN®), flucloxacillin (FLOXAPEN®), mezlocillin (MEZLIN®), methicillin (STAPHCILLIN®), nafcillin (UNIPEN®), oxa- 55 cillin (PROSTAPHLIN®), penicillin G (PENTIDS®), penicillin V (PEN-VEE-K®), piperacillin (PIPRACIL®), temocillin (NEGABAN®), ticarcillin (TICAR®), penicillin combinations (e.g., amoxicillin/clavulanate (AUGMEN-TIN®), ampicillin/sulbactam (UNASYN®), piperacillin/ 60 tazobactam (ZOSYN®), ticarcillin/clavulanate (TIMEN-TIN®), polypeptides (e.g., bacitracin, colistin (COLY-MYCIN-S®), polymyxin B, quinolones (e.g., ciprofloxacin (CIPRO®, CIPROXIN®, CIPROBAY®), enoxacin (PEN-ETREX®), gatifloxacin (TEQUIN®), levofloxacin (LEVA- 65 QUIN®), lomefloxacin (MAXAQUIN®), moxifloxacin (AVELOX®), nalidixic acid (NEGGRAM®), norfloxacin

(NOROXIN®), ofloxacin (FLOXIN®, OCUFLOX®), trovafloxacin (TROVAN®), grepafloxacin (RAXAR®), sparfloxacin (ZAGAM®), temafloxacin (OMNIFLOX®), sulfonamides (e.g., mafenide (SULFAMYLON®), sulfonamidochrysoidine (PRONTOSIL®), sulfacetamide (SULA-MYD®, BLEPH-10®), sulfadiazine (MICRO-SULFON®), silver sulfadiazine (SILVADENE®), sulfamethizole (THIO-SULFIL FORTE®), sulfamethoxazole (GANTANOL®), sulfanilimide, sulfasalazine (AZULFIDINE®), sulfisoxazole (GANTRISIN®), trimethoprim (PROLOPRIM®), TRIMPEX®), trimethoprim-sulfamethoxazole (co-trimoxazole) (TMP-SMX) (BACTRIM®, SEPTRA®), tetracyclines (e.g., demeclocycline (DECLOMYCIN®), doxycycline (VIBRAMYCIN®), minocycline (MINOCIN®), oxytetracycline (TERRAMYCIN®), tetracycline (SUMY-CIN®, ACHROMYCIN® V, STECLIN®), drugs against mycobacteria (e.g., clofazimine (LAMPRENE®), dapsone (AVLOSULFON®), capreomycin (CAPASTAT®), cycloserine (SEROMYCIN®), ethambutol (MYAMBUTOL®), ethionamide (TRECATOR®), isoniazid (I.N.H.®), pyrazi-(ALDINAMIDE®), (RIFADIN®, namide rifampin RIMACTANE®), rifabutin (MYCOBUTIN®), rifapentine (PRIFTIN®), streptomycin), and others (e.g., arsphenamine (SALVARSAN®), chloramphenicol (CHLOROMYCE-TIN®), fosfomycin (MONUROL®), fusidic acid (FUCI-DIN®), linezolid (ZYVOX®), metronidazole (FLAGYL®), mupirocin (BACTROBAN®), platensimycin, quinupristin/ dalfopristin (SYNERCID®), rifaximin (XIFAXAN®), thiamphenicol, tigecycline (TIGACYL®), tinidazole (TIN-DAMAX®, FASIGYN®).

Exemplary anti-viral agents include, but not limited to, abacavir (ZIAGEN®), abacavir/lamivudine/zidovudine (Trizivir®), aciclovir or acyclovir (CYCLOVIR®, HER-PEX®, ACIVIR®, ACIVIRAX®, ZOVIRAX®, ZOVIR®), adefovir (Preveon®, Hepsera®), amantadine (SYMME-TREL®), amprenavir (AGENERASE®), ampligen, arbidol, atazanavir (REYATAZ®), boceprevir, cidofovir, darunavir (PREZISTA®), delavirdine (RESCRIPTOR®), didanosine (VIDEX®), docosanol (ABREVA®), edoxudine, efavirenz (SUSTIVA®, STOCRIN®), emtricitabine (EMTRIVA®), emtricitabine/tenofovir/efavirenz (ATRIPLA®), enfuvirtide (FUZEON®), entecavir (BARACLUDE®, ENTAVIR®), famciclovir (FAMVIR®), fomivirsen (VITRAVENE®), (LEXIVA®, TELZIR®), fosamprenavir foscarnet (FOSCAVIR®), fosfonet, ganciclovir (CYTOVENE®, CYMEVENE®, VITRASERT®), GS 9137 (ELVITEGRA-(ALDARA®, imiquimod ZYCLARA®. BESELNA®), indinavir (CRIXIVAN®), inosine, inosine pranobex (IMUNOVIR®), interferon type I, interferon type II, interferon type III, kutapressin (NEXAVIR®), lamivudine (ZEFFIX®, HEPTOVIR®, EPIVIR®), lamivudine/ zidovudine (COMBIVIR®), lopinavir, loviride, maraviroc (SELZENTRY®, CELSENTRI®), methisazone, MK-2048, moroxydine, nelfinavir (VIRACEPT®), nevirapine (VIRA-MUNE®), oseltamivir (TAMIFLU®), peginterferon alfa-2a (PEGASYS®), penciclovir (DENAVIR®), peramivir, pleconaril, podophyllotoxin (CONDYLOX®), raltegravir (ISENTRESS®), ribavirin (COPEGUs®, REBETOL®, RIBASPHERE®, VILONA® AND VIRAZOLE®), rimantadine (FLUMADINE®), ritonavir (NORVIR®), pyramidine, saquinavir (INVIRASE®, FORTOVASE®), stavudine, tea tree oil (melaleuca oil), tenofovir (VIREAD®), tenofovir/emtricitabine (TRUVADA®), tipranavir (APTI-VUS®), trifluridine (VIROPTIC®), tromantadine (VIRU-MERZ®), valaciclovir (VALTREX®), valganciclovir (VALCYTE®), vicriviroc, vidarabine, viramidine, zalcit-

abine, zanamivir (RELENZA®), and zidovudine (azidothymidine (AZT), RETROVIR®, RETROVIS®).

Exemplary anti-fungal agents include, but not limited to, polyene antifungals (e.g., natamycin, rimocidin, filipin, nystatin, amphotericin B, candicin, hamycin), imidazole anti- ⁵ fungals (e.g., miconazole (MICATIN®, DAKTARIN®), ketoconazole (NIZORAL®, FUNGORAL®, ZOLE®), clotrimazole (LOTRIMIN®, LOTRIMIN® AF, CANESTEN®), econazole, omoconazole, bifonazole, butoconazole, fenticonazole, isoconazole, oxiconazole, sertaconazole (ERTACZO®), sulconazole, tioconazole), triazole antifungals (e.g., albaconazole fluconazole, itraconazole, isavuconazole, ravuconazole, posaconazole, voriconazole, terconazole), thiazole antifungals (e.g., abafungin), allylamines (e.g., terbinafine (LAMISIL®), naftifine (NAFTIN®), butenafine (LOTRIMIN® Ultra)), echinocandins (e.g., anidulafungin, caspofungin, micafungin), and others (e.g., polygodial, benzoic acid, ciclopirox, tolnaftate (TINAC-TIN®, DESENEX®, AFTATE®), undecylenic acid, flucy- 20 tosine or 5-fluorocytosine, griseofulvin, haloprogin, sodium bicarbonate, allicin).

Exemplary anti-protozoal agents include, but not limited to, effornithine, furazolidone (FUROXONE®, DEPENDAL-M®), melarsoprol, metronidazole (FLAGYL®), 25 ornidazole, paromomycin sulfate (HUMATIN®), pentamidine, pyrimethamine (DARAPRIM®), and tinidazole (TINDAMAX®, FASIGYN®).

Exemplary anti-parasitic agents include, but not limited to, antinematodes (e.g., mebendazole, pyrantel pamoate, 30 thiabendazole, diethylcarbamazine, ivermectin), anticestodes (e.g., niclosamide, praziquantel, albendazole), antitrematodes (e.g., praziquantel), antiamoebics (e.g., rifampin, amphotericin B), and antiprotozoals (e.g., melarsoprol, eflornithine, metronidazole, tinidazole).

Exemplary anti-prion agents include, but not limited to, flupirtine, pentosan polysuphate, quinacrine, and tetracyclic compounds.

Targeting of Pathogenic Organisms; Purification of Biological Materials.

Provided herein are methods for targeting pathogenic microorganisms, such as bacteria, yeast, protozoa, parasites, prions and the like, using modified mRNAs that encode cytostatic or cytotoxic polypeptides, e.g., anti-microbial polypeptides described herein. Preferably the mRNA introduced into the target pathogenic organism contains modified nucleosides or other nucleic acid sequence modifications that the mRNA is translated exclusively, or preferentially, in the target pathogenic organism, to reduce possible off-target effects of the therapeutic. Such methods are useful for 50 removing pathogenic organisms from biological material, including blood, semen, eggs, and transplant materials including embryos, tissues, and organs.

Targeting of Diseased Cells.

Provided herein are methods for targeting pathogenic or 55 diseased cells, particularly cells that are infected with one or more microorganisms (e.g., bacteria) or cancer cells, using modified mRNAs that encode cytostatic and/or cytotoxic polypeptides, e.g., anti-microbial polypeptides described herein. Preferably the mRNA introduced into the target 60 pathogenic cell contains modified nucleosides or other nucleic acid sequence modifications that the mRNA is translated exclusively, or preferentially, in the target pathogenic cell, to reduce possible off-target effects of the therapeutic. Alternatively, the invention provides targeting moieties that are capable of targeting the modified mRNAs to preferentially bind to and enter the target pathogenic cell.

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Methods of Protein Production.

The methods provided herein are useful for enhancing protein (e.g., an anti-microbial polypeptide described herein) product yield in a cell culture process. In a cell culture containing a plurality of host cells, introduction of the modified mRNAs described herein results in increased protein production efficiency relative to a corresponding unmodified nucleic acid. Such increased protein production efficiency can be demonstrated, e.g., by showing increased cell transfection, increased protein translation from the nucleic acid, decreased nucleic acid degradation, and/or reduced innate immune response of the host cell. Protein production can be measured by ELISA, and protein activity can be measured by various functional assays known in the art. The protein production may be generated in a continuous or a fed-batch mammalian process.

Additionally, it is useful to optimize the expression of a specific polypeptide (e.g., an anti-microbial described herein) in a cell line or collection of cell lines of potential interest, particularly an engineered protein such as a protein variant of a reference protein having a known activity. In one embodiment, provided is a method of optimizing expression of an engineered protein in a target cell, by providing a plurality of target cell types, and independently contacting with each of the plurality of target cell types a modified mRNA encoding an engineered polypeptide. Additionally, culture conditions may be altered to increase protein production efficiency. Subsequently, the presence and/or level of the engineered polypeptide in the plurality of target cell types is detected and/or quantitated, allowing for the optimization of an engineered polypeptide's expression by selection of an efficient target cell and cell culture conditions relating thereto. Such methods are particularly useful when the engineered polypeptide contains one or more posttranslational modifications or has substantial tertiary structure, situations which often complicate efficient protein production.

Modulation of Biological Pathways.

The rapid translation of modified mRNAs introduced into cells provides a desirable mechanism of modulating target biological pathways, e.g., biological pathways associated with microbial infections (e.g., bacterial infections) and/or diseases, disorders or conditions associated with microbial infections (e.g., bacterial infections). Such modulation includes antagonism or agonism of a given pathway. In one embodiment, a method is provided for antagonizing a biological pathway in a cell by contacting the cell with an effective amount of a composition comprising a modified nucleic acid encoding a recombinant polypeptide, under conditions such that the nucleic acid is localized into the cell and the recombinant polypeptide is capable of being translated in the cell from the nucleic acid, wherein the recombinant polypeptide inhibits the activity of a polypeptide functional in the biological pathway.

Alternatively, provided are methods of agonizing a biological pathway in a cell by contacting the cell with an effective amount of a modified nucleic acid encoding a recombinant polypeptide under conditions such that the nucleic acid is localized into the cell and the recombinant polypeptide is capable of being translated in the cell from the nucleic acid, and the recombinant polypeptide induces the activity of a polypeptide functional in the biological pathway. Exemplary agonized biological pathways include pathways that modulate anti-bacterial activity. Such agonization is reversible or, alternatively, irreversible.

Methods of Cellular Nucleic Acid Delivery.

Methods of the present invention enhance nucleic acid delivery into a cell population, in vivo, ex vivo, or in culture. For example, a cell culture containing a plurality of host cells (e.g., eukaryotic cells such as yeast or mammalian 5 cells) is contacted with a composition that contains an enhanced nucleic acid having at least one nucleoside modification and, optionally, a translatable region encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an antimicrobial polypeptide described herein. 10 The composition also generally contains a transfection reagent or other compound that increases the efficiency of enhanced nucleic acid uptake into the host cells. The enhanced nucleic acid exhibits enhanced retention in the cell population, relative to a corresponding unmodified nucleic 15 acid. The retention of the enhanced nucleic acid is greater than the retention of the unmodified nucleic acid. In some embodiments, it is at least about 50%, 75%, 90%, 95%, 100%, 150%, 200%, or more than 200% greater than the retention of the unmodified nucleic acid. Such retention 20 advantage may be achieved by one round of transfection with the enhanced nucleic acid, or may be obtained following repeated rounds of transfection.

In some embodiments, the enhanced nucleic acid is delivered to a target cell population with one or more additional 25 nucleic acids. Such delivery may be at the same time, or the enhanced nucleic acid is delivered prior to delivery of the one or more additional nucleic acids. The additional one or more nucleic acids may be modified nucleic acids or unmodified nucleic acids. It is understood that the initial 30 presence of the enhanced nucleic acids does not substantially induce an innate immune response of the cell population and, moreover, that the innate immune response will not be activated by the later presence of the unmodified nucleic acids. In this regard, the enhanced nucleic acid may 35 not itself contain a translatable region, if the protein desired to be present in the target cell population is translated from the unmodified nucleic acids.

Pharmaceutical Compositions

The present invention provides enhanced nucleic acids 40 (e.g., nucleic acids described herein), and complexes containing enhanced nucleic acids associated with other deliverable moieties. Thus, the present invention provides pharmaceutical compositions comprising one or more enhanced nucleic acids, or one or more such complexes, and one or 45 more pharmaceutically acceptable excipients. Pharmaceutical compositions may optionally comprise one or more additional therapeutically active substances. In some embodiments, compositions are administered to humans. For the purposes of the present disclosure, the phrase "active 50 ingredient" generally refers to an enhanced nucleic acid to be delivered as described herein.

Although the descriptions of pharmaceutical compositions provided herein are principally directed to pharmaceutical compositions which are suitable for administration to 55 humans, it will be understood by the skilled artisan that such compositions are generally suitable for administration to animals of all sorts. Modification of pharmaceutical compositions suitable for administration to humans in order to render the compositions suitable for administration to various animals is well understood, and the ordinarily skilled veterinary pharmacologist can design and/or perform such modification with merely ordinary, if any, experimentation. Subjects to which administration of the pharmaceutical compositions is contemplated include, but are not limited to, 65 humans and/or other animals (e.g., primates, mammals), including commercially relevant mammals such as cattle,

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pigs, horses, sheep, cats, dogs, mice, and/or rats; and/or birds, including commercially relevant birds such as chickens, ducks, geese, and/or turkeys.

Formulations of the pharmaceutical compositions described herein may be prepared by any method known or hereafter developed in the art of pharmacology. In general, such preparatory methods include the step of bringing the active ingredient into association with an excipient and/or one or more other accessory ingredients, and then, if necessary and/or desirable, shaping and/or packaging the product into a desired single- or multi-dose unit.

A pharmaceutical composition in accordance with the invention may be prepared, packaged, and/or sold in bulk, as a single unit dose, and/or as a plurality of single unit doses. As used herein, a "unit dose" is discrete amount of the pharmaceutical composition comprising a predetermined amount of the active ingredient. The amount of the active ingredient is generally equal to the dosage of the active ingredient which would be administered to a subject and/or a convenient fraction of such a dosage such as, for example, one-half or one-third of such a dosage.

Relative amounts of the active ingredient, the pharmaceutically acceptable excipient, and/or any additional ingredients in a pharmaceutical composition in accordance with the invention will vary, depending upon the identity, size, and/or condition of the subject treated and further depending upon the route by which the composition is to be administered. By way of example, the composition may comprise between 0.1% and 100% (w/w) active ingredient.

Pharmaceutical formulations may additionally comprise a pharmaceutically acceptable excipient, which, as used herein, includes any and all solvents, dispersion media, diluents, or other liquid vehicles, dispersion or suspension aids, surface active agents, isotonic agents, thickening or emulsifying agents, preservatives, solid binders, lubricants and the like, as suited to the particular dosage form desired. Remington's The Science and Practice of Pharmacy, 21st Edition, A. R. Gennaro (Lippincott, Williams & Wilkins, Baltimore, Md., 2006; incorporated herein by reference) discloses various excipients used in formulating pharmaceutical compositions and known techniques for the preparation thereof. Except insofar as any conventional excipient medium is incompatible with a substance or its derivatives, such as by producing any undesirable biological effect or otherwise interacting in a deleterious manner with any other component(s) of the pharmaceutical composition, its use is contemplated to be within the scope of this invention.

In some embodiments, a pharmaceutically acceptable excipient is at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% pure. In some embodiments, an excipient is approved for use in humans and for veterinary use. In some embodiments, an excipient is approved by United States Food and Drug Administration. In some embodiments, an excipient is pharmaceutical grade. In some embodiments, an excipient meets the standards of the United States Pharmacopoeia (USP), the European Pharmacopoeia (EP), the British Pharmacopoeia, and/or the International Pharmacopoeia.

Pharmaceutically acceptable excipients used in the manufacture of pharmaceutical compositions include, but are not limited to, inert diluents, dispersing and/or granulating agents, surface active agents and/or emulsifiers, disintegrating agents, binding agents, preservatives, buffering agents, lubricating agents, and/or oils. Such excipients may optionally be included in pharmaceutical formulations. Excipients such as cocoa butter and suppository waxes, coloring agents,

coating agents, sweetening, flavoring, and/or perfuming agents can be present in the composition, according to the judgment of the formulator.

Exemplary diluents include, but are not limited to, calcium carbonate, sodium carbonate, calcium phosphate, 5 dicalcium phosphate, calcium sulfate, calcium hydrogen phosphate, sodium phosphate lactose, sucrose, cellulose, microcrystalline cellulose, kaolin, mannitol, sorbitol, inositol, sodium chloride, dry starch, cornstarch, powdered sugar, etc., and/or combinations thereof.

Exemplary granulating and/or dispersing agents include, but are not limited to, potato starch, corn starch, tapioca starch, sodium starch glycolate, clays, alginic acid, guar gum, citrus pulp, agar, bentonite, cellulose and wood products, natural sponge, cation-exchange resins, calcium carbonate, silicates, sodium carbonate, cross-linked poly(vinyl-pyrrolidone) (crospovidone), sodium carboxymethyl starch (sodium starch glycolate), carboxymethyl cellulose, cross-linked sodium carboxymethyl cellulose (croscarmellose), methylcellulose, pregelatinized starch (starch 1500), microcrystalline starch, water insoluble starch, calcium carboxymethyl cellulose, magnesium aluminum silicate (Veegum), sodium lauryl sulfate, quaternary ammonium compounds, etc., and/or combinations thereof.

Exemplary surface active agents and/or emulsifiers 25 include, but are not limited to, natural emulsifiers (e.g. acacia, agar, alginic acid, sodium alginate, tragacanth, chondrux, cholesterol, xanthan, pectin, gelatin, egg yolk, casein, wool fat, cholesterol, wax, and lecithin), colloidal clays (e.g. bentonite [aluminum silicate] and Veegum® [magnesium 30 aluminum silicate]), long chain amino acid derivatives, high molecular weight alcohols (e.g. stearyl alcohol, cetyl alcohol, oleyl alcohol, triacetin monostearate, ethylene glycol distearate, glyceryl monostearate, and propylene glycol monostearate, polyvinyl alcohol), carbomers (e.g. carboxy 35 polymethylene, polyacrylic acid, acrylic acid polymer, and carboxyvinyl polymer), carrageenan, cellulosic derivatives (e.g. carboxymethylcellulose sodium, powdered cellulose, hydroxymethyl cellulose, hydroxypropyl hydroxypropyl methylcellulose, methylcellulose), sorbitan 40 fatty acid esters (e.g. polyoxyethylene sorbitan monolaurate [Tween®20], polyoxyethylene sorbitan [Tween®60], polyoxyethylene sorbitan monooleate [Tween®80], sorbitan monopalmitate [Span®40], sorbitan monostearate [Span®60], sorbitan tristearate [Span 65], glyceryl 45 monooleate, sorbitan monooleate [Span®80]), polyoxyethylene esters (e.g. polyoxyethylene monostearate [Myrj®45], polyoxyethylene hydrogenated castor oil, polyethoxylated castor oil, polyoxymethylene stearate, and Solutol®), sucrose fatty acid esters, polyethylene glycol fatty acid 50 esters (e.g. Cremophor®), polyoxyethylene ethers, (e.g. polyoxyethylene lauryl ether [Brij® 30]), poly(vinyl-pyrrolidone), diethylene glycol monolaurate, triethanolamine oleate, sodium oleate, potassium oleate, ethyl oleate, oleic acid, ethyl laurate, sodium lauryl sulfate, Pluronic®F 68, 55 Poloxamer®188, cetrimonium bromide, cetylpyridinium chloride, benzalkonium chloride, docusate sodium, etc. and/ or combinations thereof.

Exemplary binding agents include, but are not limited to, starch (e.g. cornstarch and starch paste); gelatin; sugars (e.g. 60 sucrose, glucose, dextrose, dextrin, molasses, lactose, lactitol, mannitol,); natural and synthetic gums (e.g. *acacia*, sodium alginate, extract of Irish moss, panwar gum, ghatti gum, mucilage of isapol husks, carboxymethylcellulose, methylcellulose, ethylcellulose, hydroxyethylcellulose, 65 hydroxypropyl cellulose, hydroxypropyl methylcellulose, microcrystalline cellulose, cellulose acetate, poly(vinyl-pyr-

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rolidone), magnesium aluminum silicate (Veegum®), and larch arabogalactan); alginates; polyethylene oxide; polyethylene glycol; inorganic calcium salts; silicic acid; polymethacrylates; waxes; water; alcohol; etc.; and combinations thereof.

Exemplary preservatives may include, but are not limited to, antioxidants, chelating agents, antimicrobial preservatives, antifungal preservatives, alcohol preservatives, acidic preservatives, and/or other preservatives. Exemplary antioxidants include, but are not limited to, alpha tocopherol, ascorbic acid, acorbyl palmitate, butylated hydroxyanisole, butylated hydroxytoluene, monothioglycerol, potassium metabisulfite, propionic acid, propyl gallate, sodium ascorbate, sodium bisulfite, sodium metabisulfite, and/or sodium sulfite. Exemplary chelating agents include ethylenediaminetetraacetic acid (EDTA), citric acid monohydrate, disodium edetate, dipotassium edetate, edetic acid, fumaric acid, malic acid, phosphoric acid, sodium edetate, tartaric acid, and/or trisodium edetate. Exemplary antimicrobial preservatives include, but are not limited to, benzalkonium chloride, benzethonium chloride, benzyl alcohol, bronopol, cetrimide, cetylpyridinium chloride, chlorhexidine, chlorobutanol, chlorocresol, chloroxylenol, cresol, ethyl alcohol, glycerin, hexetidine, imidurea, phenol, phenoxyethanol, phenylethyl alcohol, phenylmercuric nitrate, propylene glycol, and/or thimerosal. Exemplary antifungal preservatives include, but are not limited to, butyl paraben, methyl paraben, ethyl paraben, propyl paraben, benzoic acid, hydroxybenzoic acid, potassium benzoate, potassium sorbate, sodium benzoate, sodium propionate, and/or sorbic acid. Exemplary alcohol preservatives include, but are not limited to, ethanol, polyethylene glycol, phenol, phenolic compounds, bisphenol, chlorobutanol, hydroxybenzoate, and/or phenylethyl alcohol. Exemplary acidic preservatives include, but are not limited to, vitamin A, vitamin C, vitamin E, beta-carotene, citric acid, acetic acid, dehydroacetic acid, ascorbic acid, sorbic acid, and/or phytic acid. Other preservatives include, but are not limited to, tocopherol, tocopherol acetate, deteroxime mesylate, cetrimide, butylated hydroxyanisol (BHA), butylated hydroxytoluened (BHT), ethylenediamine, sodium lauryl sulfate (SLS), sodium lauryl ether sulfate (SLES), sodium bisulfite, sodium metabisulfite, potassium sulfite, potassium metabisulfite, Glydant Plus®, Phenonip®, methylparaben, Germall® 115, Germaben®II, NeoloneTM, KathonTM, and/or Euxyl®.

Exemplary buffering agents include, but are not limited to, citrate buffer solutions, acetate buffer solutions, phosphate buffer solutions, ammonium chloride, calcium carbonate, calcium chloride, calcium citrate, calcium glubionate, calcium gluceptate, calcium gluconate, D-gluconic acid, calcium glycerophosphate, calcium lactate, propanoic acid, calcium levulinate, pentanoic acid, dibasic calcium phosphate, phosphoric acid, tribasic calcium phosphate, calcium hydroxide phosphate, potassium acetate, potassium chloride, potassium gluconate, potassium mixtures, dibasic potassium phosphate, monobasic potassium phosphate, potassium phosphate mixtures, sodium acetate, sodium bicarbonate, sodium chloride, sodium citrate, sodium lactate, dibasic sodium phosphate, monobasic sodium phosphate, sodium phosphate mixtures, tromethamine, magnesium hydroxide, aluminum hydroxide, alginic acid, pyrogen-free water, isotonic saline, Ringer's solution, ethyl alcohol, etc., and/or combinations thereof.

Exemplary lubricating agents include, but are not limited to, magnesium stearate, calcium stearate, stearic acid, silica, talc, malt, glyceryl behanate, hydrogenated vegetable oils, polyethylene glycol, sodium benzoate, sodium acetate,

sodium chloride, leucine, magnesium lauryl sulfate, sodium lauryl sulfate, etc., and combinations thereof.

Exemplary oils include, but are not limited to, almond, apricot kernel, avocado, babassu, bergamot, black current seed, borage, cade, camomile, canola, caraway, carnauba, castor, cinnamon, cocoa butter, coconut, cod liver, coffee, corn, cotton seed, emu, eucalvptus, evening primrose, fish, flaxseed, geraniol, gourd, grape seed, hazel nut, hyssop, isopropyl myristate, jojoba, kukui nut, lavandin, lavender, lemon, litsea cubeba, macademia nut, mallow, mango seed, meadowfoam seed, mink, nutmeg, olive, orange, orange roughy, palm, palm kernel, peach kernel, peanut, poppy seed, pumpkin seed, rapeseed, rice bran, rosemary, safflower, sandalwood, sasquana, savoury, sea buckthorn, 15 sesame, shea butter, silicone, soybean, sunflower, tea tree, thistle, tsubaki, vetiver, walnut, and wheat germ oils. Exemplary oils include, but are not limited to, butyl stearate, caprylic triglyceride, capric triglyceride, cyclomethicone, diethyl sebacate, dimethicone 360, isopropyl myristate, min- 20 eral oil, octyldodecanol, oleyl alcohol, silicone oil, and/or combinations thereof.

Liquid dosage forms for oral and parenteral administration include, but are not limited to, pharmaceutically acceptable emulsions, microemulsions, solutions, suspensions, 25 syrups, and/or elixirs. In addition to active ingredients, liquid dosage forms may comprise inert diluents commonly used in the art such as, for example, water or other solvents, solubilizing agents and emulsifiers such as ethyl alcohol, isopropyl alcohol, ethyl carbonate, ethyl acetate, benzyl 30 alcohol, benzyl benzoate, propylene glycol, 1,3-butylene glycol, dimethylformamide, oils (in particular, cottonseed, groundnut, corn, germ, olive, castor, and sesame oils), glycerol, tetrahydrofurfuryl alcohol, polyethylene glycols and fatty acid esters of sorbitan, and mixtures thereof. 35 Besides inert diluents, oral compositions can include adjuvants such as wetting agents, emulsifying and suspending agents, sweetening, flavoring, and/or perfuming agents. In certain embodiments for parenteral administration, compositions are mixed with solubilizing agents such as Cremo- 40 phor®, alcohols, oils, modified oils, glycols, polysorbates, cyclodextrins, polymers, and/or combinations thereof.

Injectable preparations, for example, sterile injectable aqueous or oleaginous suspensions may be formulated according to the known art using suitable dispersing agents, 45 wetting agents, and/or suspending agents. Sterile injectable preparations may be sterile injectable solutions, suspensions, and/or emulsions in nontoxic parenterally acceptable diluents and/or solvents, for example, as a solution in 1,3-butanediol. Among the acceptable vehicles and solvents that may be employed are water, Ringer's solution, U.S.P., and isotonic sodium chloride solution. Sterile, fixed oils are conventionally employed as a solvent or suspending medium. For this purpose any bland fixed oil can be employed including synthetic mono- or diglycerides. Fatty 55 acids such as oleic acid can be used in the preparation of injectables.

Injectable formulations can be sterilized, for example, by filtration through a bacterial-retaining filter, and/or by incorporating sterilizing agents in the form of sterile solid compositions which can be dissolved or dispersed in sterile water or other sterile injectable medium prior to use.

In order to prolong the effect of an active ingredient, it is often desirable to slow the absorption of the active ingredient from subcutaneous or intramuscular injection. This may 65 be accomplished by the use of a liquid suspension of crystalline or amorphous material with poor water solubility.

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The rate of absorption of the drug then depends upon its rate of dissolution which, in turn, may depend upon crystal size and crystalline form. Alternatively, delayed absorption of a parenterally administered drug form is accomplished by dissolving or suspending the drug in an oil vehicle. Injectable depot forms are made by forming microencapsule matrices of the drug in biodegradable polymers such as polylactide-polyglycolide. Depending upon the ratio of drug to polymer and the nature of the particular polymer employed, the rate of drug release can be controlled. Examples of other biodegradable polymers include poly (orthoesters) and poly(anhydrides). Depot injectable formulations are prepared by entrapping the drug in liposomes or microemulsions which are compatible with body tissues.

Compositions for rectal or vaginal administration are typically suppositories which can be prepared by mixing compositions with suitable non-irritating excipients such as cocoa butter, polyethylene glycol or a suppository wax which are solid at ambient temperature but liquid at body temperature and therefore melt in the rectum or vaginal cavity and release the active ingredient. Solid dosage forms for oral administration include capsules, tablets, pills, powders, and granules. In such solid dosage forms, an active ingredient is mixed with at least one inert, pharmaceutically acceptable excipient such as sodium citrate or dicalcium phosphate and/or fillers or extenders (e.g. starches, lactose, sucrose, glucose, mannitol, and silicic acid), binders (e.g. carboxymethylcellulose, alginates, gelatin, polyvinylpyrrolidinone, sucrose, and acacia), humectants (e.g. glycerol), disintegrating agents (e.g. agar, calcium carbonate, potato or tapioca starch, alginic acid, certain silicates, and sodium carbonate), solution retarding agents (e.g. paraffin), absorption accelerators (e.g. quaternary ammonium compounds), wetting agents (e.g. cetyl alcohol and glycerol monostearate), absorbents (e.g. kaolin and bentonite clay), and lubricants (e.g. talc, calcium stearate, magnesium stearate, solid polyethylene glycols, sodium lauryl sulfate), and mixtures thereof. In the case of capsules, tablets and pills, the dosage form may comprise buffering agents.

Solid compositions of a similar type may be employed as fillers in soft and hard-filled gelatin capsules using such excipients as lactose or milk sugar as well as high molecular weight polyethylene glycols and the like. Solid dosage forms of tablets, dragees, capsules, pills, and granules can be prepared with coatings and shells such as enteric coatings and other coatings well known in the pharmaceutical formulating art. They may optionally comprise opacifying agents and can be of a composition that they release the active ingredient(s) only, or preferentially, in a certain part of the intestinal tract, optionally, in a delayed manner. Examples of embedding compositions which can be used include polymeric substances and waxes. Solid compositions of a similar type may be employed as fillers in soft and hard-filled gelatin capsules using such excipients as lactose or milk sugar as well as high molecular weight polyethylene glycols and the like.

Dosage forms for topical and/or transdermal administration of a composition may include ointments, pastes, creams, lotions, gels, powders, solutions, sprays, inhalants and/or patches. Generally, an active ingredient is admixed under sterile conditions with a pharmaceutically acceptable excipient and/or any needed preservatives and/or buffers as may be required. Additionally, the present invention contemplates the use of transdermal patches, which often have the added advantage of providing controlled delivery of a compound to the body. Such dosage forms may be prepared, for example, by dissolving and/or dispensing the compound

in the proper medium. Alternatively or additionally, rate may be controlled by either providing a rate controlling membrane and/or by dispersing the compound in a polymer matrix and/or gel.

Suitable devices for use in delivering intradermal pharmaceutical compositions described herein include short needle devices such as those described in U.S. Pat. Nos. 4,886,499; 5,190,521; 5,328,483; 5,527,288; 4,270,537; 5,015,235; 5,141,496; and 5,417,662. Intradermal compositions may be administered by devices which limit the effective penetration length of a needle into the skin, such as those described in PCT publication WO 99/34850 and functional equivalents thereof. Jet injection devices which deliver liquid compositions to the dermis via a liquid jet 15 injector and/or via a needle which pierces the stratum corneum and produces a jet which reaches the dermis are suitable. Jet injection devices are described, for example, in U.S. Pat. Nos. 5,480,381; 5,599,302; 5,334,144; 5,993,412; 5,649,912; 5,569,189; 5,704,911; 5,383,851; 5,893,397; 20 5,466,220; 5,339,163; 5,312,335; 5,503,627; 5,064,413; 5,520,639; 4,596,556; 4,790,824; 4,941,880; 4,940,460; and PCT publications WO 97/37705 and WO 97/13537. Ballistic powder/particle delivery devices which use compressed gas to accelerate vaccine in powder form through the outer 25 layers of the skin to the dermis are suitable. Alternatively or additionally, conventional syringes may be used in the classical mantoux method of intradermal administration.

Formulations suitable for topical administration include, but are not limited to, liquid and/or semi liquid preparations 30 such as liniments, lotions, oil in water and/or water in oil emulsions such as creams, ointments and/or pastes, and/or solutions and/or suspensions. Topically-administrable formulations may, for example, comprise from about 1% to about 10% (w/w) active ingredient, although the concentration of active ingredient may be as high as the solubility limit of the active ingredient in the solvent. Formulations for topical administration may further comprise one or more of the additional ingredients described herein.

A pharmaceutical composition may be prepared, pack- 40 aged, and/or sold in a formulation suitable for pulmonary administration via the buccal cavity. Such a formulation may comprise dry particles which comprise the active ingredient and which have a diameter in the range from about 0.5 nm to about 7 nm or from about 1 nm to about 6 nm. Such 45 compositions are suitably in the form of dry powders for administration using a device comprising a dry powder reservoir to which a stream of propellant may be directed to disperse the powder and/or using a self propelling solvent/ powder dispensing container such as a device comprising 50 the active ingredient dissolved and/or suspended in a lowboiling propellant in a sealed container. Such powders comprise particles wherein at least 98% of the particles by weight have a diameter greater than 0.5 nm and at least 95% of the particles by number have a diameter less than 7 nm. 55 Alternatively, at least 95% of the particles by weight have a diameter greater than 1 nm and at least 90% of the particles by number have a diameter less than 6 nm. Dry powder compositions may include a solid fine powder diluent such as sugar and are conveniently provided in a unit dose form. 60

Low boiling propellants generally include liquid propellants having a boiling point of below 65° F. at atmospheric pressure. Generally the propellant may constitute 50% to 99.9% (w/w) of the composition, and active ingredient may constitute 0.1% to 20% (w/w) of the composition. A propellant may further comprise additional ingredients such as a liquid non-ionic and/or solid anionic surfactant and/or a

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solid diluent (which may have a particle size of the same order as particles comprising the active ingredient).

Pharmaceutical compositions formulated for pulmonary delivery may provide an active ingredient in the form of droplets of a solution and/or suspension. Such formulations may be prepared, packaged, and/or sold as aqueous and/or dilute alcoholic solutions and/or suspensions, optionally sterile, comprising active ingredient, and may conveniently be administered using any nebulization and/or atomization device. Such formulations may further comprise one or more additional ingredients including, but not limited to, a flavoring agent such as saccharin sodium, a volatile oil, a buffering agent, a surface active agent, and/or a preservative such as methylhydroxybenzoate. Droplets provided by this route of administration may have an average diameter in the range from about 0.1 nm to about 200 nm.

Formulations described herein as being useful for pulmonary delivery are useful for intranasal delivery of a pharmaceutical composition. Another formulation suitable for intranasal administration is a coarse powder comprising the active ingredient and having an average particle from about 0.2 μm to 500 μm . Such a formulation is administered in the manner in which snuff is taken, i.e. by rapid inhalation through the nasal passage from a container of the powder held close to the nose.

Formulations suitable for nasal administration may, for example, comprise from about as little as 0.1% (w/w) and as much as 100% (w/w) of active ingredient, and may comprise one or more of the additional ingredients described herein. A pharmaceutical composition may be prepared, packaged, and/or sold in a formulation suitable for buccal administration. Such formulations may, for example, be in the form of tablets and/or lozenges made using conventional methods, and may have, for example, 0.1% to 20% (w/w) active ingredient, the balance comprising an orally dissolvable and/or degradable composition and, optionally, one or more of the additional ingredients described herein. Alternately, formulations suitable for buccal administration may comprise a powder and/or an aerosolized and/or atomized solution and/or suspension comprising active ingredient. Such powdered, aerosolized, and/or aerosolized formulations, when dispersed, may have an average particle and/or droplet size in the range from about 0.1 nm to about 200 nm, and may further comprise one or more of any additional ingredients described herein.

A pharmaceutical composition may be prepared, packaged, and/or sold in a formulation suitable for ophthalmic administration. Such formulations may, for example, be in the form of eye drops including, for example, a 0.1/1.0% (w/w) solution and/or suspension of the active ingredient in an aqueous or oily liquid excipient. Such drops may further comprise buffering agents, salts, and/or one or more other of any additional ingredients described herein. Other opthalmically-administrable formulations which are useful include those which comprise the active ingredient in microcrystalline form and/or in a liposomal preparation. Ear drops and/or eye drops are contemplated as being within the scope of this invention.

General considerations in the formulation and/or manufacture of pharmaceutical agents may be found, for example, in *Remington: The Science and Practice of Pharmacy* 21st ed., Lippincott Williams & Wilkins, 2005 (incorporated herein by reference).

The present invention provides methods comprising administering modified mRNAs and their encoded proteins or complexes in accordance with the invention to a subject in need thereof. Nucleic acids, proteins or complexes, or

pharmaceutical, imaging, diagnostic, or prophylactic compositions thereof, may be administered to a subject using any amount and any route of administration effective for preventing, treating, diagnosing, or imaging a disease, disorder, and/or condition (e.g., a disease, disorder, and/or condition 5 relating to microbial infections). The exact amount required will vary from subject to subject, depending on the species, age, and general condition of the subject, the severity of the disease, the particular composition, its mode of administration, its mode of activity, and the like. Compositions in 10 accordance with the invention are typically formulated in dosage unit form for ease of administration and uniformity of dosage. It will be understood, however, that the total daily usage of the compositions of the present invention will be decided by the attending physician within the scope of sound medical judgment. The specific therapeutically effective, prophylactially effective, or appropriate imaging dose level for any particular patient will depend upon a variety of factors including the disorder being treated and the severity of the disorder; the activity of the specific compound 20 employed; the specific composition employed; the age, body weight, general health, sex and diet of the patient; the time of administration, route of administration, and rate of excretion of the specific compound employed; the duration of the treatment; drugs used in combination or coincidental with 25 the specific compound employed; and like factors well known in the medical arts.

Devices may also be used in conjunction with the present invention. In one embodiment, a device is used to assess levels of a protein which has been administered in the form 30 of a modified mRNA. The device may comprise a blood, urine or other biofluidic test. It may be as large as to include an automated central lab platform or a small decentralized bench top device.

Kits.

The invention provides a variety of kits for conveniently and/or effectively carrying out methods of the present invention. Typically kits will comprise sufficient amounts and/or numbers of components to allow a user to perform multiple treatments of a subject(s) and/or to perform multiple experi-

In one embodiment, the levels of a modified mRNA of the present invention may be measured by immunoassay. In this embodiment, the assay may be used to assess levels of modified mRNA or its activated form or a variant delivered 45 as or in response to the administration of the modified mRNA.

Dressings and Bandages.

The invention provides a variety of dressings (e.g., wound dressings) or bandages (e.g., adhesive bandages) for conveniently and/or effectively carrying out methods of the present invention. Typically dressings or bandages will comprise sufficient amounts of pharmaceutical compositions and/or modified nucleic acids described herein to allow a user to perform multiple treatments of a subject(s).

Animal Models.

Anti-microbial agents (e.g., anti-microbial polypeptides) can be tested in healthy animals (e.g., mice) exposed to specific microbial pathogens (e.g., bacteria). Anti-microbial agents (e.g., anti-microbial polypeptides) can also be tested 60 in immunodeficient animal (e.g., mouse) models to test infection process without interference from other immune mechanisms except innate immunity.

Severe Combined Immunodeficiency (SCID) is a severe immunodeficiency genetic disorder that is characterized by the complete inability of the adaptive immune system to mount, coordinate, and sustain an appropriate immune 44

response, usually due to absent or atypical T and B lymphocytes. Scid mice are important tools for researching hematopoiesis, innate and adaptive immunity, autoimmunity, infectious diseases, cancer, vaccine development, and regenerative medicine in vivo.

Strain NOD.Cg-Prkde^{scid} Il2rg^{tm1 Wjl}/SzJ (005557 Jacson Lab), commonly known as NOD scid gamma (NSG), is the latest breakthrough in the development of immunodeficient models. It combines the innate immunity deficiencies of the NOD/ShiLtJ background, the scid mutation, and IL2 receptor gamma chain (Il2rg) deficiency. The latter two deficiencies combine to eliminate mature T cells, B cells, and NK cells. Because the Il2rg knockout prevents the development of lymphoma, NSG mice survive longer than other scid strains, enabling long-term experiments.

The B6 scid—strain B6.CB17-Prkdc^{scid}/SzJ (001913, Jacson Lab), B6 scid mice lack most B and T cells. B6 scid is more severely immunodeficient and supports better engraftment of allogeneic and xenogeneic cells, tissues, and tumors than Foxn1^{nu} mutant strains.

The humanized mouse model of HIV infection to investigate mechanisms of viral dissemination, of HIV-induced immune activation, and of HIV-induced immune dysfunction can be used too MGH. Another mouse model—EcoHIV infected about 75 percent of the mice tested, an efficiency rate comparable with that of HIV in humans. The EcoHIV infection was present in immune cells and white blood cells, the spleen, abdominal cavity and brain.

C57BL/6-Btk^{tm1.Arte} 9723-F-mouse model for Bruton's disease. Bruton's tyrosine kinase (Btk) is a member of the Tec kinase family and has been implicated in the primary immunodeficiency X-linked agammaglobulinemia. Btk is thought to play multiple roles in the haematopoietic system, including B-cell development, stimulation of mast cells and the onset of autoimmune diseases. The Btk (Bruton's tyrosine kinase) KinaseSwitch mouse strain carries point mutations at the genomic level at positions T474A/S538A in the ATP binding pocket of the Btk kinase domain (BtkT474A/S538A).

Definitions

Therapeutic Agent: The term "therapeutic agent" refers to any agent that, when administered to a subject, has a therapeutic, diagnostic, and/or prophylactic effect and/or elicits a desired biological and/or pharmacological effect.

Administered in combination: As used herein, the term "administered in combination" or "combined administration" means that two or more agents (e.g., a modified nucleic acid encoding an anti-microbial polypeptide (e.g., an anti-bacterial polypeptide), e.g., an anti-microbial polypeptide described herein and an anti-microbial agent (e.g., an anti-microbial polypeptide or a small molecule anti-microbial compound described herein)) are administered to a subject at the same time or within an interval such that there is overlap of an effect of each agent on the patient. In some embodiments, they are administered within about 60, 30, 15, 10, 5, or 1 minute of one another. In some embodiments, the administrations of the agents are spaced sufficiently close together such that a combinatorial (e.g., a synergistic) effect is achieved.

Animal: As used herein, the term "animal" refers to any member of the animal kingdom. In some embodiments, "animal" refers to humans at any stage of development. In some embodiments, "animal" refers to non-human animals at any stage of development. In certain embodiments, the non-human animal is a mammal (e.g., a rodent, a mouse, a

rat, a rabbit, a monkey, a dog, a cat, a sheep, cattle, a primate, or a pig). In some embodiments, animals include, but are not limited to, mammals, birds, reptiles, amphibians, fish, and worms. In some embodiments, the animal is a transgenic animal, genetically-engineered animal, or a clone.

Approximately: As used herein, the term "approximately" or "about," as applied to one or more values of interest, refers to a value that is similar to a stated reference value. In certain embodiments, the term "approximately" or "about" refers to a range of values that fall within 25%, 20%, 19%, 10 18%, 17%, 16%, 15%, 14%, 13%, 12%, 11%, 10%, 9%, 8%, 7%, 6%, 5%, 4%, 3%, 2%, 1%, or less in either direction (greater than or less than) of the stated reference value unless otherwise stated or otherwise evident from the context (except where such number would exceed 100% of a possible value).

Associated with: As used herein, the terms "associated with," "conjugated," "linked," "attached," and "tethered," when used with respect to two or more moieties, means that the moieties are physically associated or connected with one 20 another, either directly or via one or more additional moieties that serves as a linking agent, to form a structure that is sufficiently stable so that the moieties remain physically associated under the conditions in which the structure is used, e.g., physiological conditions. As used herein, the 25 terms "associated with," when used with respect to a microorganism (e.g., a bacterium) and a disease, disorder, or condition, means the microorganisms (e.g., bacterium) is found more frequently (e.g., at least 10%, 25%, 50%, 75%, 100%, 200%, 500%, 1000% more frequently) in patients 30 with the disease, disorder, or condition than in healthy controls and/or there is a frequent co-occurrence of the microorganisms (e.g., bacterium) in the disease, disorder, or condition. In some embodiments, the microorganisms (e.g., bacterium) can be a direct and/or singular cause of the 35 disease, disorder, or condition. In some embodiments, the microorganisms (e.g., bacterium) can be a necessary, but not sufficient, cause of the disease, disorder, or condition (e.g., only causes the disease, disorder or condition in combination with one or more other causal factors (e.g., genetic 40 factors, or toxin exposure)). In some embodiments, the bacterium can predispose to the development of or increase the risk of getting the disease, disorder, or condition. In some embodiments, the microorganisms (e.g., bacterium) can also be an "innocent bystander" that plays no significant role in 45 the etiology of the disease, disorder, or condition but is more prevalent in patients with the disease, disorder, or condition for some reason such as the compromised immune response caused by the disease, disorder, or condition.

Biologically active: As used herein, the phrase "biologically active" refers to a characteristic of any substance that has activity in a biological system and/or organism. For instance, a substance that, when administered to an organism, has a biological effect on that organism, is considered to be biologically active. In particular embodiments, where a nucleic acid is biologically active, a portion of that nucleic acid that shares at least one biological activity of the whole nucleic acid is typically referred to as a "biologically active" portion.

Conserved: As used herein, the term "conserved" refers to 60 nucleotides or amino acid residues of a polynucleotide sequence or amino acid sequence, respectively, that are those that occur unaltered in the same position of two or more related sequences being compared. Nucleotides or amino acids that are relatively conserved are those that are 65 conserved amongst more related sequences than nucleotides or amino acids appearing elsewhere in the sequences. In

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some embodiments, two or more sequences are said to be "completely conserved" if they are 100% identical to one another. In some embodiments, two or more sequences are said to be "highly conserved" if they are at least 70% identical, at least 80% identical, at least 90% identical, or at least 95% identical to one another. In some embodiments, two or more sequences are said to be "highly conserved" if they are about 70% identical, about 80% identical, about 90% identical, about 95%, about 98%, or about 99% identical to one another. In some embodiments, two or more sequences are said to be "conserved" if they are at least 30% identical, at least 40% identical, at least 50% identical, at least 60% identical, at least 70% identical, at least 80% identical, at least 90% identical, or at least 95% identical to one another. In some embodiments, two or more sequences are said to be "conserved" if they are about 30% identical, about 40% identical, about 50% identical, about 60% identical, about 70% identical, about 80% identical, about 90% identical, about 95% identical, about 98% identical, or about 99% identical to one another.

Cytostatic: As used herein, "cytostatic" refers to inhibiting, reducing, suppressing the growth, division, or multiplication of a cell (e.g., a mammalian cell (e.g., a human cell)), bacterium, virus, fungus, protozoan, parasite, prion, or a combination thereof.

Cytotoxic: As used herein, "cytotoxic" refers to killing or causing injurous, toxic, or deadly effect on a cell (e.g., a mammalian cell (e.g., a human cell)), bacterium, virus, fungus, protozoan, parasite, prion, or a combination thereof.

Expression: As used herein, "expression" of a nucleic acid sequence refers to one or more of the following events: (1) production of an RNA template from a DNA sequence (e.g., by transcription); (2) processing of an RNA transcript (e.g., by splicing, editing, 5' cap formation, and/or 3' end processing); (3) translation of an RNA into a polypeptide or protein; and (4) post-translational modification of a polypeptide or protein.

Functional: As used herein, a "functional" biological molecule is a biological molecule in a form in which it exhibits a property and/or activity by which it is characterized

Homology: As used herein, the term "homology" refers to the overall relatedness between polymeric molecules, e.g. between nucleic acid molecules (e.g. DNA molecules and/or RNA molecules) and/or between polypeptide molecules. In some embodiments, polymeric molecules are considered to be "homologous" to one another if their sequences are at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% identical. In some embodiments, polymeric molecules are considered to be "homologous" to one another if their sequences are at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% similar. The term "homologous" necessarily refers to a comparison between at least two sequences (nucleotides sequences or amino acid sequences). In accordance with the invention, two nucleotide sequences are considered to be homologous if the polypeptides they encode are at least about 50% identical, at least about 60% identical, at least about 70% identical, at least about 80% identical, or at least about 90% identical for at least one stretch of at least about 20 amino acids. In some embodiments, homologous nucleotide sequences are characterized by the ability to encode a stretch of at least 4-5

uniquely specified amino acids. Both the identity and the approximate spacing of these amino acids relative to one another must be considered for nucleotide sequences to be considered homologous. For nucleotide sequences less than 60 nucleotides in length, homology is determined by the 5 ability to encode a stretch of at least 4-5 uniquely specified amino acids. In accordance with the invention, two protein sequences are considered to be homologous if the proteins are at least about 50% identical, at least about 60% identical, at least about 80% identical, or 10 at least about 90% identical for at least one stretch of at least about 20 amino acids.

Identity: As used herein, the term "identity" refers to the overall relatedness between polymeric molecules, e.g., between nucleic acid molecules (e.g. DNA molecules and/or 15 RNA molecules) and/or between polypeptide molecules. Calculation of the percent identity of two nucleic acid sequences, for example, can be performed by aligning the two sequences for optimal comparison purposes (e.g., gaps can be introduced in one or both of a first and a second 20 nucleic acid sequences for optimal alignment and nonidentical sequences can be disregarded for comparison purposes). In certain embodiments, the length of a sequence aligned for comparison purposes is at least 30%, at least 40%, at least 50%, at least 60%, at least 70%, at least 80%, 25 at least 90%, at least 95%, or 100% of the length of the reference sequence. The nucleotides at corresponding nucleotide positions are then compared. When a position in the first sequence is occupied by the same nucleotide as the corresponding position in the second sequence, then the 30 molecules are identical at that position. The percent identity between the two sequences is a function of the number of identical positions shared by the sequences, taking into account the number of gaps, and the length of each gap, which needs to be introduced for optimal alignment of the 35 two sequences. The comparison of sequences and determination of percent identity between two sequences can be accomplished using a mathematical algorithm. For example, the percent identity between two nucleotide sequences can be determined using methods such as those described in 40 Computational Molecular Biology, Lesk, A. M., ed., Oxford University Press, New York, 1988; Biocomputing: Informatics and Genome Projects, Smith, D. W., ed., Academic Press, New York, 1993; Sequence Analysis in Molecular Biology, von Heinje, G., Academic Press, 1987; Computer Analysis 45 of Sequence Data, Part I, Griffin, A. M., and Griffin, H. G., eds., Humana Press, New Jersey, 1994; and Sequence Analysis Primer, Gribskov, M. and Devereux, J., eds., M Stockton Press, New York, 1991; each of which is incorporated herein by reference. For example, the percent identity 50 between two nucleotide sequences can be determined using the algorithm of Meyers and Miller (CABIOS, 1989, 4:11-17), which has been incorporated into the ALIGN program (version 2.0) using a PAM120 weight residue table, a gap length penalty of 12 and a gap penalty of 4. The percent 55 identity between two nucleotide sequences can, alternatively, be determined using the GAP program in the GCG software package using an NWSgapdna.CMP matrix. Methods commonly employed to determine percent identity between sequences include, but are not limited to those 60 disclosed in Carillo, H., and Lipman, D., SIAM J Applied Math., 48:1073 (1988); incorporated herein by reference. Techniques for determining identity are codified in publicly available computer programs. Exemplary computer software to determine homology between two sequences 65 include, but are not limited to, GCG program package, Devereux, J., et al., Nucleic Acids Research, 12(1), 387

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(1984)), BLASTP, BLASTN, and FASTA Atschul, S. F. et al., J. Molec. Biol., 215, 403 (1990)).

Inhibit expression of a gene: As used herein, the phrase "inhibit expression of a gene" means to cause a reduction in the amount of an expression product of the gene. The expression product can be an RNA transcribed from the gene (e.g., an mRNA) or a polypeptide translated from an mRNA transcribed from the gene. Typically a reduction in the level of an mRNA results in a reduction in the level of a polypeptide translated therefrom. The level of expression may be determined using standard techniques for measuring mRNA or protein.

In vitro: As used herein, the term "in vitro" refers to events that occur in an artificial environment, e.g., in a test tube or reaction vessel, in cell culture, in a Petri dish, etc., rather than within an organism (e.g., animal, plant, or microbe).

In vivo: As used herein, the term "in vivo" refers to events that occur within an organism (e.g., animal, plant, or microbe).

Isolated: As used herein, the term "isolated" refers to a substance or entity that has been (1) separated from at least some of the components with which it was associated when initially produced (whether in nature or in an experimental setting), and/or (2) produced, prepared, and/or manufactured by the hand of man. Isolated substances and/or entities may be separated from at least about 10%, about 20%, about 30%, about 40%, about 50%, about 60%, about 70%, about 80%, about 90%, or more of the other components with which they were initially associated. In some embodiments, isolated agents are more than about 80%, about 95%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or more than about 99% pure. As used herein, a substance is "pure" if it is substantially free of other components.

Preventing: As used herein, the term "preventing" refers to partially or completely delaying onset of a microbial infection; partially or completely delaying onset of one or more symptoms, features, or clinical manifestations of a particular disease, disorder, and/or condition associated with a microbial infection; partially or completely delaying onset of one or more symptoms, features, or manifestations of a particular disease, disorder, and/or condition prior to an identifiable microbial infection; partially or completely delaying progression from an latent microbial infection to an active microbial infection or a particular disease, disorder and/or condition; and/or decreasing the risk of developing pathology associated with the microbial infection or the disease, disorder, and/or condition.

Similarity: As used herein, the term "similarity" refers to the overall relatedness between polymeric molecules, e.g. between nucleic acid molecules (e.g. DNA molecules and/or RNA molecules) and/or between polypeptide molecules. Calculation of percent similarity of polymeric molecules to one another can be performed in the same manner as a calculation of percent identity, except that calculation of percent similarity takes into account conservative substitutions as is understood in the art.

Subject: As used herein, the term "subject" or "patient" refers to any organism to which a composition in accordance with the invention may be administered, e.g., for experimental, diagnostic, prophylactic, and/or therapeutic purposes. Typical subjects include animals (e.g., mammals such as mice, rats, rabbits, non-human primates, and humans) and/or plants.

Substantially: As used herein, the term "substantially" refers to the qualitative condition of exhibiting total or

near-total extent or degree of a characteristic or property of interest. One of ordinary skill in the biological arts will understand that biological and chemical phenomena rarely, if ever, go to completion and/or proceed to completeness or achieve or avoid an absolute result. The term "substantially" is therefore used herein to capture the potential lack of completeness inherent in many biological and chemical phenomena.

Suffering from: An individual who is "suffering from" a disease, disorder, and/or condition has been diagnosed with 10 or displays one or more symptoms of a disease, disorder, and/or condition.

Susceptible to: An individual who is "susceptible to" a disease, disorder, and/or condition has not been diagnosed with and/or may not exhibit symptoms of the disease, 15 disorder, and/or condition. In some embodiments, an individual who is susceptible to a disease, disorder, and/or condition (for example, cancer) may be characterized by one or more of the following: (1) a genetic mutation associated with development of the disease, disorder, and/or condition; 20 (2) a genetic polymorphism associated with development of the disease, disorder, and/or condition; (3) increased and/or decreased expression and/or activity of a protein and/or nucleic acid associated with the disease, disorder, and/or condition; (4) habits and/or lifestyles associated with devel- 25 opment of the disease, disorder, and/or condition; (5) a family history of the disease, disorder, and/or condition; and (6) exposure to and/or infection with a microbe associated with development of the disease, disorder, and/or condition. In some embodiments, an individual who is susceptible to a 30 disease, disorder, and/or condition will develop the disease, disorder, and/or condition. In some embodiments, an individual who is susceptible to a disease, disorder, and/or condition will not develop the disease, disorder, and/or condition.

Therapeutically effective amount: As used herein, the term "therapeutically effective amount" means an amount of an agent to be delivered (e.g., nucleic acid, drug, therapeutic agent, diagnostic agent, prophylactic agent, etc.) that is sufficient, when administered to a subject suffering from or 40 susceptible to a disease, disorder, and/or condition, to treat, improve symptoms of, diagnose, prevent, and/or delay the onset of the disease, disorder, and/or condition.

Transcription factor: As used herein, the term "transcription factor" refers to a DNA-binding protein that regulates 45 transcription of DNA into RNA, for example, by activation or repression of transcription. Some transcription factors effect regulation of transcription alone, while others act in concert with other proteins. Some transcription factor can both activate and repress transcription under certain conditions. In general, transcription factors bind a specific target sequence or sequences highly similar to a specific consensus sequence in a regulatory region of a target gene. Transcription factors may regulate transcription of a target gene alone or in a complex with other molecules.

Treating: As used herein, the term "treating" refers to partially or completely alleviating, ameliorating, improving, relieving, delaying onset of, inhibiting progression of, reducing severity of, and/or reducing incidence of one or more symptoms, features, or clinical manifestations of a 60 particular disease, disorder, and/or condition. For example, "treating" microbial infections may refer to inhibit or reduce the survival, growth, and/or spread of the microbial pathogens. "Treating" cancer may refer to inhibiting survival, growth, and/or spread of a tumor. Treatment may be administered to a subject who does not exhibit signs of a disease, disorder, and/or condition (e.g., prior to an identifiable

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microbial infection) and/or to a subject who exhibits only early signs of a disease, disorder, and/or condition for the purpose of decreasing the risk of developing pathology associated with the disease, disorder, and/or condition.

Unmodified: As used herein, "unmodified" refers to the protein or agent prior to being modified.

Equivalents And Scope

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments, described herein. The scope of the present invention is not intended to be limited to the above Description, but rather is as set forth in the appended claims.

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments in accordance with the invention described herein. The scope of the present invention is not intended to be limited to the above Description, but rather is as set forth in the appended claims.

In the claims articles such as "a," "an," and "the" may mean one or more than one unless indicated to the contrary or otherwise evident from the context. Claims or descriptions that include "or" between one or more members of a group are considered satisfied if one, more than one, or all of the group members are present in, employed in, or otherwise relevant to a given product or process unless indicated to the contrary or otherwise evident from the context. The invention includes embodiments in which exactly one member of the group is present in, employed in, or otherwise relevant to a given product or process. The invention includes embodiments in which more than one, or all of the group members are present in, employed in, or 35 otherwise relevant to a given product or process. Furthermore, it is to be understood that the invention encompasses all variations, combinations, and permutations in which one or more limitations, elements, clauses, descriptive terms, etc., from one or more of the listed claims is introduced into another claim. For example, any claim that is dependent on another claim can be modified to include one or more limitations found in any other claim that is dependent on the same base claim. Furthermore, where the claims recite a composition, it is to be understood that methods of using the composition for any of the purposes disclosed herein are included, and methods of making the composition according to any of the methods of making disclosed herein or other methods known in the art are included, unless otherwise indicated or unless it would be evident to one of ordinary skill in the art that a contradiction or inconsistency would arise.

Where elements are presented as lists, e.g., in Markush group format, it is to be understood that each subgroup of the elements is also disclosed, and any element(s) can be removed from the group. It should it be understood that, in general, where the invention, or aspects of the invention, is/are referred to as comprising particular elements, features, etc., certain embodiments of the invention or aspects of the invention consist, or consist essentially of, such elements, features, etc. For purposes of simplicity those embodiments have not been specifically set forth in haec verba herein. It is also noted that the term "comprising" is intended to be open and permits the inclusion of additional elements or steps.

Where ranges are given, endpoints are included. Furthermore, it is to be understood that unless otherwise indicated or otherwise evident from the context and understanding of

one of ordinary skill in the art, values that are expressed as ranges can assume any specific value or subrange within the stated ranges in different embodiments of the invention, to the tenth of the unit of the lower limit of the range, unless the context clearly dictates otherwise.

In addition, it is to be understood that any particular embodiment of the present invention that falls within the prior art may be explicitly excluded from any one or more of the claims. Since such embodiments are deemed to be known to one of ordinary skill in the art, they may be 10 excluded even if the exclusion is not set forth explicitly herein. Any particular embodiment of the compositions of the invention (e.g., any nucleic acid or protein encoded thereby; any method of production; any method of use; etc.) can be excluded from any one or more claims, for any 15 reason, whether or not related to the existence of prior art.

All cited sources, for example, references, publications, databases, database entries, and art cited herein, are incorporated into this application by reference, even if not expressly stated in the citation. In case of conflicting statements of a cited source and the instant application, the statement in the instant application shall control.

EXAMPLES

Modified mRNAs (mmRNAs) according to the invention can be made using standard laboratory methods and materials. The open reading frame (ORF) of the gene of interest is flanked by a 5' untranslated region (UTR) containing a strong Kozak translational initiation signal and a 3' UTR 30 (e.g., an alpha-globin 3' UTR) terminating with an oligo(dT) sequence for templated addition of a polyA tail. The mmR-NAs can be modified with pseudouridine (ψ) and 5-methylcytidine (5 meC) to reduce the cellular innate immune response. Kariko K et al. Immunity 23:165-75 (2005), 35 Kariko K et al. Mol Ther 16:1833-40 (2008), Anderson B R et al. NAR (2010).

The cloning, gene synthesis and vector sequencing can be performed by DNA2.0 Inc. (Menlo Park, Calif.). The ORFs can be restriction digested and used for cDNA synthesis 40 using tailed-PCR. This tailed-PCR cDNA product can be used as the template for the modified mRNA synthesis reaction using 25 mM each modified nucleotide mix (modified U/C was manufactured by TriLink Biotech, San Diego, Calif., unmodifed A/G was purchased from Epicenter Bio- 45 technologies, Madison, Wis.) and CellScript MegaScript™ (Epicenter Biotechnologies, Madison, Wis.) complete mRNA synthesis kit. The in vitro transcription reaction can be run for 3-4 hours at 37° C. PCR reaction can use HiFi PCR 2X Master MixTM (Kapa Biosystems, Woburn, Mass.). 50 The in vitro transcribed mRNA product can be run on an agarose gel and visualized. mRNA can be purified with Ambion/Applied Biosystems (Austin, Tex.) MEGAClear RNA™ purification kit. PCR reaction can be purified using PureLinkTM PCR purification kit (Invitrogen, Carlsbad, 55 Calif.) or PCR cleanup kit (Qiagen, Valencia, Calif.). The product can be quantified on NanodropTM UV Absorbance (ThermoFisher, Waltham, Mass.). Quality, UV absorbance quality and visualization of the product can be performed on a 1.2% agarose gel. The product can be resuspended in TE 60

When transfected into mammalian cells, the modified mRNAs may have a stability of between 12-18 hours.

For animal experiments, the IV delivery solution can be 150 mM NaCl, 2 mM CaCl2, 2 mM Na+-phosphate, and 0.5 65 mM EDTA, pH 6.5 and 10 µl lipofectamine (RNAiMaxTM, Invitrogen, Carlsbad, Calif.).

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Example 1

Use of Synthetic Modified mRNAs to Produce Functional Anti-Microbial Peptides and Proteins by Human Cells

The goal of this example is to express several functional AMPs from modified RNA in several human cell lines to test antibacterial effect of AMPs with distinct patterns of natural distribution and activities.

Each AMP (hBD-2, LL-37, or RNAse-7) is cloned into propagation plasmid in connection with sequences required for efficient translation and prolonged life of mRNA in cell with globin 5' and 3' UTRs and polyA tail. The mRNAs containing modified nucleotides and/or backbone modifications are transcribed using a standard T7 RNA polymerasedependent transcription system from plasmid templates. Those mRNAs are transfected into a panel of primary human cell lines including keratinocytes and fibroblasts using a lipophilic carrier. The intensive optimization of expression is performed in matrix-type experiments focusing on dose, media and delivery reagents selection. Then a dose titration curve of AMP expression can be established in a repeat administration protocol. As a positive transfection control, each construct encodes the EGFP gene for visualization. The expressed and secreted polypeptides are detected by corresponded antibodies by ELISA and Western blots. The specific antimicrobial activity is tested in corresponded microbiological plate assays or antibacterial neutralization assays required for the selection of targeted microorganisms. The strain collection can be tested for sensitivity to AMPs by determining their minimal inhibitory concentration (MIC) using those methods. Apoptosis is monitored using FACS with Annexin VCy5.5 and DAPI staining Apoptotic DNA fragmentation can also be observed by agarose gel electrophoresis. Interferon production is assayed from the cell supernatant using standard ELISA techniques and qPCR of inflammatory gene products. Experiments can be carried out with a collection of different microorganisms including Listeria monocytogenes strains and Staphylococcus aureus strains representing different lineages and serotypes (L. monocytogenes), spa types (S. aureus), and origins (food processing environment, food products, and human clinical isolates).

Example 2

The Combinatory Effect of Modified mRNAs for Polypeptides with Different Antimicrobial Mechanisms on Bacterial Resistance

The goal of this example is to show increase in antibacterial potency of AMP by co-expression of combination of several functional AMPs with distinct patterns of natural distribution and activities in human cell lines and test antibacterial effect of combination of AMPs on microorganisms partially resistant against one of AMP. AMPs can interact with the membrane lipids and form a channel through which ions can escape, upsetting homeostasis and eventually leading to cell lysis. However, other mechanisms for AMP activity may include activation of autolysis as well as nonlytic mechanisms such as inhibition of protein synthesis, degradation of proteins required for DNA replication, and interference with the transport and energy metabolism 7(Gob). The approach described in this example is to reduce bacterial resistance and cover a wider variety of pathogenic microorganisms by applying the desired mixture of two and

more in vitro-generated, modified synthetic mRNAs encoded AMPs (e.g., hBD-2, LL-37, RNAse-7). The library of AMPs with most studied and different mechanisms of action can be cloned and transcribed as above. Following by the developed optimal protocol for modified mRNA transfection, many possible combinations of target AMPs can be expressed in a panel of human cell lines including keratinocytes and fibroblasts using a lipophilic carrier in described anti-bacterial assays in a systematic manner looking for the lowest possible dose for bacteriostatic effect on selected panel of microorganisms.

Example 3

The Combinatory Effect of Modified mRNA for AMP and Conventional Antibiotic on Bacterial Resistance

The goal of this example is to show increase in antibacterial potency of AMP distinct patterns of natural distribution and activities expressed in human cell lines from modified RNA by combination with one or more traditional antibiotic drugs test antibacterial effect of combination of AMPs on microorganisms partially resistant against those 25 antibiotic drugs.

Human peptide antibiotics, in combination with wide variety of other natural polypeptides from other species and conventional antibiotics can be used as therapeutic agents, avoiding the problems of acquired resistance. The approach 30 described in this example is to reduce bacterial resistance and cover a wider variety of pathogenic microorganisms by applying the desired mixture of one or more in vitrogenerated, modified synthetic mRNAs encoded AMPs and one or more traditional antibiotics and to show synergetic 35 effect of combination of panel of traditional antibiotics and panel of AMPs. For example, the hBD-2, LL-37, and RNAse-7 can be used along and in all possible combinations. The following exemplary antibiotics can be used in this example: penicillins such as penicillin and amoxicillin; 40 cephalosporins such as cephalexin (Keflex); macrolides such as erythromycin (E-Mycin), clarithromycin (Biaxin), and azithromycin (Zithromax); fluoroquinolones such as ciprofloxacin (Cipro), levofloxacin (Levaquin), and ofloxacin (Floxin); sulfonamides such as co-trimoxazole (Bactrim) 45 and trimethoprim (Proloprim); tetracyclines such as tetracycline (Sumycin, Panmycin) and doxycycline (Vibramycin); and aminoglycosides such as gentamicin (Garamycin) and tobramycin (Tobrex). The library of AMPs with most studied and different mechanisms of action can be cloned 50 and transcribed as above. Following by the developed optimal protocol for modified mRNA transfection, many possible combinations of target AMPs can be expressed in a panel of human cell lines including keratinocytes and fibroblasts using a lipophilic carrier in described antibacterial 55 assays in a systematic manner looking for the lowest possible dose for bacteriostatic effect on selected panel of microorganisms including microorganisms known to be resistant to one or more traditional antibiotics.

Example 4

The Modified mRNA Technology as a Tool for Developing Novel Antibiotic Activity

The goal of this example is to develop efficient protocol for discovery, validation and development of new AMPs.

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The AMP validation protocol in high throughput manner can be developed. There have been many new AMPs recently discovered, but their mechanisms of action and utility for therapeutic applications remain unknown. Modified RNA technology allows for the simultaneous testing of new AMPs for human cell toxicity and antimicrobial activities. The sequence of newly discovered candidates can be cloned for in vitro RNA synthesis and testing in high throughput screens without actual peptide expression. Following by the optimal protocol for modified mRNA transfection, several new AMPs expressed in human cells against a panel of microorganisms can be tested. The AMP improvement protocol can be developed. 2-3 known AMPs are selected and a systematic walkthrough mutagenesis by PCR ¹⁵ and clone resulting constructs in plasmid vectors are performed. The library of those mutants can be tested one-byone in a high throughput screen according to developed protocols in comparison to wild type peptides. Functional domains in testing proteins and peptides associated with human cytotoxicity and domains linked to certain mechanisms of antimicrobial activities can be identified. The results of those scanning efforts can allow engineering AMPs with optimal non-toxic but rapid bacteriostatic activi-

Example 5

The Effect of Synthetic Modified mRNAs Coding Intracellular Communication Factors on the Expression of AMPs in Human Cells

The goal of this example is to use modified mRNAs coding intracellular communication factors to induce innate immune system including expression of AMPs.

The expression of AMP genes in a variety of epithelial cells can be enhanced using specific nutrients, vitamins (D) and other short chain fatty acids as therapeutic treatment. The opportunity for more specific signal for expression of AMP can be investigated. hBD-2 messenger RNA expression in foreskin-derived keratinocytes was greatly up-regulated with TNF-α within 1 h of stimulation and persisted for more than 48 h. The TNF- α gene can be used for synthesis of modified mRNA and transfected into a panel of primary human cell lines including keratinocytes and fibroblasts using a lipophilic carrier. It can be used to test expression of several AMPs including hBD-2 in human cells. The expressed TNF-α and secreted AMPs can be detected by corresponded antibodies by ELISA and Western blots. The specific anti-microbial activity can be tested in corresponded microbiological plate assays or anti-bacterial neutralization assays required for the selection of targeted microorganisms. Apoptosis can be monitored using FACS with Annexin VCy5.5 and DAPI staining Apoptotic DNA fragmentation can also be observed by agarose gel electrophoresis. Interferon production can be assayed from the cell supernatant using standard ELISA techniques and qPCR of inflammatory gene products.

Example 6

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Use of Synthetic Modified mRNAs to Produce Functional Antimicrobial Peptides and Proteins by Animal Cells for Development of Antibiotics for Agriculture Industry

The goal of this example is to express several functional AMPs from modified RNA in several animal cell lines to test

anti-bacterial effect of AMPs with distinct patterns of natural distribution and activities to test possibility to use modified RNAs as antibiotics in agriculture.

Each AMP (hBD-2, LL-37, and RNAse-7) can be cloned into propagation plasmid in connection with sequences required for efficient translation and prolonged life of mRNA in cell with globin 5' and 3' UTRs and polyAtail. The mRNAs containing modified nucleotides and/or backbone modifications can be transcribed using a standard T7 RNA polymerase-dependent transcription system from plasmid templates. Those mRNAs are transfected into a panel of primary human cell lines including keratinocytes and fibroblasts using a lipophilic carrier. The intensive optimization of expression can be performed in matrix-type experiments focusing on dose, media and delivery reagents selection. A dose titration curve of AMP expression can be established in a repeat administration protocol. As a positive transfection control, each construct encodes the EGFP gene for visualization. The expressed and secreted polypeptides can be detected by corresponded antibodies by ELISA and Western blots. The specific antimicrobial activity can be tested in corresponded microbiological plate assays or antibacterial neutralization assays required for the selection of targeted microorganisms. Apoptosis is monitored using FACS with Annexin VCy5.5 and DAPI staining Apoptotic DNA fragmentation can also be observed by agarose gel electrophoresis. Interferon production can be assayed from the cell supernatant using standard ELISA techniques and qPCR of inflammatory gene products.

Example 7

In Vitro Selection of Anti-Viral Inhibitory Peptides Encoded by Synthetic Modified mRNA

The viral lifecycle may be inhibited by antibody mimetic anti-viral peptides at a number of points. Viral entry into the host cell can be prevented by inhibitory peptides that ameliorate the proper folding of the viral hairpin fusion complex. Alternatively, intracellular viral propagation may be inhibited by antiviral peptides directed against viral capsid assembly thereby preventing the formation of functional infectious viral particles. The goal of this example is to identify anti-viral peptides using mRNA-display technology directed

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against specific viral capsid proteins or viral envelope proteins from HIV, herpes or influenza viruses. The mRNA display in vitro selection can be performed similar to previously described methods (Wilson et al., PNAS USA, 2001, 98(7):375). Briefly, a synthetic oligonucleotide library is constructed containing ~10¹³ unique sequences in a 30-nt randomized region for selection of a 10aa antiviral peptide. The oligonucleotide library is synthesized containing a 3'-puromycin nucleotide analog used to covently attach the nascent peptide chain to its encoded mRNA during the in vitro translation step in rabbit reticulocyte lysate. A preselection round can filter the mRNA peptide-display library over a ligand-free column to remove non-specific binding partners from the pool. The selection rounds can then proceed through passage and incubation over a target viralprotein functionalized selection column followed by a wash through selection buffer (20 mM Tris-HCl, pH7.5; 100 mM NaCl). The bound peptides are eluted with an alkaline elution buffer (0.1M KOH) and the sequence information in the peptide is recovered through RT-PCR of the attached mRNA. Mutagenic PCR may be performed between selection rounds to further optimized binding affinity and peptide stability. Based on previous mRNA-display selections (Wilson et al., PNAS USA, 2001, 98(7):375), this selection is expected to recover high affinity (K ~50 pM-50 nM) antiviral peptides after 15-20 rounds of selection. To test in vivo functionality of the anti-viral peptide, synthetic modified mRNAs encoding the anti-viral peptide are transfected into target cells. Post-transfection culture transduction with infectious virus or mock-virus are performed and viral propagation can be monitored through standard pfu counts and qPCR of viral genomic material. Cells transfected with synthetic mRNAs encoding the appropriate anti-viral peptide inhibitor are expected to reduce viral propagation, display reduced pfu counts, reduced viral RNA or DNA in culture, and increase cell survival. In vivo efficacy, PK and toxicology can be studied in appropriate animal models.

Lengthy Table

The patent application contains a lengthy table section. A copy of the table is available in electronic form from the USPTO web site. An electronic copy of the table will also be available from the USPTO upon request and payment of the fee set forth in 37 CFR 1.19(b)(3).

SEQUENCE LISTING

The patent contains a lengthy "Sequence Listing" section. A copy of the "Sequence Listing" is available in electronic form from the USPTO web site (http://seqdata.uspto.gov/?pageRequest=docDetail&DocID=US09464124B2). An electronic copy of the "Sequence Listing" will also be available from the USPTO upon request and payment of the fee set forth in 37 CFR 1.19(b)(3).

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What is claimed is:

- 1. A pharmaceutical composition comprising:
- a lipid-based nanoparticle comprising a synthetic messenger ribonucleic acid (mRNA) encoding a defensin polypeptide in an amount effective to permit production of the defensin polypeptide in a cell, wherein the 65 synthetic mRNA comprises a translatable region that contains at least one nucleoside modification, and
- wherein 75-100% of uridine nucleotides in the synthetic mRNA are modified.
- 2. The pharmaceutical composition of claim 1, wherein the at least one nucleoside modification is selected from the group consisting of pyridin-4-one ribonucleoside, 5-azauridine, 2-thio-5-aza-uridine, 2-thio-pseudouridine, 4-thio-pseudouridine, 2-thio-pseudouridine, 5-hydroxyuridine, 3-methyluridine, 5-carboxymethyl-uridine, 1-carboxymethyl-pseudouridine, 5-propynyl-uridine, 1-propynyl-

pseudouridine, 5-taurinomethyluridine, 1-taurinomethylpseudouridine. 5-taurinomethyl-2-thio-uridine, 1-taurinomethyl-4-thio-uridine, 5-methyl-uridine, 1-methylpseudouridine, 4-thio-1-methyl-pseudouridine, 2-thio-1methyl-pseudouridine, 1-methyl-1-deaza-pseudouridine, 2-thio-1-methyl-1-deaza-pseudouridine. dihydrouridine. dihydropseudouridine, 2-thio-dihydrouridine, 2-thio-dihydropseudouridine, 2-methoxyuridine, 2-methoxy-4-thio-uri-4-methoxy-pseudouridine, 4-methoxy-2-thiopseudouridine, 5-aza-cytidine, pseudoisocytidine, 3-methyl-N4-acetylcytidine, 5-formylcytidine, N4-methylcytidine, 5-hydroxymethylcytidine, 1-methylpseudoisocytidine, pyrrolo-cytidine, pyrrolo-pseudoisocytidine, 2-thio-cytidine, 2-thio-5-methyl-cytidine, 4-thiopseudoisocytidine, 4-thio-1-methyl-pseudoisocytidine, 4-thio-1-methyl-1-deaza-pseudoisocytidine, 1-methyl-1deaza-pseudoisocytidine, zebularine, 5-aza-zebularine, 5-methyl-zebularine, 5-aza-2-thio-zebularine, 2-thio-zebularine, 2-methoxy-cytidine, 2-methoxy-5-methyl-cytidine, 20 4-methoxy-pseudoisocytidine, 4-methoxy-1-methylpseudoisocytidine, 2-aminopurine, 2, 6-diaminopurine, 7-deaza-adenine, 7-deaza-8-aza-adenine, 7-deaza-2-aminopurine, 7-deaza-8-aza-2-aminopurine, 7-deaza-2,6-diamin-7-deaza-8-aza-2,6-diaminopurine, 1-methyladenosine, N6-methyladenosine, N6-isopentenyladenosine, N6-(cis-hydroxyisopentenyl)adenosine, 2-methylthio-N6-(cis-hydroxyisopentenyl) adenosine, N6-glycinylcarbamoyladenosine, N6-threonylcarbamoyladenosine, 2-methylthio-N6,N6- 30 N6-threonyl carbamoyladenosine, 7-methyladenine, dimethyladenosine, 2-methylthioadenine, 2-methoxy-adenine, inosine, 1-methyl-inosine, wyosine, wybutosine, 7-deaza-guanosine, 7-deaza-8-azaguanosine, 6-thio-guanosine, 6-thio-7-deaza-guanosine, 6-thio-7-deaza-8-aza-guanosine, 7-methyl-guanosine, 35 6-thio-7-methyl-guanosine, 7-methylinosine, 6-methoxyguanosine. 1-methylguanosine, N2-methylguanosine, N2,N2-dimethylguanosine, 8-oxo-guanosine, 7-methyl-8oxo-guanosine, 1-methyl-6-thio-guanosine, N2-methyl-6thio-guanosine, and N2,N2-dimethyl-6-thio-guanosine.

- 3. The pharmaceutical composition of claim 1, wherein the composition is formulated for administration via a route selected from the group consisting of: systemic, local, intravenous, topical, oral, administration via a dressing, administration via a bandage, rectal, vaginal, intramuscular, transarterial, intraperitoneal, intranasal, subcutaneous, endoscopic, transdermal and intrathecal.
- **4**. The pharmaceutical composition of claim **3**, wherein the route is intravenous.
- 5. The pharmaceutical composition of claim 1, wherein the defensin polypeptide is selected from the group consisting of α -defensins, and β -defensins and θ -defensins.

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- **6**. The pharmaceutical composition of claim **1**, wherein the anti-microbial polypeptide is hBD-2 (SEQ ID NO: 191 or 192).
 - 7. A kit comprising:
 - (a) a synthetic messenger ribonucleic acid (mRNA) encoding a defensin polypeptide packaged in a container, wherein the synthetic mRNA comprises a translatable region that contains at least one nucleoside modification, and wherein 75-100% of uridine nucleotides in the synthetic mRNA are modified; and
 - (b) a pharmaceutically acceptable carrier packaged in a container.
- 8. The pharmaceutical composition of claim 5, wherein the defensin is an α -defensin selected from the group consisting of neutrophil defensin 1, defensin alpha 1, neutrophil defensin 3, neutrophil defensin 4, defensin 5 and defensin 6.
- 9. The pharmaceutical composition of claim 5, wherein the defensin is a β -defensin selected from the group consisting of beta-defensin 1, beta-defensin 2, beta-defensin 103, beta-defensin 110 and beta-defensin 136
- 10. The pharmaceutical composition of claim 5, wherein the at least one nucleoside modification is pseudouridine.
- 11. The pharmaceutical composition of claim 5, wherein 100% of uridine nucleotides in the synthetic mRNA are modified.
- 12. The pharmaceutical composition of claim 5, wherein the synthetic mRNA further comprises a 5' untranslated region that contains at least one nucleoside modification.
- 13. The pharmaceutical composition of claim 5, wherein the synthetic mRNA further comprises a 5' untranslated region that contains at least two nucleoside modifications.
- **14.** The pharmaceutical composition of claim **5**, wherein the synthetic mRNA further comprises a 3' untranslated region that contains at least one nucleoside modification.
- 15. The pharmaceutical composition of claim 5, wherein the synthetic mRNA further comprises a 3' untranslated region that contains at least two nucleoside modifications.
- 16. A lipid-based nanoparticle comprising a synthetic messenger ribonucleic acid (mRNA) encoding a defensin polypeptide in an amount effective to permit production of the defensin polypeptide in a cell,
 - wherein 75-100% of uridine nucleotides in the synthetic mRNA are modified, and
 - wherein the synthetic mRNA comprises a translatable region, a 5' untranslated region and a 3' untranslated region, each of which contains at least one nucleoside modification.
- 17. The lipid-based nanoparticle of claim 16, wherein the at least one nucleoside modification is pseudouridine.

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